

Max-Planck-Institut für molekulare Physiologie



Novel Transition Metal-Free C-H Bond Functionalization Methods for Biologically Important Heterocycles Synthesis

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Abstract

The development of transition metal-free reaction methodologies for the direct functionalization of C–H bonds is an attractive area of research. These reactions offer atom-economic methods for the synthesis and functionalization of valuable organic molecules in the absence of precious metal catalysts under mild conditions. Novel methodologies have been established for biologically important nitrogen containing molecules (Figure 1). A regioselective cross-coupling method for the benzylation and alkynylation of privileged heterocyclic scaffolds like quinolines, isoquinolines and pyridines were evaluated under organocatalytic conditions. Oxidized *N*-heterocyclic compounds were reacted with organosilanes in presence of a fluoride catalyst. This method offers a highly selective route to access C₁-benzylated isoquinolines, C₂-benzylated quinolines and pyridines. Additionally, a unique sigma bond metathesis strategy is explored to obtain symmetrically disubstituted acetylenes containing privileged scaffolds (Chapter 3).

Electroorganic chemistry has witnessed a renewed interest in recent years because of its profound advantages over other methods such as oxidants mediated oxidative coupling reactions. Many oxidants used in the organic reactions are potentially hazardous and even toxic. The risk of handling such oxidants in stoichiometric amounts is high. Alternatively, electricity itself can be employed as sole oxidant to carry out many redox reactions. Combining the field of transition metal-free C–H functionalization chemistry with the revived electroorganic chemistry offers unique advantages. Inspired by this proposal, an electrochemical dehydrogenative amination including both intramolecular and intermolecular variants have been established. Detailed mechanism involving a nitrenium ion intermediate has been proposed revealing the possible generation of nitrenium ion intermediates under electrochemical oxidative conditions for the first time (Chapter 4).

Direct C-H bond functionalization methods frequently give access to the kind of molecules which were never accessed before. These molecules however might be relevant in terms of biological properties. Therefore, the evaluation of bioactivities for novel molecules synthesized by direct C-H bond functionalization methods are highly significant. A novel physiological, morphogenic cellular screening system that is focused on the bone morphogenetic pathway revealed chromones as potential BMP activator chemotypes. The key step in the synthesis of these chromone compounds is an oxidative C-H amination in presence of molecular iodine and base. The active

hits were resynthesized in adequate amounts for *in vitro* and *in vivo* studies. Additionally, based on the molecular structure of active hits, novel derivatives were designed and synthesized. Afterwards, biological experiments were carried out in collaboration to understand the structure activity relationship of various chromones as BMP effectors (Chapter 5).

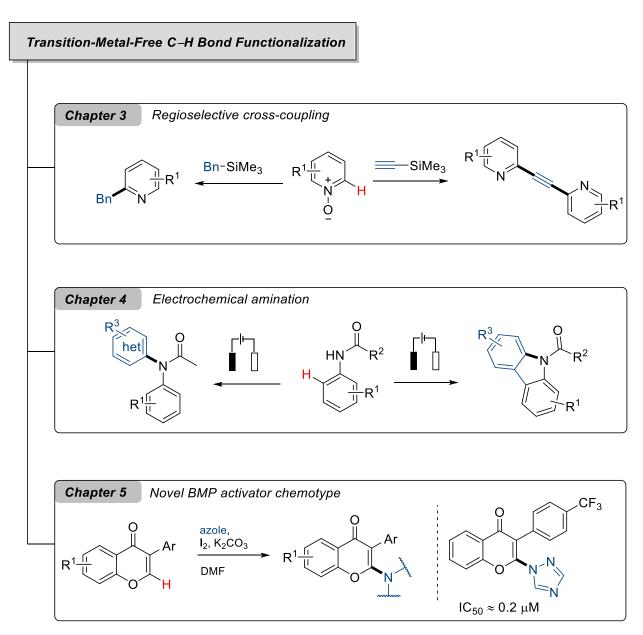


Figure 1 Overview of the projects described in this thesis

Zusammenfassung

Die Entwicklung übergangsmetallfreien Reaktionsmethoden für die direkte von Funktionalisierung von C-H-Bindungen ist ein attraktives Forschungsgebiet. Diese Reaktionen bieten verbesserte, atomökonomische Methoden für die Synthese und Funktionalisierung wertvoller organischer Moleküle in Abwesenheit von Edelmetallkatalysatoren unter relativ milden Bedingungen. In diesem Zusammenhang wurden neue Methoden für biologisch relevante stickstoffhaltige Moleküle entwickelt (Abbildung 1). Zunächst wurde eine regioselektive Kreuzkupplungsmethode für die Benzylierung und Alkynylierung privilegierter heterozyklischer Strukturgerüste wie Chinoline, Isochinoline und Pyridine unter organokatalytischen Bedingungen untersucht. Voroxidierte N-heterozyklische Verbindungen wurden mit Organosilanen in Anwesenheit eines Fluoridaktivators umgesetzt. Diese Methode bietet einen hochselektiven Zugang zu C₁-benzylierten Isochinolinen, C₂-benzylierten Chinolinen und Pyridinen. Zusätzlich wird eine einzigartige Sigma-Bindungs-Metathesestrategie untersucht, um symmetrisch disubstituierte Acetylene zu erhalten, die an beiden Enden privilegierte Gerüste aufweisen. Diese Verbindungen wurden auch auf potenzielle Bioaktivität untersucht (Kapitel 3). Die elektroorganische Chemie hat in den letzten Jahren aufgrund ihrer tiefgreifenden Vorteile gegenüber anderen Methoden, wie z. B. durch chemische Oxidationsmittel vermittelte oxidative Kupplungsreaktionen, ein neues Interesse geweckt. Viele verwendeten Oxidationsmittel in organischen Reaktionen sind potenziell gefährlich und sogar giftig. Der Umgang mit solchen Oxidationsmitteln in stöchiometrischen Mengen ist mit hohem Risiko verbunden. Alternativ kann Elektrizität selbst als gezieltes Oxidationsmittel eingesetzt werden, um viele Redoxreaktionen durchzuführen. Die Kombination von übergangsmetallfreien C-H-Funktionalisierungschemie mit der wiederbelebten elektroorganischen Chemie bietet einzigartige Vorteile. Inspiriert von diesem Ansatz wurde eine elektrochemische dehydrierende Aminierung entwickelt, die sowohl intramolekulare als auch intermolekulare Varianten umfasst. Ein detaillierter Mechanismus, an dem ein Nitrenium-Ionen-Zwischenprodukt beteiligt ist, welcher zum ersten Mal die mögliche Bildung von Nitrenium-Ionen Zwischenprodukten unter elektrochemischen oxidativen Bedingungen aufzeigt, wurde vorgeschlagen (Kapitel 4).

Direkte C-H-Bindungsfunktionalisierungsmethoden ermöglichen häufig den Zugang zu Molekülen, die bisher nicht bzw. erschwert zugänglich waren. Diese Moleküle könnten jedoch im

Hinblick auf biologische Eigenschaften von hoher Bedeutung sein. Daher ist die Untersuchung der Bioaktivität Moleküle, die mit Hilfe direkter neuartiger Bindungsfunktionalisierungsmethoden synthetisiert wurden, von großer Bedeutung. Ein neuartiges physiologisches, morphogenetisches zelluläres Screening-System, das sich auf den Knochenmorphogenese-Weg (BMP-Weg) konzentriert, hat zum Beispiel Chromone als potenziellen BMP-Aktivator-Chemotyp enthüllt. Der Schlüsselschritt bei der Synthese dieser Chromonverbindungen ist eine oxidative CH-Aminierung in Gegenwart von molekularem Jod und einer Base. Die aktiven Verbindungen wurden in ausreichenden Mengen für In-vitro- und In-vivo-Studien resynthetisiert. Darüber hinaus wurden auf der Grundlage der Molekularstruktur der aktiven Moleküle neue Derivate entworfen und synthetisiert. Anschließend wurden in Zusammenarbeit biologische Experimente durchgeführt, um die Struktur-Aktivitäts-Beziehung der verschiedenen Chromone als BMP-Effektoren zu analysieren.

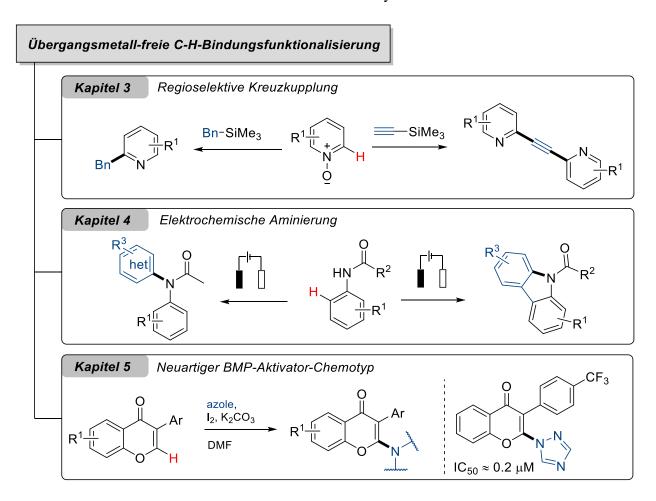
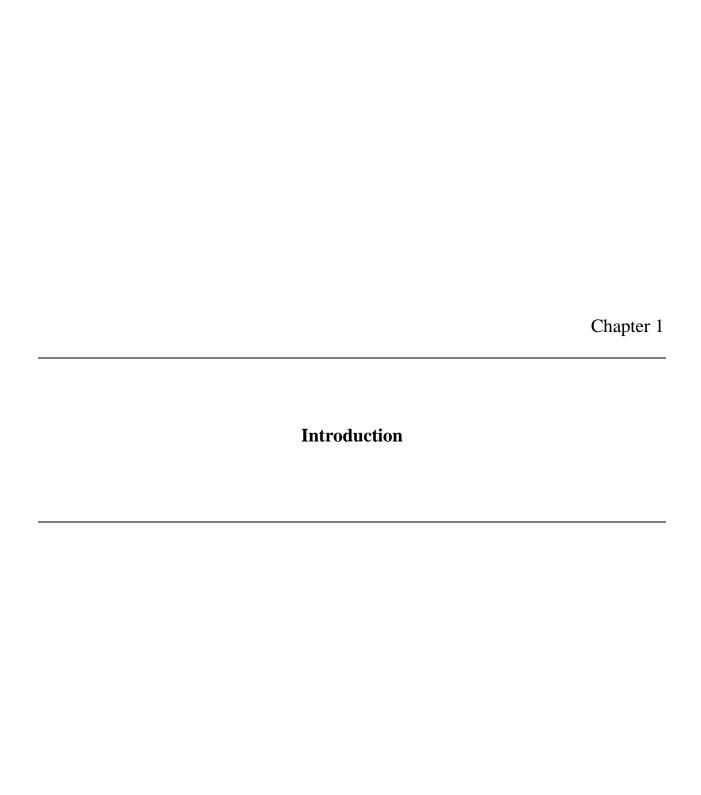


Abbildung 1 Überblick über die in dieser Arbeit beschriebenen Projekte

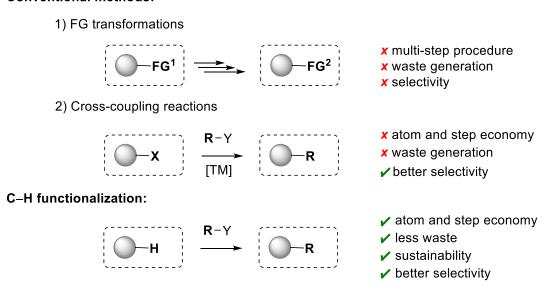


1. Introduction

1.1 C–H functionalization

Direct C–H bond functionalization methods have attracted the attention of the synthetic community because of their undisputed advantages over the traditional methods. Since the 1960s, following the discovery of Shilov's platinum chemistry, this research area has experienced tremendous developments. Considering their new-found applications in synthetic organic chemistry and medicinal chemistry such as late-stage functionalization, researchers around the globe continue to develop novel reactions as well as solve existing challenges in this field. C–H functionalization is used to directly functionalize otherwise unreactive C–H bonds without the requirement of their pre-functionalization. This offers a number of advantages over traditional methods of organic synthesis (Scheme 1.1). Development of atom and step economical reactions were realized with minimal amounts of waste generation in the form of reaction by-products. In addition, from the sustainability point of view, the catalytic C–H functionalization methods offer milder conditions, more efficient routes in comparison to cross-coupling reactions. Due to their large functional group tolerance and cost effectiveness up to an extent, these methodologies are highly significant for the synthesis and functionalization of biologically relevant natural products and pharmaceuticals.

Conventional methods:

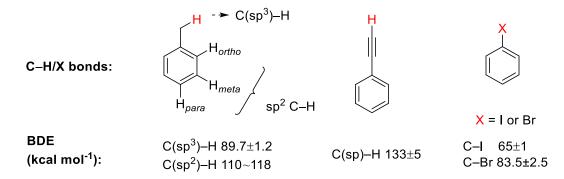


Scheme 1.1 Progress in organic synthesis towards C–H functionalization

Techniques like late-stage functionalization enable synthesis of large numbers of derivatives from the same intermediate *via* direct functionalization of carbon-hydrogen bonds^[4] and hence helping in the structure-activity relationship studies during drug discovery. In short, a method once considered as highly challenging and less practical is now becoming an important aspect of synthetic route design and offers a new set of retrosynthetic connections for molecularly diverse architectures (Scheme 1.2).^[5]

Scheme 1.2 Late-stage functionalization of a complex organic molecule

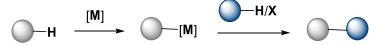
However, the challenges to overcome in the area of C–H activation/functionalization chemistry are still many in terms of industrial applications. The limitations pertaining to catalytic C–H functionalization methods are mainly attributed to the ubiquitous nature of carbon-hydrogen bonds, their relatively higher bond dissociation energies and lack of major electronic differences between various C–H bonds. These challenges ultimately result in harsh reaction conditions. Moreover, C–H functionalization methods advancing the sustainability of chemical reactions are also an active area of research "to meet the chemistry needs of the present without compromising those of future" (Scheme 1.3).^[6]



Scheme 1.3 Relative bond dissociation energies (BDEs) of different C–H/X bonds

The majority of developed C-H functionalization methodologies use various transition metal catalysts (TM) in order to facilitate the activation of inert and thermodynamically stable C-H bonds. In this regard, a large number of site-selective transformations have been realized employing external or internal directing group coordination chemistry.^[7] This not only includes selective functionalization of adjacent C–H bonds to the coordinating group, but also C–H bonds located at remote positions of coordinating functionalities.^[8] Currently, the development of enantioselective reactions by means of C-H activation has gained attention as well. [9] Despite the undisputed advantages over other existing methods C-H activation has its own shortcomings (Scheme 1.4). Most of these methods necessitate the use of precious transition metal catalysts. With a clear focus on sustainable chemistry for the future, it is obvious that dependence on such scarce metals should be minimized. Many such literature methods employ high catalyst loadings of high valent transition metals as well. In general, these methods also employ high temperatures and solvents which are not well suited for green and sustainable chemistry. Another problem is the dependence on hazardous oxidants for the completion of catalytic cycles. Also, additional directing group manipulations are required for selective insertion of functional groups. However, a large number of methods have already been published with improved sustainability parameters for C-H activation in the past two decades.^[10] These include methodologies using early transition metals, reduced catalyst loading and alternative oxidant systems are to name a few. Catalysis under mild and efficient reaction conditions in environmentally friendly solvents as well as innate functional group directed transformations are also gaining attention.^[11]

Key issues of C-H Activation:



- precious transition metals
 stoichiometric oxidants
 sensitive reactions
- directing groups metal impurities in final products

Scheme 1.4 Key challenges of sustainable C–H functionalization methods

1.1.1 Transition metal-free C–H functionalization

An appealing alternative could be the development of a greater number of C–H functionalization reactions in the absence of any transition metals.^[12] New avenues in this field of research like

organocatalytic and oxidative C–H functionalizations are progressing rapidly (Figure 1.1). As two significant areas of modern organic chemistry, C–H functionalization and transition metal-free (TM-free) catalysis results in the development of viable, green, selective and efficient transformations. In addition, such methods allow synthesis of important intermediates of pharmaceuticals without the high risk of contamination with heavy metal impurities. Otherwise removal of these impurities from the final products are a time consuming and cost-ineffective process.

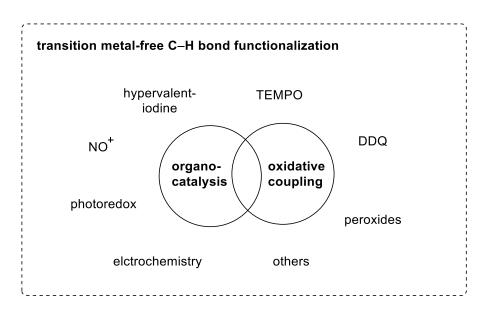


Figure 1.1 Methods of TM-free C–H bond functionalizations

Despite the advances made in the past couple of decades, more selective and efficient methods are in high demand at present. In this context, rapid advances in the area of organocatalysis along with other transition metal-free methodologies are well foreseeable. [13] According to existing literature reports, transition metal-free C–H functionalization reactions can be broadly categorized as organocatalytic methods and oxidative C–H functionalization reactions such as oxidative homoand hetero- couplings. The former class visibly involves a complete catalytic cycle while the latter could be oxidation-initiated C–C or C–X bond forming reactions. [14]

There are different classes of organocatalysts employed in inert C–H functionalization reactions.^[15] A renaissance in electroorganic synthesis and growing popularity of photoredox catalysis are also complimenting organocatalytic research well.^[16-17] Oxidative C–H bond

functionalization reactions are also gaining significant popularity owing to their ability to carry out efficient organic transformations without the requirement of pre-functionalization.

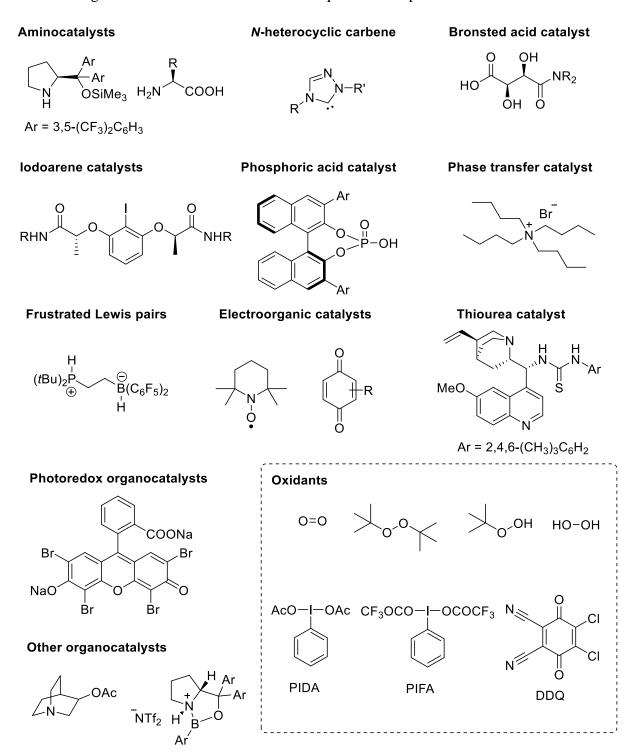


Figure 1.2 Representative organocatalysts and oxidants for C–H bond functionalization

The amounts of waste generated is significantly lower. However, to functionalize stable C–H bonds is a challenge. For this reason, the electron/hydrogen transfer reagents are employed to significantly increase the acidity of C–H bonds and to decrease the BDE in oxidative C–H functionalization as in the case of photoredox or electroorganic catalysis. Taking advantage of this chemistry, various chemicals like 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ), 2,2,6,6-tetramethylpiperidin-1-yl)oxyl (TEMPO), phenyl iodine (III) diacetate (PIDA), bis(trifluoroacetoxy)iodo)benzene (PIFA), nitrosonium salts, etc. are used for oxidative homocoupling and heterocoupling reactions in the absence of any transition metals (Figure 1.2).^[18]

Innate C–H functionalization reactions are based on their innate reactivity at certain positions of molecules as in the case of heteroaromatic *N*-oxide chemistry, aromatic substitution reactions, sulfoxides directed C–H couplings for example.^[19-23] In this regard, heterocyclic *N*-oxides are an important class of substrates in organic chemistry and they are well studied chemicals for direct C–H functionalization methods to obtain densely functionalized nitrogen heterocycles. They are also present in some biologically active natural products and synthetic drugs.^[24] A general route for the synthesis of C₂ substituted pyridine or quinoline compounds is the nucleophilic substitution of corresponding heteroaryl halides. However, these reactions usually require prefunctionalization, harsh conditions and still the yield of reactions are quite low in most of the cases (Scheme 1.5).

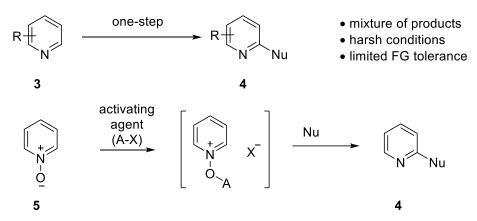
Scheme 1.5 Nucleophilic substitution of 2-halo-heteroarenes

Hartwig's group used a combination of fluorination and a following nucleophilic aromatic substitution for the site-selective late-stage functionalization of nitrogen containing aromatic

compounds. The reaction is broadly applicable for the α -functionalization of pyridine containing compounds (3) and various nucleophiles like cyano-, amino-, thio-, etc. could be employed to obtain corresponding C₂-substituted pyridines (4) (Scheme 1.5).^[25]

1.1.2 Transition metal free C–H functionalization of heteroaromatic N-oxides

Another strategy is to start with *N*-oxides in the presence of an activating agent, which will eventually increase the electrophilic nature of the 2-position. Hence, reactions will occur under relatively mild conditions. However, there are chances of undesired side reactions in presence of activating agents and 2- or 4- position selectivity is challenging to control in this type of *cine*-substitution reactions. (Scheme 1.6).^[26]



A-X = Ts_2O , TsCI, MsCI, RCOCI, PyBroP, $PhI(OAc)_2$, $POCI_3$, I_2 , etc.

Scheme 1.6 Nucleophilic substitution reactions of pyridine and heteroaromatic *N*-oxides

Many efficient deoxygenative functionalization reactions have been reported following this strategy. Among these, phosphorus containing activating groups are attractive reagents as the formation of strong phosphorus-oxygen bonds will drive the reaction to proceed. On this basis, Londregan and co-workers developed a facile amination strategy for the synthesis of 2-aminopyridines using commercially available bromo-tris-pyrrolidino-phosphonium hexafluorophosphate (PyBroP, 7) as activating agent. The reaction had a diverse substrate scope and could be used for the synthesis of various C₂-aminated pyridines and quinolines. The reaction was proposed to proceed through the formation of intermediate 5a. The nucleophilic attack of amine to the intermediate and following rearomatization resulted in the formation of selective amination products (8).^[27] In a following report by the same group, this method was further extended to

various carbon, sulfur and oxygen nucleophiles to obtain selectively C₂-substituted nitrogen heterocyclic compounds (Scheme 1.7).^[28-29]

Scheme 1.7 C–H functionalization of heteroaromatic *N*-oxides using activating agent

Baran group developed a mild and efficient approach for the regioselective bromination of azine N-oxides using tosic anhydride as activator. The methodology was successful for the bromination of diverse quinolines and isoquinolines. However, the reaction scope was not extendable to pyridine N-oxides. The authors also presented a one-pot oxidation-bromination strategy as well as a chlorination reaction using tetrabutylammonium chloride to demonstrate the versatility of the developed method (Scheme 1.8). [30]

Scheme 1.8 Regioselective C–H bromination of heteroaromatic *N*-oxides

In case of fluorination, a direct approach was not feasible because of the low nucleophilic reactivity of fluoride ions. Hence, the pyridine *N*-oxides were first converted into 2-pyridylalkylammonium salts (**10**). These salts are isolable and treating them with nucleophilic fluorine reagents like tetrabutylammonium fluoride or potassium fluorides give access to 2-fluoropyridine compounds. Interestingly, this method can be used in the synthesis of flortaucipir (¹⁸F) employing K¹⁸F in the presence of Kryptofix 2.2.2. Flortaucipir (¹⁸F) (**11**) is a radioactive diagnostic agent used for brain imaging with the help of positron emission tomography (PET) (Scheme 1.9). [31]

Scheme 1.9 Application of regioselective C–H fluorination in synthesis

Scheme 1.10 Three component reaction of *N*-oxides, acyl chlorides and cyclic (thio)ethers

Tomkinson's group revealed a strategy to introduce complex substituents at C₂-position of electron deficient pyridines through novel three component reactions. The reaction proceeds in presence of acyl chlorides as an activating agent and triethylamine as base. The product formation while using cyclic ethers such as tetrahydrofuran as nucleophile was different from those while using cyclic sulfides (12, 13). The reason was proposed to be the nucleophilic attack of chloride ion to the reaction intermediate (12a) instead of the acyloxy anion (13a) in the latter case (Scheme 1.10).^[32-33]

A highly regioselective direct C–H trifluoromethylation of pyridine, quinoline, isoquinoline and two or three nitrogen containing aromatic *N*-oxides was discovered by Kuninobu and Kanai *et al.* trifluoromethyldifluoroborane (**14**) was found to be an effective activating reagent to drive reaction under mild conditions even for gram-scale reactions with an impressive scope. The methodology possesses high potential considering the abundance of trifluoromethyl group in biologically active molecules (Scheme 1.11). [34]

Scheme 1.11 Direct C–H trifluoromethylation of *N*-oxides using trifluoromethyldifluoroborane

External activating group strategies for the site-selective C–H functionalization of *N*-oxides have been explored to access diversely functionalized nitrogen heterocycles. Moreover, the formation of new C–C, C–S, C–O, etc. (19-21) have been achieved *via* the direct functionalization of C–H bonds. Even though the transition metals are employed in many reported reactions, several metal-free alternatives are also available for a wide range of transformations in this class of reactions. In addition to common external activation reagents such as PyBroP, PPh₃, anhydrides, *H*-phosphonates, hypervalent iodine reagents and acid chlorides (16-18) have also been employed successfully to achieve useful functionalizations (Scheme 1.12).^[35-37]

On the other hand, reactions have also been developed where the use of external activating agents have been negotiated by employing reagents which can simultaneously act as nucleophile source

and activating agent. For instance, dialkyl phosphonates are common activating reagents for nucleophilic *cine*- substitution reactions of *N*-oxides to introduce a phosphonate group at α -position (Scheme 1.13).^[38-39]

$$R^{1} \stackrel{\downarrow}{ \downarrow} \stackrel{\downarrow}$$

Scheme 1.12 Other external activation reagents for C–H functionalization of *N*-oxides

Scheme 1.13 *cine*- substitution reactions of *N*-oxides with dialkyl phosphonates

Keith *et al.* revealed a method to access α -imidazole substituted *N*-heteroarenes in presence of commercially available sulfuryl diimidazole (23). The reaction showed high site selectivity. [40] Later, it was also shown that *N*-tosylated triazoles and diazoles (25) can also be used as *N*-heterocycle source in presence of base. Similarly, heating *N*-aryltriflimides (27) with heteroaromatic *N*-oxides in presence of diisopropylethylamine yielded *N*-aryltriflamidoazines. However, because of the harsh reaction conditions and low functional group tolerance, the applications of this methodology are limited. (Scheme 1.14). [41-42]

Scheme 1.14 Amination reactions of *N*-oxides in the absence of external activation

In many cases, a strong base can initiate the reaction by abstracting a proton from α -position of azine-N-oxides. Following nucleophilic attack of the dehydrogenative coupling partner to the resultant carbanion yields the biheteroaryl compounds (**30**). This methodology provides easy and efficient access to introduce heteroaromatic compounds to the *ortho* position of N-oxides (Scheme 1.15 and Scheme 1.16). [43]

Scheme 1.15 Base mediated *ortho* C–H functionalization of azine *N*-oxides

Scheme 1.16 Base mediated homocoupling of azine *N*-oxides

Scheme 1.17 Regioselective C₂-alkylation of diazine *N*-Oxides using Wittig reagents

In another approach an active nucleophilic species is generated *in situ* in presence of a base. This further attacks the *ortho*- position of heterocyclic *N*-oxide and ultimately leads to desired product formation.^[44] Taking inspiration from Cho's alkylation approach employing diborylalkanes as alkylating agents, Kim, Han and co-workers reported a metal-free deoxygenative alkylation

strategy for pyridine, quinoline N-oxides and diazene N-oxides. The method demonstrates the potential of Wittig reagents (33) in site-selective metal-free C-H bond functionalization. Wittig reagent reacts with the heteroaromatic N-oxides to generate intermediate 33b, from which the proton abstraction by the base yield the desired products. Various C_2 -alkylated diazine compounds (34a-f) were accessible via this method (Scheme 1.17). [45-47]

Heteroaromatic *N*-oxides are also 1,3-dipoles. This property has been exploited to develop novel methods of C–H functionalization reactions *via* 1,3-dipolar cycloadditions. In many cases, the intermediate cycloadduct undergoes further modifications to form more stable final products. Methodologies for the C₂-alkenylation of *N*-oxide containing heterocycles using activated olefins are available in literature. However, unactivated olefins (35) have also been successfully used for alkenylation reactions under Brønsted acid catalysis (Scheme 1.18). [50]

Scheme 1.18. 1,3-dipolar cycloaddition for alkenylation of *N*-oxides

Similarly, C₂- and C₃- arylations of azaarene *N*-oxides are also reported in reactions involving formation of aryne intermediate. The reaction offers a metal-free route to access hydroxyphenyl pyridines in presence of silylaryl triflates as a precursor for benzyne intermediate. Larock group revealed a methodology in which treating pyridine *N*-oxides with silylaryl triflates in presence of cesium fluoride (CsF) provided 3-(2-hydroxyphenyl)pyridines (**38**).^[51] Liu's group modified the conditions by introducing a base to obtain 2-(2-hydroxyphenyl) substituted pyridines (**39**) (Scheme 1.19).^[32]

Free radical intermediates are highly reactive reaction intermediates and they have been used widely in the functionalization of otherwise non-reactive C–H bonds. In this regard, cross dehydrogenative non deoxygenative coupling (CDC) reactions between azine *N*-oxides and ethers have been reported recently. The reaction occurs in presence of *tert*-butyl hydroperoxide as a radical initiator to generate alkyl radical intermediates. This intermediates further undergo coupling with heterocyclic *N*-oxides and results in non-deoxygenative C–H alkylation (Scheme 1.20).^[52]

Scheme 1.19 Benzyne mediated C–H functionalization of pyridine *N*-oxides

R¹
$$\stackrel{\bigcirc}{}$$
 $\stackrel{\bigcirc}{}$ $\stackrel{}{}$ $\stackrel{\bigcirc}{}$ $\stackrel{}{}$ $\stackrel{\bigcirc}{}$ $\stackrel{}{}$ $\stackrel{}{}$ $\stackrel{}{}$ $\stackrel{}{}$ $\stackrel{}{}$ $\stackrel{}{}$ $\stackrel{}{$

Scheme 1.20 Nondeoxygenative C–H functionalization via radical intermediates

1.1.3 Electrochemical C–H functionalization in the absence of transition metal catalysts

Electricity as an electron source for carrying out direct functionalization of inert C–H bonds have attracted attention in recent years. This is mainly attributed to their environmentally benign properties. [53] Electrochemistry also allows versatile and nonclassical disconnection approaches for retrosynthesis in an atom efficient manner. Conventionally used redox reagents are hazardous and toxic. Usage of such chemicals in stoichiometric amounts at higher temperature involves a great risk. Many of the common redox reagents like lead acetate, osmium tetroxide, sodium hydride, etc. and radical initiators like azobisisobutyronitrile (AIBN) are the recommended

reagents. For these reasons, organic chemists are showing a renewed interest in electrochemistry. [54] In addition to the safety, atom efficiency and mild reaction conditions, the electrochemical methods for organic synthesis possess other advantages as well. As electricity is a syringe pump of electrons with numerous variables, it can help chemists to develop unprecedented redox reactions. Electrochemistry can overcome the limitations of synthetic route design based on redox potentials of available oxidizing and reducing agents. In addition, reaction kinetics could also be improved. Recently, there are a number of reports showcasing the potential of electroorganic synthesis in transition metal catalyzed cross-coupling reactions as a terminal oxidant in complex catalytic cycles. [55]

In comparison to a classical organic reaction, there are a number of variable reaction parameters and cell types for an electrochemical reaction set up. The electrolysis can be carried out in an undivided cell or in a divided cell. The undivided cell electrolysis involves only one chamber where all the reagents, solvents, catalyst, etc. are put together to carry out the electrolysis. This setup is preferred in terms of ease of set up. However, it is not always possible to carry out electrolysis in a single cell altogether because of the undesired side reactions from counter electrodes. In this case, the cells need to be separated by a semipermeable membrane to avoid such side reactions (Figure 1.3). [56]

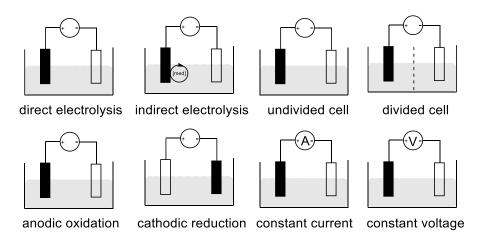


Figure 1.3 Types of electrolysis for electrochemical reactions

A redox mediator could be employed for electrolysis if the direct electrolysis is not feasible.^[57] It will undergo single electron transfer (SET) on the electrode surface which further oxidizes or reduces the substrate to initiate desired chemical transformations. Variation and modifications in

the electrochemical parameters, cell types and electrode arrangements are allowing electroorganic chemists to discover novel reactivities of organic molecules including PROTACs in a highly selective manner as shown in the Scheme 1.21.^[58]

Scheme 1.21 Chemoselective electrochemical reduction of a PROTAC-relevant molecule

Application of electrochemistry in organic synthesis has a long history. In the 1830s, Faraday pioneered the application of electric current for preparative organic chemistry. After a decade, Kolbe's electrolysis was discovered for the decarboxylative C–C bond formation *via* anodic oxidation of carboxylic acids. An important milestone in this area was achieved when the concept of redox mediator was introduced in the mid-20th century. Soon after, various powerful redox catalysts such as triarylamine mediators and nitroxyl radical mediators were applied in electrosynthesis. Also, transition metals and halide mediators are playing a significant role as redox mediators in electrosynthesis. Moeller's work on chiral electrodes in the 1970s opened up a new area of asymmetric electrosynthesis.

Yoshida and co-worker's enormous contributions include the electroauxillary concept to decrease the redox potential of compounds by introduction of special functional groups and cation pool strategy to carry out electrolysis at lower temperatures. [63] Now, researchers are showing a renewed interest in the area of electroorganic synthesis. Baran's electrooxidation methods for unreactive C–H bonds, Waldvogel's studies on biaryl coupling reactions, Ackermann and Mei's work on C–H activation using transition metals, cross dehydrogenative coupling reactions (CDC) developed by Lei *et al.*, Xu's work on electrochemical generation of nitrogen centered radicals

and Lin's asymmetric electrocatalytic methods are promoting this field. In the context of scale-up and efficiency, development of electrochemical reactions under flow conditions is an advancing field. [54, 64-66] The likes of Noël and Wirth are making significant contributions for electroorganic synthesis under flow conditions. [67-68] Photo-electrochemistry methods combine the power of light and electricity to enable powerful transformations which would have been extremely challenging otherwise. This research area is making rapid progress as indicated by the reports from Lambert group and Wickens group. [69] Moreover, availability of standardized commercial equipment has made the adoption of electroorganic synthesis easier and reproducible in research laboratories and chemical industries. These factors have allowed electrochemistry to become more accessible and enabled the application of novel chemistry.

1.1.4 Electrochemical direct C–H bond amination methods

As two complementary research areas, electroorganic synthesis and C–H functionalization methods go hand in hand. Numerous discoveries in this field have allowed more efficient and green synthetic routes. Among them, electrochemical C–H bond amination in the absence of transition metals as redox mediators is one of the prominent areas of research. Nitrogen-containing compounds are of great importance when it comes to biologically active molecules. Direct formation of C–N bond *via* inert C–H bond functionalization is an important strategy for synthesis and derivatization of *N*-heterocyclic compounds. To carry out these transformations in the absence of any transition metals and hazardous redox reagents is highly appealing.^[56]

Direct electrolysis is a simple setup electrolysis where the substrates are oxidized or reduced directly at the electrode surface in a chemical transformation. However, the risk of side reactions is more if there are other redox active substrates inside the cell or if the products formed can further undergo overoxidations. Nevertheless, by careful optimization of reaction parameters, many useful intramolecular and intermolecular C–H amination methodologies are reported in literature. To this end, Moeller's report on synthesis of *N*-tosyl pyrrolidines (44) by intramolecular coupling of electrochemically generated sulfonamidyl radical (43a) to olefins is highly relevant (Scheme 1.22).^[70]

Later in 2013, Yoshida and coworkers revealed an efficient amination of arenes using pyridines. Selective anodic oxidation of arenes in a divided cell in presence of pyridine generates *N*-

arylpyridinium ions (**46**). Reacting this intermediate with an alkylamine furnished the aromatic primary amines (**47**) under mild conditions. However, the reaction conditions are mostly compatible with activated arenes containing electron donating groups (**47a-d**) while the unactivated arenes are less favored for amination (Scheme 1.23).^[71]

R3 R4 R2 Et₄NOTs (0.1 M) N R1 OMe N-BuLi (0.5 equiv.) Ts MeOH

R3 R4 TsN R2 TsN
$$R^2$$
 R^3 R^4 R^2 R^3 R^4 R^4 R^2 R^4 R^4

Scheme 1.22 Direct electrolysis of *N*-sulfonamidyl group for pyrrolidine synthesis

selected examples:

Scheme 1.23 Yoshida's C–H amination of electron rich *via N*-arylpyridinium ions

Recently, a metal-free approach to synthesize isoxazolidine-fused isoquinoline-1(2*H*)-ones (49) was demonstrated by Li and coworkers. The reaction takes place *via* direct electrolysis of *N*-alkoxy amide scaffolds (48) in 95% ethanol as green solvent (Scheme 1.24). Generation of *N*-centered radicals through indirect electrolysis is an important strategy for C–H bond amination reactions. [72]

Scheme 1.24 Tricyclic *N*-heterocycles via cascade cyclization

As mentioned earlier, employing a redox mediator as an electrocatalyst has its own advantages. Many reactions have been developed with high selectivity under milder conditions using this approach. One of the earliest reports is when Shono demonstrated electrolysis of tosylamides (50) in presence of halide mediators and base to furnish pyrrolidines as in Hofmann-Löffler-Freytag-type cyclization (Scheme 1.25).^[73]

Scheme 1.25 Electrochemical Hofmann-Löffler-Freytag-type cyclization

Very recently an intermolecular [3+2] annulation for the synthesis of indolines was discovered by Lei and co-workers. The methodology uses DDQ as an electrocatalyst. The same reaction in the absence of a redox catalyst proceeds with a much lower yield. Mechanistically a *N*-radical formed (**52a**) *via* anodic oxidation is proposed to resonate to form the C-radical (**52b**). This after radical addition with the olefin followed by a second oxidation by DDQ or anode provides the indoline products (**54**) (Scheme 1.26). [74]

In an attempt to obtain radical diazidation of alkenes using TEMPO as a redox catalyst, Lin *et al.* observed the azidoxygenation reaction to be favored over diazidation. This eventually requires stoichiometric amounts of TEMPO because of its consumption during the reaction.^[75] To overcome this issue and to obtain diazidation products under metal-free conditions, the authors synthesized a number of analogues of TEMPO with an increased steric crowd at the *N*-oxyl center. This revealed cyclohexane-substituted (4-acetamidopiperidin-1-yl)oxyl (**56**, CHAMPO) as an efficient catalyst for the diazidation of various types of alkenes effectively (Scheme 1.27).^[76]

Scheme 1.26 DDQ mediated [3+2] annulation for the synthesis of indolines

$$R^{2} \xrightarrow{R^{3}} R^{3} \xrightarrow{C \bowtie Pt} R^{3} \xrightarrow{R^{3} \times R^{4}} R^{2}$$

$$R^{4} \xrightarrow{R^{4} \times R^{2}} R^{4} \xrightarrow{R^{2} \times R^{2}} R^{4$$

Scheme 1.27 CHAMPO as an efficient catalyst for the diazidation

A novel tetraarylhydrazine catalyst (**59**) capably catalyzed intramolecular [3+2] annulation to yield imidazo-fused heteroaromatic compounds (**58**). The reaction proceeds between *N*-centered radical and *C*-radical generated by indirect electrolysis in presence of base under constant current electrolysis under reflux temperature (Scheme 1.28).^[77]

Scheme 1.28 Tetraarylhydrazine as redox mediator for *N*-heteroaryl synthesis

$$R^{1} = R^{1} + R^{2}$$

$$R^{1} + R^{2} + R^{3}$$

$$R^{2} + R^{4} + R^{3}$$

$$R^{4} + R^{4} + R^{3} + R^{4} + R^{4$$

Scheme 1.29 Electrochemical generation of nitrene intermediate for synthesis of aziridines

Electrochemical generation of nitrene and nitrenium species enable many interesting amination reactions. Harsh oxidizing agents such as lead tetraacetate (Pb(OAc)₄) had been used to generate reactive intermediate nitrene from *N*-aminophthalimide (**60**). Yudin's group revealed that under constant potential electrolysis of +1.8 V (vs Ag/AgCl), it is possible to generate such intermediates and can be applied for the aziridination of olefins (**62**). The concept of overpotential was brilliantly exploited to obtain aziridines from electron rich olefins which have more or less similar oxidation potential as that of phthalimide moiety.^[78] The reaction conditions were modified by Little, Zeng and coworkers to make it more accessible. A redox mediator in the form of tetrabutylammonium

iodide was employed to carry out electrolysis under constant current in an undivided cell using cheaper electrodes (Scheme 1.29).^[79] Halogens are used as electrochemical mediators in many other reactions as well.

The concept of electrochemical generation of hypervalent iodine and subsequent use of it to carry out important transformations has been studied in detail. One such example is for the oxidative spirocyclization reaction for azacyclic derivatives as reported by Nishiyama group. Electrolysis of iodobenzene in presence of trifluoroethanol as solvent results in the formation of hypervalent iodine intermediate (64). This acts as an oxidizing agent to generate nitrenium ion intermediate from alkoxy amide substrate. The following nucleophilic attack of the aromatic ring resulted in the spirocyclic product which are synthetic intermediates (65) of complex bioactive compounds (Scheme 1.30).^[80]

Scheme 1.30 Electrochemical generation of a hypervalent iodine reagent and its application

Scheme 1.31 Shono type electrochemical amination of γ -lactams

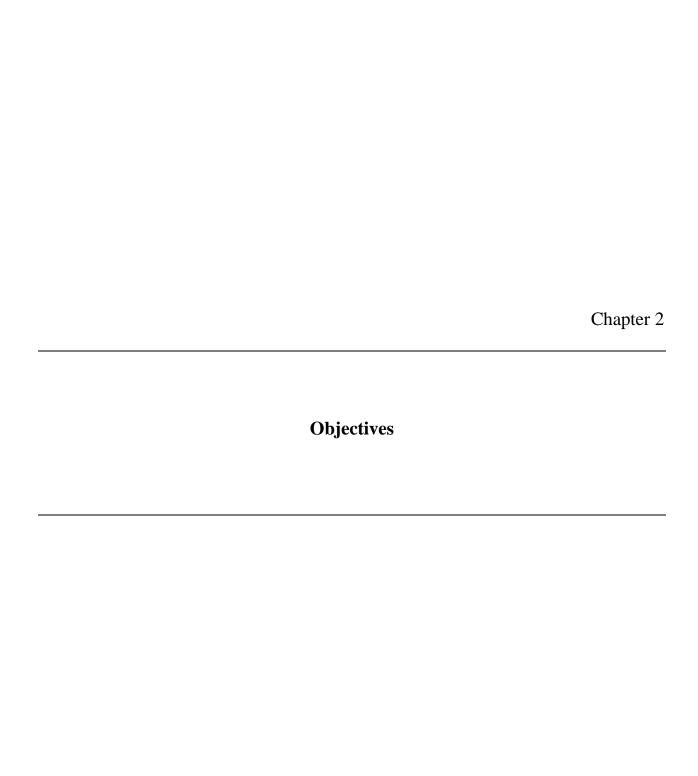
Shono oxidation is a classical approach to functionalize α -position of N-acyl compounds. Usually anodic oxidation of compounds ultimately results in the formation of iminium ions and alcoholic solvent attacks the iminium carbon to get corresponding functionalized products. Numerous

modifications of this method have been reported over years. Direct C–H amination reactions of *N*-methyl pyrrolidones (**66**) involving an iminium cation (**66b**) under metal-free conditions have also been reported recently (Scheme 1.31).^[81]

Flow chemistry has gained significant attention owing to its unique advantages over conventional flask methods. Efficient scale up and reproductivity of electrochemical reactions with control over reactivities can be achieved through combining it with continuous flow synthesis. Wirth *et al.* designed an interesting electrochemical microflow reactor and demonstrated its efficiency by successfully generating nitrogen and sulfur radicals for the synthesis of corresponding heterocycles. For instance, an intramolecular reaction of electrochemically generated *N*-acyl radical in flow reacts with alkene and following radical coupling of TEMPO give rise to corresponding TEMPO substituted *N*-aryl isoindolinones (**69**) (Scheme 1.32).^[82]

Scheme 1.32 Isoindolinone synthesis in an electrochemical flow microreactor

It is noteworthy to mention that care should be taken to categorize a reaction as a transition metal free reaction. The starting material preparation before the key C–H bond functionalization step might involve a transition metal. It is possible to carry forward trace amounts of these metals for the next steps as well. In a way, it is more correct to coin the term "externally added TM-free C–H bond functionalization". A number of studies related to TM-free cross coupling reactions such as organocatalytic Suzuki-Miyaura coupling are considered doubtful in this aspect.^[83] Even the metal contaminated magnetic stir bars seemed to catalyze the classic cross-coupling reactions to an extent.^[84] These studies however are still beneficial for the science to develop new reactions with extremely low catalytic loading of expensive and precious transition metals.



2 Objectives

Organocatalysis and oxidative coupling reactions allow access to novel organic molecules without the use of any transition metal catalysts. Requirement for the pre-functionalization of starting materials in conventional cross-coupling reactions will increase the number of steps to synthesize a desired product. In this aspect, development of novel direct C–H functionalization methods in a transition metal-free manner is highly advantageous. These methods do not need harsh reaction conditions and pre-functionalization of starting materials to corresponding halides or other functionalities. Instead, direct coupling reaction between the non-pre-functionalized substrates could be achieved. Considering these benefits, efforts will be made to develop novel reaction methodologies of this class especially for the easier access to highly functionalized nitrogen-containing compounds.

Isoquinoline and other closely related nitrogen heterocycles are abundant in natural products and synthetic pharmaceuticals. Chemical architectures of naturally occurring alkaloids such as papaverine and noscapine are based on a C₁-benzylic substitution of isoquinoline and tetrahydroisoquinoline moieties. To access these classes of compounds methodologies based on organocatalytic inert C–H functionalization are rare. Keeping this in mind, development of a cross-coupling method between heteroaromatic *N*-oxides and organosilanes was proposed. This method will provide easy access to benzylated *N*-heterocycles without the use of expensive transition metals. Furthermore optimization, evaluation of the scope and mechanistic studies were also planned. The feasibility of methodology to enable alkynylation of *N*-heterocyclic compounds will be examined.

Scheme 2.1 Regioselective cross-coupling of *N*-oxides with organosilanes

Nitrenium ions are important reaction intermediates and development of methods to access these intermediates electrochemically would help to access various *N*-heterocycles in a sustainable

Objectives 29

manner. The existing methodologies for the electrochemical C–H bond amination based on nitrenium ion generation use stoichiometric quantities of redox mediators and electrolytes. Therefore, a novel direct electrolytic amination method involving an electrochemically generated nitrenium ion intermediate was proposed. The feasibility of the proposed reaction will be tested. The optimization of reaction conditions as well as substrate scope evaluation will be carried out. The compatibility of the methodology for both intramolecular and intramolecular aminations needed to be examined.

Scheme 2.2 Electrochemical dehydrogenative C–H bond aminations

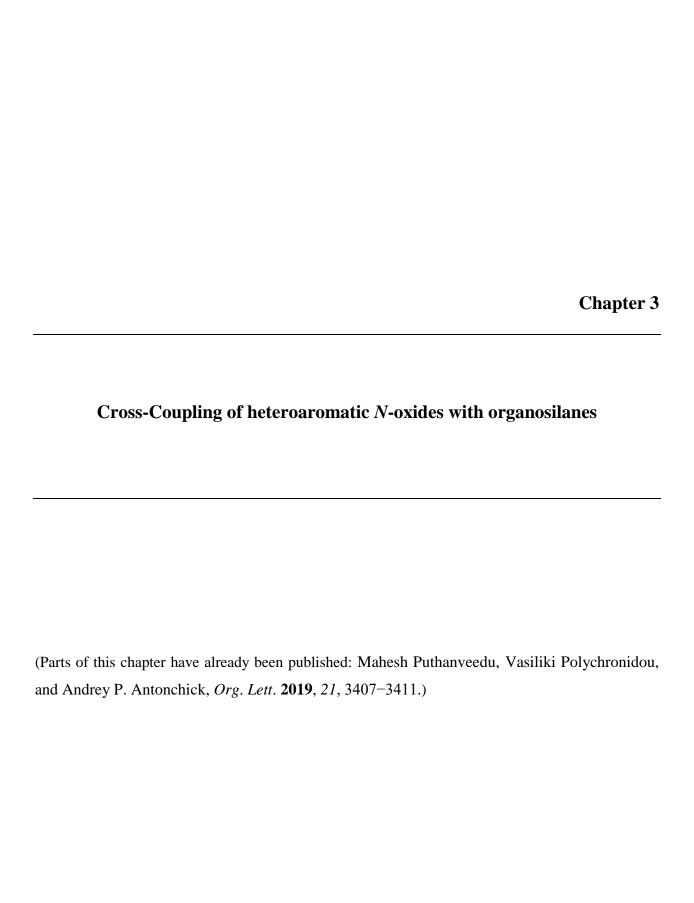
A 2015 report based on oxidative C–H amination of chromones using azoles as coupling partner reported a number of novel C₂-aminated chromones as Hedgehog inhibitors. Later on, a novel biological assay found these compounds to be potent bone morphogenetic pathway effectors. Based on these results, the synthesis of the most active analogue for various biological studies will be carried out. In addition, to have a better insight into the structure-activity relationship, synthesis of a number of closely related analogues will be carried out to evaluate their biological effects. Also, the compounds lacking specific substituents are required for chemical biology studies as inactive probes.

$$R^{1} \xrightarrow{O} \qquad \frac{\text{azole,}}{\text{DMF}} \qquad R^{1} \xrightarrow{O} \qquad R^{2}$$

Scheme 2.3 Design and synthesis of C₂-aminated chromones

All novel molecules obtained during the reaction developments, substrate scope evaluation and mechanistic studies would be submitted to the COMAS (Compound Management and Screening Centre) in Dortmund.

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3 Cross-Coupling of Heteroaromatic N-oxides with Organosilanes

3.1 C-H Benzylation

3.1.1 Introduction

Nitrogen containing heteroaromatic compounds are important scaffolds in organic chemistry. They are encountered very often in bioactive natural products and synthetic drugs. Therefore, their synthesis and direct functionalization reactions are of particular interest. Direct C–H functionalization methods are gaining popularity in drug discovery and process chemistry because of their unique advantages. Application of this methodology to synthesize and functionalize pharmaceutical compounds allows atom economic production of drugs in a sustainable way. Not only that, C–H functionalization methods allow faster structure activity relationship studies and in some cases synthesis of previously inaccessible analogues. Privileged scaffolds such as quinoline, isoquinoline and pyridine are highly relevant in medicinal chemistry. Direct C–H bond functionalization reactions of these scaffolds have been studied widely in recent years with most of them using precious transition metals as catalysts.

Figure 3.1 Representative bioactive compounds having benzyl substituted *N*-heterocycle

In this regard a transition metal-free approach for the synthesis of benzylated nitrogen heterocycles *via* direct C–H bond functionalization was highly demanding. This is because many naturally occurring alkaloids and synthetic drugs such as papaverine, berberine, moxaverine, etc. carries benzylated azacycles, especially isoquinoline, as a key scaffold in their structure (Figure 3.1). Historically, isoquinoline derivatives were synthesized using classical Bischler-Napieralski cyclization reactions (Scheme 3.1).

 R^1 = alkyl, aryl, halogen, etc.

 R^2 = alkyl, aryl, benzyl, etc.

Scheme 3.1 Bischler-Napieralski cyclization reactions for isoquinoline synthesis

With the rapid development of cross-coupling chemistry, approaches to readily synthesize benzylated isoquinolines and the natural products within the class were revealed. These methodologies however demand harsh reaction conditions, highly expensive metal catalysts and specific metal coordination ligands.^[85]

Scheme 3.2 Transition metal mediated direct C–H benzylation of isoquinolines

Minisci reaction is a radical functionalization method to obtain functionalized pyridines in presence of a radical source. This concept has been modified by several groups to obtain various functionalized nitrogen heterocycles *via* C–H bond functionalization methods.^[86] However, more

than one reactive position for radical reactions as in the case of quinoline (C₂- and C₄-) will result in the formation of a mixture of regioisomers. One such successful example of a selective Minisci type C–H benzylation is an oxidative decarbonylative coupling of aliphatic aldehydes with azaarenes in presence of di-*tert*-butyl peroxide as radical initiator and oxidant.^[87] An independent report from Liu and coworkers used simple methyl arenes as coupling partner for azacycles enabling a cross dehydrogenative coupling (CDC) for the synthesis of C₁-benzyl isoquinolines (75). The reaction proceeds in the presence of catalytic amounts of ytterbium triflate and di-*tert*-butyl peroxide as oxidant (Scheme 3.2).^[88]

Scheme 3.3 Regioselective benzylation of heteroaromatic *N*-oxides

Because of the advantages such as improved regioselectivity and step economy of deoxygenative functionalization, heteroaromatic *N*-oxides have gained importance as substrates to synthesize functionalized nitrogen heterocycles. On the other hand, organosilanes are a class of stable, affordable and less toxic alternatives for cross-coupling reactions. Trimethylsilyl cyanide has been successfully employed for the direct deoxygenative *cine*- C–H cyanation of pyridine *N*-oxides to afford 2-cyanopyridine compounds (73).^[89] Murakami, Miura and coworkers developed a strategy to synthesize benzylated and secondary alkyl substituted pyridines by means of photocatalysis. The authors proposed a radical species generated photocatalytically from alkenes will be highly electrophilic. It will then react with pyridine *N*-oxide followed by intramolecular radical addition to *ortho*- position and consequent elimination of a carbonyl compound to furnish C₂- alkylated pyridine compounds.^[90] However, the long reaction time up to 7 days of stirring at room

temperature might interrupt adopting this strategy largely for benzyl pyridine synthesis by organic chemists (Scheme 3.3).

3.1.2 Motivation and objectives

Bering and Antonchick revealed a facile strategy for the C–H bond functionalization of quinoline N-oxides using boronic acids as a coupling partner. The methodology enabled transition metal and external oxidant free synthesis of C_2 -(hetero)arylated and alkenylated quinolines in an efficient manner. Interestingly, the reaction design was based on an analogy of the structure of quinoline N-oxides to a key reaction intermediate formed during the Petasis reaction. This was the first instance of application of the Petasis reaction for direct functionalization of heterocycles. Even though the scope of reaction was broad with respect to quinolines, (hetero)aromatic compounds and alkenes, the reaction was not applied to isoquinoline and pyridine scaffolds. In addition, only $C(sp^2)$ - $C(sp^2)$ bond formation was successful under reported conditions (74) (Scheme 3.4).

Scheme 3.4 Regioselective arylation and alkenylation of *N*-oxides with boronic acids

Considering the abundance of various C₂-substituted nitrogen heterocycles such as isoquinolines and pyridines, novel approaches based on the concept of transition metal-free C–H functionalization methods would be highly appealing. As in the case of boronic acids, organosilicon compounds are also important coupling partners to make new C–C bonds *via* crosscoupling reactions. Based on this similarity, it was hypothesized that replacing boronic acids with organosilicon compounds could help to overcome the limitations of known methods (Scheme 3.5).

Scheme 3.5 Proposed benzylation of isoquinoline *N*-oxides using organosilanes

3.1.3 Initial results and optimization

The feasibility of proposed cross-coupling reaction was examined by reacting isoquinoline Noxide with benzyltrimethylsilane in presence of substoichiometric amount of tetrabutylammonium fluoride (TBAF) under air at room temperature using tetrahydrofuran (THF) as solvent (Table 3.2). Fluoride activation is important to initiate the reaction. Nucleophilic attack of isoquinoline N-oxide to this activated silane complex and following nucleophilic migration of benzyl group was proposed to furnish benzylated isoquinoline product. As proposed, a trace amount of desired product formation was observed upon initial screening (entry 1 and entry 2). From screening a number of solvents, either THF or dimethylformamide (DMF) were found to be most suitable (entry 3-5). However, in all cases additional undesired products were also observed. Careful isolation and characterization of side products from the reaction identified two major side products impacting the better conversion of starting materials to desired product. One of them was 1benzoylisoquinoline which was formed by the oxidation at the benzylic position of the formed product. However, this side reaction was suppressed to a large extent by carrying out the reaction under argon atmosphere. 45% yield of product was obtained by reacting isoquinoline N-oxide with benzyltrimethylsilane under argon atmosphere in presence of 10 mol% of TBAF as catalyst and DMF as solvent (entry 6). Increased catalyst loading was not beneficial and furnished lower yields of product (entry 7). Using 3 equivalents of silane coupling partner was found to be advantageous. Addition of TBAF made the reaction vial hot. But lowering the temperature during the catalyst addition did not facilitate better conversion (entry 8). Another important observation made was that the benzylated isoquinoline N-oxides are also formed in high quantities during reaction. The main target was to develop the deoxygenative C-H functionalization to directly obtain C₁functionalized isoquinolines. So, the efforts were directed towards in situ reduction of unreduced

Table 3.1 Representative conditions of the optimization for the cross-coupling.^[a]

Entry	76a (equiv.)	Cat. (mol%)	Additives (equiv.)	Temp. (°C)	Solvent (1 mL)	Time (h)	Yield (%) ^[b]
1 ^[c]	2	TBAF (10)		rt	THF	24	trace
2 ^[c]	2	TBAF (10)		rt	DMF	24	trace
3	2	TBAF (10)		rt	NMP	24	n.d.
4	2	TBAF (10)		rt	EtOH	24	n.d.
5	3	TBAF (10)		rt	THF	6	37
6	3	TBAF (10)		rt	DMF	6	45
7	3	TBAF (20)		rt	DMF	6	19
8	3	TBAF (10)		$0 \rightarrow rt$	DMF	6	trace
9	3	TBAF (10)	PhB(OH) ₂ (2)	rt	DMF	24	trace
10 ^[d]	3	TBAF (10)	$Et_3SiH(5)$	rt	DMF	12	59
11 ^[d]	3	TBAF (10)	$Et_3SiH(5)$	rt	DMF	24	62
12 ^[e]	3	TBAF (10)	$Et_3SiH(5)$	rt	DMF	24	41
13 ^[f]	3	TBAF (10)	Et ₃ SiH (5)	rt	DMF	24	78
14	3	CsF (10)	$Et_3SiH(5)$	rt	DMF	24	30
15	3	TBAT (10)	$Et_3SiH(5)$	rt	DMF	24	29
16	3	TBAF (10	$Et_3SiH(5)$	rt	THF	24	7
17	3	TBAF (10	Et ₃ SiH (5)	rt	DCE	24	n.d.
18	3	TBAF (10	Et ₃ SiH (5)	rt	Dioxane	24	7

Entry	76a (equiv.)	Cat. (mol%)	Additives (equiv.)	Temp. (°C)	Solvent (1 mL)	Time (h)	Yield (%) ^[b]
19	3	TBAF (10	Et ₃ SiH (5)	rt	EtOAc	24	12
20	3	TBAF (10)	$Et_3SiH(7)$	rt	DMF	24	66
21	3	TBAF (10)	Et ₃ SiH (10)	rt	DMF	24	68
22	3	TBAF (10)	$Et_3SiH(3)$	rt	DMF	24	65
23	3	TBAF (10)	PMHS (1)	rt	DMF	24	trace
24	3	TBAF (10)	PhMe ₂ SiH (5)	rt	DMF	24	58
25	3	TBAF (10)	Me ₃ Si-SiMe ₃	rt	DMF	24	42
26	3		$Et_3SiH(5)$	rt	DMF	24	n.d.

 $^{[a]}Reaction$ conditions: **75a** (0.4 mmol), solvent (0.4 M), argon. $^{[b]}yields$ are given for isolated products (**77a**) after column chromatography. $^{[c]}under$ air atmosphere. $^{[d]}Et_3SiH$ was added before the TBAF addition. $^{[e]}Et_3SiH$ after 12 h. $^{[f]}Et_3SiH$ just after TBAF addition. TBAF = tetrabutylammonium fluoride. PMHS = polymethylhydrosiloxane. TBAT = tetrabutylammonium difluorotriphenylsilicate. rt = room temperature (~28 °C). n.d. = not detected.

product to C₁-benzyl isoquinolines. Along this line, various additives were screened (entry 9-25). It was noted that silyl hydride as additives lead to formation of desired product in higher yields. 5 equivalents of triethylsilane (Et₃SiH) was added to a mixture of isoquinoline *N*-oxide and benzyltrimethylsilane. After addition of 10 mol% of TBAF the reaction mixture was stirred under argon for 12 hours to isolate 59% of cross-coupled product (entry 10). Increasing the reaction time to 24 hours resulted in a slightly higher yield of product (entry 11). Interestingly, time of addition of triethylsilane was crucial for optimal yield of product. A diminished yield was observed when Et₃SiH was added after 12 hours of stirring of starting materials with catalyst (entry 12). The optimal yield of 78% of desired product was obtained when the additive was added just after the addition of catalytic fluoride source, which was found to be TBAF (entry 13). Other fluoride sources such as cesium fluoride (CsF) and tetrabutylammonium difluorotriphenylsilicate (TBAT) were also examined (entry 14 and entry 15). In addition to Et₃SiH, other silane compounds such as dimethylphenylsilane, hexamethyldisilane, polymethylhydrosiloxane (PMHS), etc. were also evaluated for the efficacy in this reaction (entry 22-25). Similarly, using THF, ethyl acetate (EtOAc) or dioxane as solvents were not ideal. The best condition was found to be cross-coupling

in presence of TBAF and Et₃SiH using DMF as solvent, stirring at room temperature under argon atmosphere for 24 hours. A reaction in the absence of any catalysts did not furnish 1-benzylsioquinoline (entry 26).

3.1.4 Scope of heterocyclic *N*-oxides and benzyltrimethylsilanes

With the optimal reaction conditions in hand, the substrate scope of reaction was examined. All the heterocyclic *N*-oxides were synthesized by reacting them with *meta*-chloroperbenzoic acid (*m*CPBA) (Scheme 3.6).^[28] Various substituted benzyltrimethylsilane derivatives are prepared by reacting trimethylsilyl chlorides with *in situ* generated Grignard reagents of corresponding benzyl bromides (Scheme 3.7).^[91]

Scheme 3.6 Preparation of heteroaromatic *N*-oxides

Scheme 3.7 Preparation benzyltrimethylsilanes

To evaluate the scope, respective N-oxides of isoquinoline, quinoline and pyridine were reacted with simple benzyltrimethylsilane initially. It was found that all three classes of these important nitrogen heterocycles provided the corresponding benzylated N-heterocyclic product in good yields. Isoquinoline N-oxide and quinoline N-oxide were reacted with benzyltrimethylsilane under standard conditions to obtain 78% and 79% of C_1 - and C_2 - benzylated products respectively (77a and 77b). In case of pyridine N-oxide, better yield of product was obtained in the absence of triethylsilane additive (77c). But, the catalyst loading was increased from 10 mol% to 20 mol% for optimum product formation. Various substituted heterocyclic N-oxides were also examined. For instance, electron donating substituents on the C_6 - position of quinoline was well tolerated

(77d and 77e). Similarly, effects of substitutions at other positions of *N*-oxide compounds were also studied (77f-77h). With respect to the time taken to consume the starting materials, quinoline analogues were found to be faster to provide benzylated quinoline compounds, followed by pyridines and then isoquinolines.

Later on, different benzyltrimethylsilane analogues as substituted benzyl group donors were tested. In case of isoquinoline *N*-oxides, a *para*- isopropyl group and a *para*- phenyl group on the phenyl ring reacted to yield corresponding products in moderate yields (77i and 77j). In addition, 1-napthyl and 2-naphthyl derivatives were also successfully coupled with isoquinoline *N*-oxide (77k and 77l). Synthesis of quinoline derivatives using developed protocol was superior to that of isoquinolines. For instance, substitutions on the *para*- position of the phenyl ring of the silane component had no significant deteriorating effect on the reaction performance. Electron donating groups such as methoxy, isopropyl group and electron withdrawing halogen were all compatible (77m-77o) under these conditions. Highly substituted trimethyl(2,4,6-trimethylbenzyl)silane effectively coupled with quinoline *N*-oxide to furnish 77p in 70% yield in 6 hours. Furthermore, C₂- substituted pyridine derivatives were synthesized by established cross-coupling protocol. For these products 77q and 77r, the reaction was catalyzed solely by tetrabutylammonium fluoride and use of additional triethylsilane as an additive was negated (Scheme 3.8).

With respect to the limitations of this protocol, it was found that highly electron deficient heterocycles are not suited. This could be justified because of the decreased nucleophilicity of oxide functionality. A nitro- group or a cyano- group would decrease the electron density of heterocyclic *N*-oxide and thereby prevent the nucleophilic attack of the *N*-oxide group to activate silanes. This limitation however provides useful directions for the plausible mechanism of reaction (Scheme 3.9). Delightedly, the reaction scale-up was also found to be successful. 1 g scale reaction of isoquinoline *N*-oxide (**75a**) with benzyltrimethylsilane (**76a**) under optimized conditions provided 68% of isolated product (**77a**). No further modifications were required to the optimal conditions of the reaction (Scheme 3.10).

Scheme 3.8 Scope of cross-coupling between heterocyclic *N*-oxides and organosilanes. ^[a]Reaction conditions: **75** (0.4 mmol), **76** (1.2 mmol, 3 equiv.), TBAF (10 mol%), DMF (0.4 M). ^[b]Yields are given for isolated products (**77**). ^[c]20 mol% of TBAF was used. ^[d]Without Et₃SiH.

Scheme 3.9 Limitations of cross-coupling methodology. n.d. = not detected

Scheme 3.10 Scale-up experiment of the developed benzylation methodology

3.1.5 Mechanistic considerations

To understand the reaction mechanism various control experiments were performed (Table 3.2). Various fluoride sources catalyzed the reaction under standard conditions (entry 1-3). However, using a stoichiometric amount of base failed to initiate the transformation (entry 4). The starting materials remain unconsumed in the absence of any fluoride source (entry 5). This supported the initial thought of fluoride activation of silanes in the first step of reaction. The reaction happens in the absence of triethylsilane as additive but with lower yields (entry 6). To understand the role of triethylsilane in the reaction, a control experiment was carried out in the absence of a coupling partner, which is benzyltrimethylsilane. Interestingly, the *N*-oxide was simply reduced to yield isoquinoline **78** (Scheme 3.11). In this case, the necessity of addition of triethylsilane after addition of fluoride catalyst was understood. The role of triethylsilane was hypothesized to be a reducing

Table 3.2 Control experiments for fluoride catalysis^[a]

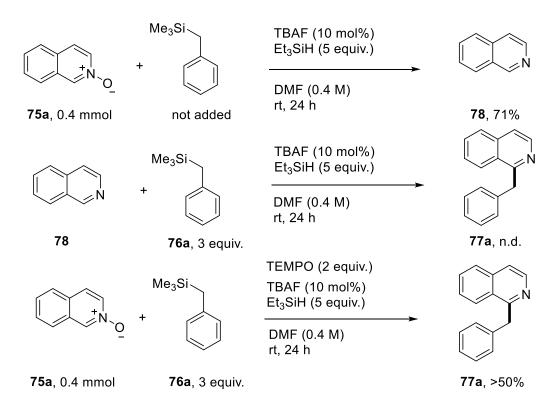
Entry	Cat.	Additives (5 equiv.)	Yield (%) ^[b]
1	TBAF (10 mol%)	Et ₃ SiH	78
2	CsF (10 mol%)	Et ₃ SiH	30
3	TBAT (10 mol%)	Et ₃ SiH	29
4	KOtBu (2 equiv.)	Et ₃ SiH	n.d.
5	-	Et ₃ SiH	n.d.
6	TBAF (10 mol%)	-	45

[a]Reaction conditions: 75a (0.4 mmol), 76a (1.2 mmol, 3 equiv.), DMF (0.4 M), rt, argon. n.d. = not detected.

agent to transform C₁-benzylated isoquinoline *N*-oxides to C₁-benzylated isoquinoline. The preoxidation of heterocycle is necessary, because simple isoquinoline failed to react with the organosilanes coupling partner under fluoride catalysis to give coupling product. A high C₁ regioselectivity for isoquinolines and C₂ regioselectivity for quinolines and pyridines indicated a 1,2-migration type of mechanism as in the case of previously reported boronic acid coupling. However, to negate a radical mediated reaction, radical trap experiments were performed. The reaction performance had no significant decrease in presence of 2 equiv. of TEMPO as radical scavenger. These results from control experiments and the pattern of substrate scope suggested a reaction mechanism involving formation of a hypervalent silicon intermediate (Scheme 3.11).

Silicon can afford 4-,5- and 6- coordinate complexes. Benzyltrimethylsilane has a 4-coordinate silicon. This is a poor Lewis acid because it cannot accept a lone pair of electrons. The R group attached to 4-coordinate silicon is a poor nucleophile (Scheme 3.12). A nucleophilic attack of

fluoride ion to benzyltrimethylsilane **76a** results in the formation of a 5-cooridnate silicon **75b**. In comparison to the 4-coordinate silicon, pentacoordinate silicon is a good Lewis acid. Hence, the nucleophilic oxygen attacks the silicon which results in the formation of hexacoordinate silicon intermediate **75c**. The R group, which is a benzyl group attached to a hexacoordinate silicon, is a very good nucleophile. It will undergo a nucleophilic migration to the C₁ position of isoquinoline *N*-oxide. The following rearomatization step gives the desired product **77a**. with the regeneration of the catalyst. In the process, trimethylsilanol **79** is formed as a byproduct which will polymerize to form hexamethyldisiloxane and other polysiloxanes. [92-93]



Scheme 3.11 Control experiments for mechanistic studies of benzylation.

Scheme 3.12 Characteristics of hypervalent silicon complexes

Scheme 3.13 Proposed reaction mechanism of C₁-benzylation

3.1.6 Identification of biological activity

Compound libraries of C₁-benzylated isoquinoline were submitted to COMAS (Compound Management and Screening Centre) in Dortmund to screen for the biological activities. Cell-based assays to screen for autophagy inhibitor activities and hedgehog signaling pathway inhibition were carried out. After screening, the compound **77p** was identified as an Autophagy inhibitor in the low micro molar range. However, a rapamycin induced autophagy inhibition assay in presence of compound showed no activity.

Autophagy $IC_{50} = 6.86 \mu M$

RAPA IC_{50} = inactive

Furthermore, the phenotypic screening of compounds was also carried out using Cell painting assay which is highly useful for retrieving the biological information from morphological characteristics of cellular components.^[94] From the library, 4 compounds were reported to have significant biosimilarity to reference compounds. However, the reference compounds for active hits were not very similar and the hit frequencies of these reference compounds are high. The compounds are suggested to possess the highest similarity to functional clusters of Lysosomotrop.

Table 3.3 Representative results of cell painting assay

Compound	Conc.	Activity	Induction [%]	Highest sim. to a reference [%]	Toxic
77 p	30	active	6.0	78.9	No
77 p	10	inactive	1.2		No
77k	10	active	5.0	73.9	No
77d	10	active	11.1	84.4	No
77h		active	14.0	86.2	No

3.2 C-H Alkynylation

3.2.1 Introduction

Having success on developing a metal-free methodology for $C(sp^2)$ - $C(sp^3)$ bond formation, focus was diverted to other silanes which would make $C(sp^2)$ - $C(sp^2)$ and $C(sp^2)$ - C(sp) connections with N-heterocycles. In this regard, trimethyl(phenylethynyl)silane was first tested as a coupling partner because of the biological significance of C_2 -alkynylated scaffolds (Figure 3.2).

Figure 3.2 Representative bioactive compounds having alkynyl substituted N-heterocycle

Scheme 3.14 Deoxygenative alkynylation of quinoline *N*-oxides using. phosphazene super base

Kondo and co-workers elucidated a method for deoxygenative alkynylation of quinoline *N*-oxides using. phosphazene super base **80** as catalyst in presence of silylated nucleophiles (Scheme 3.14).

Afterwards, the same group modified the conditions and reported a procedure for coupling terminal alkynes with heteroaromatic *N*-oxides to provide 2-alkynyl substituted nitrogen heteroaromatics.^[95] The phosphazene organic superbase was replaced by an onium amide base in this method and thereby improving the ease of access for reagents.^[96] Chupakhin *et al.* had reported a complementary method to Sonogashira cross-coupling reaction for the synthesis of ethynylazines using potassium *tert*-butoxide. In the reaction, phenylacetylide of potassium is generated. Quinoxaline *N*-oxide readily reacts with this reagent at -20 °C in anhydrous DMF to yield corresponding ethynylazines.^[97] Recently a potassium hydroxide catalyzed visible light mediated reaction has also been developed for selectively alkynylating the azine *N*-oxides (Scheme 3.15).^[98]

Scheme 3.15 C–H functionalization methods 2-alkynyl substituted nitrogen heteroaromatics

3.2.2 Motivation and objectives

A most common approach for introducing alkynyl group is carrying out Sonogashira cross-coupling reactions. This reaction usually requires transition metal catalysts, base and harsh reaction conditions (Scheme 3.16).^[99] Alternatively, aforementioned methods for the synthesis of alkynylated quinoline and pyridine are known. The reported methodologies are carried out in presence of strong bases such as phosphazene super base, potassium *tert*- butoxide, etc. So, there is still room for further improvements. Additionally, the synthesis of selectively C–H alkynylated

isoquinoline derivatives under transition metal-free conditions is not well studied. For these reasons, the feasibility of cross-coupling between isoquinoline *N*-oxide and trimethyl(phenylethynyl)silane was examined initially.

Scheme 3.16 Sonogashira coupling for alkynylation

3.2.3 Initial results and optimization

The cross-coupling reaction between isoquinoline *N*-oxide **75a** and trimethyl(phenylethynyl)silane proceeded smoothly to yield 91% of the coupled product **83** in just 2 hours (Scheme 3.17). Tetrabutylammonium fluoride was an efficient fluoride source for this transformation for carrying out the reaction in DMF at room temperature under argon atmosphere.

Scheme 3.17 C–H alkynylation of heterocyclic N-oxide using organosilanes

Scheme 3.18 Sonogashira coupling for 1,2-diheteroarylacetylenes

Unexpectedly using ethynyltrimethylsilane as a coupling partner resulted in the formation of a double coupling product (Table 3.4). Symmetrically heteroaryl disubstituted acetylene **85a** was obtained under transition metal-free conditions. This observation was encouraging in the sense that, an alternative Sonogashira coupling takes place only in presence of metals such as copper and palladium and additionally a base has to be added. More importantly, handling of gaseous alkyne is required and reaction conditions were harsh and long as well (Scheme 3.18). With these initial observations, further optimization of the reaction conditions was carried out. THF found to be the only suitable solvent in this transformation while DMF, MeCN or DCE was found to be not suitable for this cross-coupling between compound **75a** and compound **84** (entry 1-6).

Table 3.4 Representative conditions for the optimization of alkynylation^[a]

Entry	Equiv. of 84a	TBAF (mol%)	Solvent (1 mL)	Time (h)	Yield of 85a (%) ^b
1	3	10	THF	12	42
2	3	10	DMF	12	n.d.
3	3	10	MeCN	12	n.d.
4	3	10	DCE	12	n.d.
5	6	10	THF	12	72
6	6	5	THF	3	82
7	4	5	THF	3	89
8	4	3	THF	3	35
9	3	5	THF	3	36

[[]a]Reaction conditions: **75a** (0.2 mmol), solvent (0.2 M). [b]Isolated yields. n.d. = not detected.

In case of synthesis of 1,2-di(isoquinolin-1-yl)ethyne, 5 mol% of TBAF and 6 equivalents of ethynyltrimethylsilane was found to be optimal. The reaction yielded 89% of the desired cross-coupled product at room temperature in just 3 hours (entry 7). This was really encouraging to see, because unlike the Sonogashira method, inexpensive catalysts and harsh reaction conditions were not required at all. Instead, reaction at room temperature using easy to handle reagents and simple laboratory set up was more than sufficient to obtain these interesting classes of compounds. Further experiments did not improve the yield of product **85a** (entry 8 and entry 9).

3.2.4 Scope of heterocyclic *N*-oxides

After the optimization of reaction conditions, the methodology was applied for the synthesis of different coupled products. Different isoquinoline and quinoline compounds were evaluated for their compatibility.

It was found that, in addition to the isoquinoline scaffold, various substituted quinoline heterocyclic *N*-oxides also undergo double deoxygenative coupling reaction to produce disubstituted acetylene compounds. A 6-methyl substituted quinoline *N*-oxide reacted with ethyne containing trimethylsilyl group at 1-position to yield 1,2-bis(6-methylquinolin-2-yl)ethyne under mild conditions. However, a slight increase of catalyst loading to 10 mol% was important to get 65% of the product (85c). A strong electron donating substituent at the same position was also tolerated (85d). Interestingly, *N*-oxide of isoquinoline ring possessing methoxy substitution at both 6 and 7 position reacted well with 6 equivalents of ethynyltrimethylsilane to provide the product 85f. Also, R groups such as halogen or phenyl substitution of only 7-position of isoquinoline *N*-oxide substrate were also acceptable for the developed cross-coupling method (85g and 85h). In short, the scope of reaction is good and this method proved to be a powerful alternative for Sonogashira type coupling reactions for such a class of compounds. Unfortunately, the methodology could not be applied for pyridine *N*-oxides.

More importantly, most of the reported compounds are synthesized for the first time and this will allow to unveil interesting applications of this 1,2-heteroarylsubstituted acetylene scaffold for various applications. The results are summarized in the scheme below (Scheme 3.19). [a] [b]

Scheme 3.19 Scope of cross-coupling between heterocyclic *N*-oxides and organosilanes. ^[a]Reaction conditions: **75** (0.4 mmol), **84** (6 equiv.), TBAF (10 mol%), DMF (0.2 M). ^[b]Yields are given for isolated products (**77**). ^[c] 4 equiv. of **84** and 5 mol% TBAF was used. n.d = not detected.

3.2.5 Mechanistic considerations

A plausible mechanistic pathway was hypothesized in the scheme. Initially a proton abstraction from the terminal alkynes results in the generation of an alkynide ion. Fluoride ions act as a base in this process.^[100] The nucleophilic attack of alkynide ion to the silicon of ethynyltrimethylsilane results in the generation of bis(trimethylsilyl)acetylene. A molecule of ethyne is eliminated as gas from the reaction mixture. This results in strong effervescence upon addition of catalyst. Also, requirement of excess acetylene partner could be justified for this reason (Scheme 3.20). Literature reports suggest that this type of sigma bond metathesis of terminal alkynes has been reported earlier as well.^[101-102] Furthermore, a reaction starting with bis(trimethylsilyl)acetylene **86** and

isoquinoline *N*-oxide proceeded smoothly to yield 93% of the double coupling product **85a**. Heteroaromatic *N*-oxide first couple with *in situ* generated bis(trimethylsilyl)acetylene **86** to give the mono coupling intermediate. Another molecule of *N*-oxide moiety then attacks this intermediate to eventually give a double coupled final product **85a**.

Scheme 3.20 Mechanistic consideration of alkynylation reaction methology

3.2.6 Identification of biological activity

Compound libraries of symmetrically heteroaryl disubstituted acetylenes were submitted to COMAS (Compound Management and Screening Centre) in Dortmund to screen for the biological activities. Cell-based assays to screen for autophagy inhibitory activity and hedgehog signaling pathway inhibition were carried out. After screening, the compound **85a** was identified as a hedgehog pathway inhibitor in the low micro molar range.

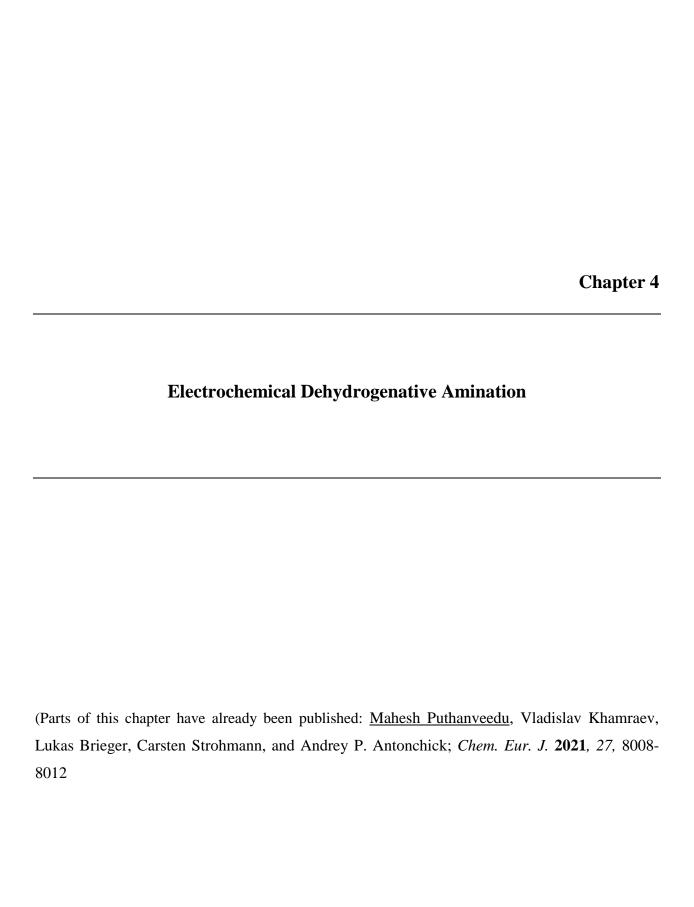
 $Hh~IC_{50}=0.26~\mu M$

Furthermore, the phenotypic screening of compounds was also carried out using cell painting assay (Table 3.5). From the library, 2 compounds were reported to have significant biosimilarity to reference compounds. Unfortunately, 10 µM concentration of isoquinoline was toxic to the cells.

Lowering the concentration leads to loss of activity. In the case of quinoline analogue (COMAS compound Id 409532), $10~\mu M$ solution showed a significant induction and similarity to the reference compound. Activity of reference compound suggests a potential to inhibit glucose transporter GLUT1. The frequency of this reference compound is only 0.16%, which is really good. But this compound also has a mild toxicity. Activity studies with lower concentration needs to be carried out.

Table 3.5 Representative results of cell painting assay

Compound	Conc.	Activity	Induction [%]	Highest sim. to a ref. [%]	Toxic
85a	10	active	31.4		Yes
85a	3	inactive	2.4		No
85a	1	inactive	0.7		No
85b	10	active	34.7	84.4	No



4 Electrochemical Dehydrogenative Amination

4.1 Introduction

Nitrenium ions are important intermediates in organic synthesis. Various transition metal-free C–H amination methodologies have been developed exploiting the high reactivity of *in situ* generated nitrenium ions. In this regard, Antonchick and co-workers discovered various oxidative annulation reactions and direct C–H bond amination reactions using hypervalent iodine reagents. For instance, an intramolecular C–H amination method towards carbazole scaffold (89) was developed under hypervalent iodine catalysis. The methodology was also applicable for the amination of unactivated arenes and also for diarylation of anilides (91) in presence of excess arene and stoichiometric amount of phenyliodine (III) diacetate (PIDA) (Scheme 4.1). [103]

Scheme 4.1 Hypervalent iodine mediated direct C-H bond aminations

Scheme 4.2 Organocatalytic annulation of benzamide compounds with alkynes

Also, an organocatalytic annulation of benzamide compounds with alkynes has been reported for the straight forward synthesis of isoquinolines (Scheme 4.2). The reaction was proposed to proceed *via* an *in-situ* generation of PIDA in presence of peracetic acid as an oxidant. The hypervalent iodine reagent then oxidizes the benzamide nitrogen to form electrophilic nitrenium ions. The attack of alkynes to the and followed nucleophilic attack of aryl ring results in isoquinoline scaffold. Similarly, nucleophilic attack of aryl ring to the nitrenium intermediates of benzamide derivatives results in direct C–H bond amination of aromatic and heteroaromatic compounds (Scheme 4.3). [105-107]

Scheme 4.3 Intermolecular amination mediated by hypervalent iodine reagents

Owing to their broad applicability and unique advantages in organic synthesis, efforts have been made to combine electroorganic chemistry with transition metal-free C–H bond functionalization. Nishiyama and coworkers applied electrochemistry to oxidize iodine(I) precursors to iodine(III) reagents (100) and employed these electrogenerated hypervalent iodine reagents to C–H amination reactions involving nitrenium ion intermediates (Scheme 4.4).

Scheme 4.4 Electrochemical generation of hypervalent iodine reagents

 $N-1-\beta$ -D-ribofuranosylmakaluvamine

Scheme 4.5 Electrochemical total synthesis of tetrahydropyrroloiminoquinone alkaloids

Moreover, the synthetic utility of these methods was demonstrated by applying them in the total synthesis of natural products such as tetrahydropyrroloiminoquinone alkaloids (101) (Scheme 4.5).^[108]

4.2 Intramolecular amination

4.2.1 Motivation and objectives

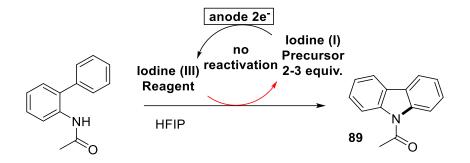
Atom economy of organic reactions is an important parameter with respect to sustainable chemistry. Direct amination of C–H bonds is an atom economic method to synthesize nitrogen heterocyclic compounds. In case of carbazoles, Buchwald and coworkers reported a highly atom economic strategy to access this class of compounds. The method was a novel C–H activation approach catalyzed by expensive palladium catalysts in presence of copper(II) acetate as an oxidant. Afterwards, numerous reactions of similar chemistry were reported using different transition metal catalysts such as copper, rhodium, iridium etc. In 2011, Antonchick and coworkers reported a novel transition metal-free organocatalytic approach for intramolecular C–H bond amination. The approach was based on the generation of electrophilic nitrenium ion intermediate in presence of hypervalent iodine reagents. This report later paved the way to the electrochemical synthesis of carbazole derivatives (Scheme 4.6).

Buchwald, 2005: Pd catalyzed

Antonchick, 2011: Hypervalent Iodine catalyzed

Scheme 4.6 Intramolecular amination of 2-amidobiaryls towards carbazoles

It is claimed that direct electrochemical synthesis is hard because of overoxidation, homo coupling and electropolymerization reactions. [113-114] For these reasons, the existing methodologies use an indirect electrosynthetic approach. This method is otherwise called the *ex cell* approach. In an *ex cell* electrosynthesis using iodine based redox mediator as a precursor for hypervalent iodine reagent, at first the electrolysis of iodine precursor is carried out in presence of fluorinated solvents to generate the active hypervalent iodine reagent. Then the electric current is turned off and the substrate is added to the electrolytic solution and stirred under specified conditions. [115-116] In such reactions, it is mandatory to use a stoichiometric amount of redox mediator because the active catalyst is just consumed and not regenerated after (Scheme 4.7).



Scheme 4.7 Ex cell electrochemical methods for amination reactions

Also, in case of intermolecular reactions, it is found that an excess of coupling partners has to be used with stoichiometric quantities of supporting electrolytes. Even though using quantities of supporting electrolytes and non-friendly constant potential electrolysis, a very recent report has allowed *in cell* electrolysis for C–H amination using hypervalent iodine catalysts.^[117]

An alternative could be the direct electrolysis of acetanilide derivatives to generate corresponding nitrenium ions. Waldvogel and co-workers delineated a possibility to generate nitrenium ion by a two-step oxidation of acetanilide. Not only they succeeded in the mechanistic investigations including a series of cyclic voltammetric studies to prove the two-step oxidation of nitrogen, but also demonstrated the utility by benzoxazole synthesis *via* intramolecular C–O bond formation. [118] However, a direct electrolytic strategy for such C–N bond formation remained unexplored. In this regard, a novel direct electrochemical strategy for intramolecular amination of C–H bonds mediated by nitrenium ion intermediate would be highly appreciated. This would open the door to more synthetic methods based on the direct electrochemical generation of nitrenium ions (Scheme 4.8).

Scheme 4.8 Proposed direct electrolytic approach to nitrenium ions

4.2.2 Initial results and optimization

Scheme 4.9 Initial results of direct electrolytic approach towards carbazole synthesis

Table 4.1 Representative conditions for the optimization of intramolecular amination^[a]

T4	D()	Electrolyte	Calaran 4	\$70 1 1[b]
Entry	Base (equiv.)	(1 equiv.)	Solvent	Yield ^[b]
1 ^[c]	-	n-Bu ₄ NBF ₄	HFIP	n.d.
2	-	n-Bu ₄ NBF ₄	HFIP	trace
3	KOtBu (2)	n-Bu ₄ NBF ₄	HFIP	57
4	KOtBu (2)	n-Bu ₄ NBF ₄	toluene	n.d.
5	KOtBu (2)	n-Bu ₄ NBF ₄	TFE	n.d
6	KOtBu (2)	n-Bu ₄ NBF ₄	DCE/HFIP (1:1)	40
7	KOtBu (2)	n-Bu ₄ NBF ₄	MeCN/HFIP (1:1)	33
8	KOtBu (1)	<i>n</i> -Bu ₄ NOAc	HFIP	43
9	KOtBu (1)	<i>n</i> -Bu ₄ NOAc	HFIP	43
10	KOtBu (1)	n-Bu ₄ NPF ₆	HFIP	47
11	KOtBu (1)	Et ₄ NOTs	HFIP	45
12	KOtBu (1)	-	HFIP	52
13	LiOtBu (1)	-	HFIP	59
14	Et ₃ N (2)	-	HFIP	56
15	DBU (2)	-	HFIP	45
16	NaOEt (1)	-	HFIP	60
17 ^[d]	NaOEt (0.75)	-	HFIP	63

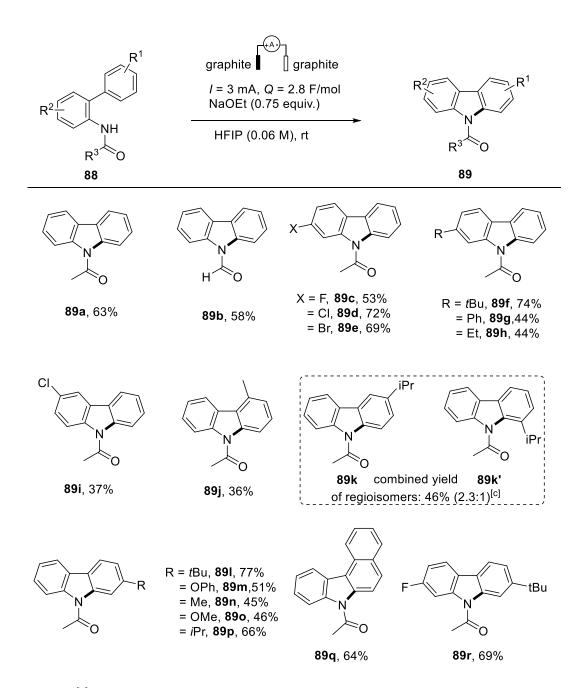
[[]a] Reaction conditions: constant current electrolysis (5 mA) for 3 h (3.2 F/mol) using graphite electrodes: **88a** (0.2 mmol), solvent (0.06 M). [b] Isolated yield of **89a** [c] Glassy carbon anode and platinum plate cathode. [d] 3 mA current and 2.8 F/mol charge passed in 5 h. n.d. = not detected.

The initial efforts to realize a direct electrolytic approach to generate nitrenium ion intermediate and to initiate the intramolecular cyclization of 2-amidobiaryl compounds were not successful. Instead of the desired products, a dimerization of the starting material was consistently observed. This is due to the ready formation and high reactivity of *N*-acyl radicals upon electrolysis. The N–N radical dimerization was initially reported by Baran and co-workers as a key step in the synthesis of natural product Dixiamicine B.^[119] In addition, mechanistic studies by Waldvogel,

Moeller and co-workers have also reported methodologies based on N-N radical dimerization reactions (Scheme 4.9). [118] At this point, modification of reaction conditions was necessary to shift the reaction pathway from radical homo-coupling to intramolecular C-H amination (Table 4.1). Only switching to graphite electrodes did not provide useful results (entry 2). A detailed literature review led to a previous report from Zeng group in which a base mediated proton abstraction from amide was proposed as the first step of an electrochemical benzylic C-H amination.^[120] Based on this, a base was added additionally to the reaction mixture for electrolysis. A constant current electrolysis using graphite electrodes in presence of 2 equivalents of potassium tert-butoxide in hexafluoroisopropanol solvent and tetrabutylammonium tetrafluoroborate as electrolyte provided a promising yield of 57% of desired product (entry 3). Various solvents including toluene, TFE and solvent mixtures were also screened (entry 4-7). Screening different electrolytes did not improve the yield of desired product (entry 8-11). However, using a base in an acidic solvent was found beneficial. Because it was found that the reaction still proceeds without huge resistance to render the reaction even in the absence of an external electrolyte (entry 12). Then further optimization revealed sodium ethoxide as a good choice (entry 13-16) which results in good yield of desired product with sub-stoichiometric quantities of base (entry 17).

4.2.3 Scope of intramolecular amination

To evaluate the substrate scope of the developed method, various substrates containing different substitutions at different positions of the aryl rings were prepared according to the known literature methods. Acetanilide derivatives were prepared using acetyl chlorides and triethylamine base. To make biaryl substrates, Suzuki coupling with respective boronic acids or palladium catalyzed *ortho* C–H activation was applied. [121-122] Afterwards, the synthesized substrates were examined for the compatibility of the developed electrochemical intramolecular amination reaction.



Scheme 4.10. ^[a] Reaction conditions: constant current electrolysis (5 mA) for 3 h (~ 3.2 F/mol) using graphite electrodes: **88** (0.2 mmol), solvent (0.06 M). ^[b] Isolated yield. ^[c] *n*-Bu₄NBF₄ (0.2 mmol, 1 equiv.) as supporting electrolyte. ^[d] Glassy carbon anode and platinum plate cathode. ^[e] 3 mA current and 2.8 F/mol charge passed in 5 h. n.d. = not detected.

Delightedly, many substrates reacted under electrochemical oxidation in presence of base to yield corresponding carbazole derivatives (Scheme 4.10). In addition to the acetyl protection, a formyl protected 2-amidobiaryl was also examined. Contrary to previous reports, the current method selectively yielded a five membered carbazole product instead of the usual six membered

isoquinolone analogue (**89b**). Afterwards, substrates containing halogen substitution at 4- position of the biaryl ring was electrolyzed under standard conditions. All tested halogen containing substrates cyclized to yield *N*-protected 2-halocarbazoles (**89c-e**). Similarly, different

2-amidobiaryl derivatives containing 4-alkyl or 4-phenyl substitutions were also successful substrates for this intramolecular (89f-h) electrochemical C–H bond amination. Furthermore, the synthesis of 3-chlorocarbazole and 4-methyl carbazole derivatives were realized albeit in lower yields (89i and 89j). 3'-isopropyl group containing biarylacetamide substrate resulted in a mixture of regioisomers as expected, with the combined yield of 46% in a ratio of 2.3 to 1 for two possible regioisomers (89k and 89k'). However, all tested substrates with 4' electron donating substitutions such as *tert*-butyl, methoxy, etc. provided corresponding carbazoles in good yields (89l-p). Substrates containing polyaromatic rings such as 1-naphthyl were also found to be an excellent candidate for this electrosynthesis (89q). Also, biaryl compounds with more than one substitution were also tested and found successful (89r). Electron rich substitution on the phenyl ring decreases the redox potential of the substrate. In addition, it also increases the reactivity towards electron-poor nitrenium ions. This property is reflected clearly in the trend of substrate scope of reaction as well.

4.2.4 Mechanistic considerations

Considering the pKa values of ethoxide ion and 1,1,1,3,3,3-hexafluoropropan-2-ol (OEt 16 and HFIP 9.3), the added base will react with solvent to form a salt. This salt negates the need of addition of external supporting electrolytes to carry out the electrolysis. This *in situ* electrolyte formation was further confirmed by carrying out the reaction using readily prepared sodium salt of HFIP as supporting electrolyte for the electrolysis (Scheme 4.11).^[123]

Scheme 4.11 Sodium salt of HFIP as supporting electrolyte

Also, during the optimization and scope studies, much evidence for the nitrenium ion intermediate were observed. Most important being the consistent formation of HFIP adducts. A control experiment using 4-methoxy acetanilide in HFIP resulted in the formation of mono and di adduct of HFIP with the substrate. Furthermore, the product **103**° was isolated and confirmed by NMR studies. The HFIP ether at both the *ortho*- positions of the substrate was also formed in trace amounts (**105**) (Scheme 4.12). Similarly, in case of intramolecular amination using potassium *tert* -butoxide as a base and tetrabutylammonium tetrafluoroborate as an electrolyte, almost 1:1 ratio of cyclized product (**89a**) and HFIP ether (**88a**°) of the biaryl compound was isolated (Scheme 4.13). It has to be noted that single electron transfer pathways for such compounds are not favored. Mostly, the HFIP ethers are formed by the nucleophilic addition reactions which strongly support the formation of nitrenium ions and following resonance structures.

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, $Q = 2.8 \text{ F/mol}$
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 $I = 3 \text{ mA}$, $Q = 2.8 \text{ F/mol}$
 $I = 3 \text{ mA}$, $Q = 2.8 \text{ M$

Scheme 4.12 Mono and di adduct of HFIP with the acetamido substrate

Scheme 4.13 Cyclized product and HFIP ether of the biaryl compound

Moreover, electron rich arenes are easy to oxidize as they possess a low oxidation potential. Once the nitrenium ion is formed, they prefer carbazole formation *via* nucleophilic attack. The less

nucleophilic electron deficient arenes with higher oxidation potentials favored HFIP adduct formation (eg.: 88s') via positive charge delocalization. This indicates a strong dependence on the nature of substitution on aryl rings (Scheme 4.14). Afterwards, cyclic voltammetric studies were carried out to get more insights into the reaction mechanism. In the voltammogram of compound 88a two distinct oxidation peaks are not visible (Figure 4.1). However, depending on the compounds and substitution pattern, the $E_{ox,1}$ peak and $E_{ox,2}$ can overlap with the current at the potential limit. The characteristic properties of the solvent 1,1,1,3,3,3-hexafluoroisopropanol (HFIP) play an important

.....

Scheme 4.14 Reaction pathway dependence on electron rich and poor substituents

role in this electrolysis. The ability to stabilize the charged intermediate, acidity to generate *in situ* electrolyte and an observed reduction of onset potential (1.45 V) are some of the important aspects to consider.^[125]

Scheme 4.15 Plausible mechanism of the reaction

Based on the control experiments and observations made during reaction development, a plausible mechanism has been outlined in the following scheme. A speculation was made considering the pKa values of involved chemicals. It suggests that a direct proton abstraction of amidyl nitrogen is possible only with a stronger base. However, the sodium ethoxide used here in the reaction mixture is small in quantity which would eventually form the salt with acidic solvent hexafluoroisopropanol. Also, the anion of solvent cannot directly abstract a proton from the substrate. For these reasons, it has to be believed that the reaction starts with an oxidation of 2-amidoarylsubstrate at anode resulting in the formation of a cation radical **A**. This cation radical however is highly acidic in nature. The oxyanion of solvent then abstracts the proton to generate the highly reactive *N*-acyl radical intermediate **B**. Which actually can undergo homodimerization as observed in the initial screening reactions or would eventually undergo a second oxidation to

generate the key nitrenium ion intermediate **C**. A nucleophilic attack of the aryl ring to nitrenium ion to form **D** and following aromatization step provide the desired carbazole derivative **89a**. Another possibility is that the resonance of the positive charge results in a Wheland intermediate and form adduct **88a**' with hexafluoroisopropanol anion.

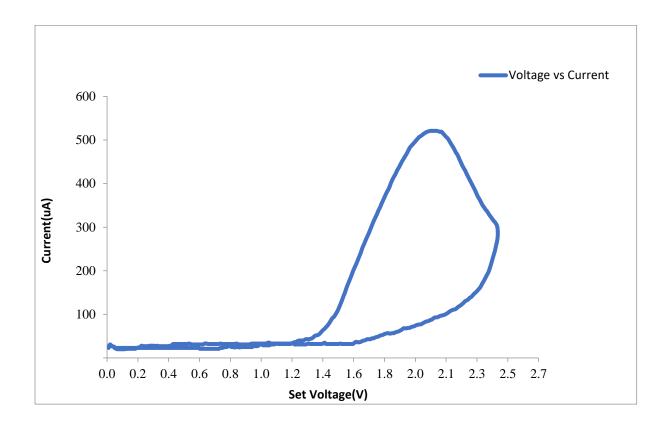


Figure 4.1. Cyclic voltammograms of **88a** (1 mM) and 0.75 equiv. NaOEt (21% solution in EtOH) in HFIP (3 mL) using supporting electrolyte n-Bu₄NBF₄ (0.1 M). WE, Glassy Carbon; CE, Platinum plated electrode; RE; Ag/AgCl in 3 M aqueous KCl. Scan rate = 200 mV/S. X-axis, voltage (V); Y-axis, current (μ A).

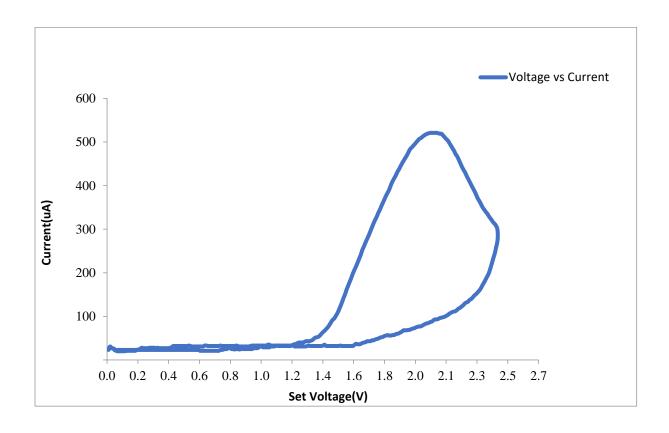


Figure S3. Cyclic voltammograms of **88a** (1 mM) and 0.75 equiv. NaOEt (21% solution in EtOH) in HFIP (3 mL) using supporting electrolyte n-Bu₄NBF₄ (0.1 M). WE, Glassy Carbon; CE, Platinum plated electrode; RE; Ag/AgCl in 3 M aqueous KCl. Scan rate = 200 mV/S. X-axis, voltage (V); Y-axis, current (μ A).

4.3 Intermolecular amination

4.3.1 Motivation and objectives

Direct amination of unactivated C–H bonds under electrochemical conditions is an appealing strategy to obtain *N*-arylated compounds in a sustainable manner. Francke and coworkers first reported a redox mediator mediated methodology towards this research. An interesting iodine-based mediator was designed and synthesized for this purpose with the characteristics of recyclability and reusability (Scheme 4.16). The iodine precursor **104** was also linked with an alkyl chain constituting a supporting electrolyte at the *para* position of the phenyl ring. The benzylic position was protected from oxidation by equipping a carbonyl group there. In this indirect electrosynthesis, the iodine(I) mediator was electrochemically oxidized to the active iodine(III) reagent **105** in HFIP (Scheme 4.16). Afterwards an excess of arene coupling partner along with the acetanilide derivatives were added to the pre-electrolyzed solution. This enabled the C–N coupling reaction between the coupling partners. After the reaction the mediator-salt is readily recovered and can be reused. Based on this appealing strategy, various *N*-heterocycles and acetanilide derivatives were arylated successfully (Scheme 4.17). [115]

Scheme 4.16 Indirect electrolysis using a recyclable mediator

Very recently, Powers *et al* came up with another method which enabled the catalytic use of iodine(I) reagent. In this redox mediated electrolysis, the iodine(I)/iodine(III) redox process was continuous over the whole reaction time by electrochemically reactivating the iodine(I) precursor. However, the reaction uses an undesired amount of two different electrolytes and 5 equivalents of acid additives under constant potential conditions (4.18).^[117] A direct electrolytic approach similar

to aforementioned intramolecular amination methodology might drastically simplify the reaction conditions and improve the green characteristics. For these reasons, a constant current electrolytic cross-amination of arenes with in situ generated electrolyte using an easily available and simple inorganic base would be highly appealing (Scheme 4.19).

Scheme 4.17 Indirect electrolysis for C–H amination of arenes

Scheme 4.18 Hypervalent iodine catalyzed electrochemical amination

$$R^{1} \xrightarrow{H} R^{2} + \underbrace{het} R^{3} \xrightarrow{het} R^{$$

Scheme 4.19 Proposed strategy for direct electrolytic intermolecular amination

4.3.2 Initial results and optimization

N-(4-methoxyphenyl)acetamide (108a) and 1-methylnaphthalene (109a) were initially tested under electrochemical conditions (Table 4.2). As for the intramolecular amination part, potassium tert-butoxide was first used as the base for this arylation of acetanilide substrate. Interestingly, a constant current electrolysis using graphite electrodes in hexafluoroisopropanol as solvent successfully cross-aminated the 1-methylnaphthalene in excellent yields (entry 1). So, the focus was given to reduce the amount of coupling partners for this reaction (entry 2). Indeed, 2 equivalents of aryl coupling partner was sufficient to react with acetanilide and provide the product 110a in excellent yield of 92% (entry 3). Decreased amount of base had a slight impact on the reaction performance. Other bases and an electrolyte were also effective in this intermolecular coupling reaction even though with slightly lower efficiency (entry 4-6).

Table 4.2 Representative conditions of the optimization for the cross-coupling.^[a]

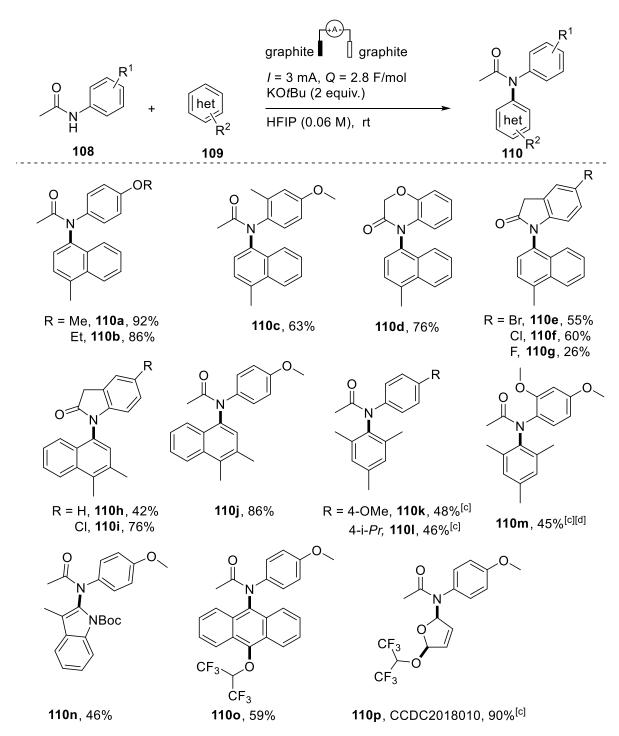
graphite(+)lgraphite(-)
$$I = 3 \text{ mA, } Q = 2.8 \text{ F/mol}$$
base (x equiv.)
$$HFIP (0.06 \text{ M}), \text{ rt}$$

$$108a \qquad 109a \qquad 110a$$

Entry	109a (equiv.)	Base (equiv.)	Yield ^[b] (%)
1	5	KOtBu (2)	90
2	2	KOtBu (2)	92
3	2	KO <i>t</i> Bu (1)	88
4	2	NaOEt (2)	90
5	2	Et ₃ N (2)	86
6 ^[c]	2	-	88

[[]a]Reaction conditions: constant current electrolysis (3 mA) until 2.8 F/mol charge was passed using graphite electrodes: **3a** (0.2 mmol), solvent (0.06 M, 3 mL) [b]Isolated yield. [c]*n*-Bu₄NBF₄ (0.2 mmol, 1 equiv.) as supporting electrolyte. rt = room temperature.

4.3.3 Scope of intermolecular amination



Scheme 4.20 Substrate scope of intermolecular amination. [a][b] [a] Reaction conditions: constant current electrolysis (I = 3 mA) until 2.8 F/mol charge (Q) was passed using graphite electrodes for ~ 5 h: 3 (0.2 mmol), 4 (0.4 mmol, 2 equiv.), potassium *tert*-butoxide (0.4 mmol, 2 equiv.), HFIP (0.06 M, 3 mL). [b] Isolated yield. [c] 5 equiv. of arene was used as coupling partner. [d] 1:1 DCM: HFIP (3 mL).

The optimized reaction conditions were then applied for the arylation and heteroarylation of numerous compounds (Scheme 4.20). Various N-phenylacetamide containing substitution at paraposition undergo cross coupling reactions with 1-methylnaphthalene 109a to give the corresponding products in moderate to excellent yields (110a-110c). Heterocyclic compound benzoxazinone (108d) was also found to be a suitable coupling partner for the electrochemical arylation of N-containing compounds. In addition, oxindole derivatives such as 5-bromo, 5-chloro and 5-fluoro oxindole reacted with 1-methyl and 1,2-dimethylnaphthalene coupling partners under constant current electrolysis to yield products 110e-110i. The yields of these products were also very good. Other arene and hetero(arene) coupling partners also partook in our electro-oxidative cross-amination method. However, in case of mesitylene as arylation partner, an excess of this partner was necessary to get decent yield. Similarly, conditions were slightly modified for arylation of substrate with 2,4-dimethoxy functionality (110k-110m).C₂-arylation of N-Boc protected 3methylindole was also achieved under standard conditions to give 110n. Surprisingly, the electrochemical amination reaction of a polyaromatic ring containing anthracene resulted in the simultaneous formation of C-N and C-O bonds as a result of amination plus HFIP ether formation (1100). When furan was used as a partner for heteroarylation of acetanilide, an unprecedented dual C–H functionalization resulted in the formation of compound **110p**.

4.3.4 Identification of biological activity

Compound libraries of electrochemical C-H amination were submitted to COMAS (Compound Management and Screening Centre) in Dortmund to screen for the biological activities. Cell-based assays to screen for autophagy inhibitory activity and hedgehog signaling pathway inhibition were carried out. After screening, the compound was identified as an autophagy inhibitor at low micromolar concentration.

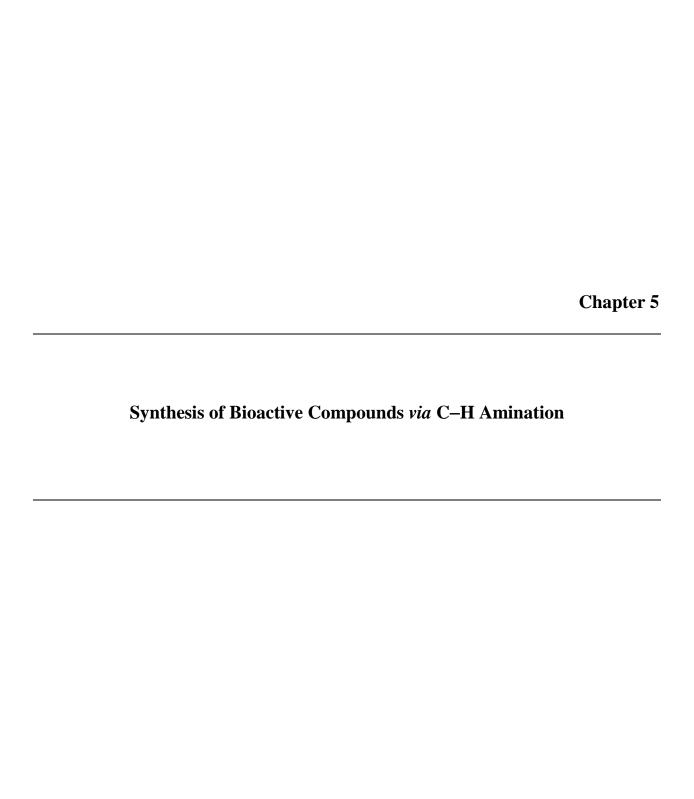
Furthermore, the phenotypic screening of compounds was also carried out using cell painting assay. From the library, 2 compounds were reported to have significant biosimilarity to reference compounds. But in most of the cases the frequency of hits for these similar compounds is high. For instance, acetanilide compound had a similarity of 94.2 with a RNA polymerase inhibitor Dasabuvir. But the frequency percentage was as high as 3.45%. Also, lowering the concentration to $10 \,\mu\text{M}$ leads to loss of activity.

Autophagy $IC_{50} = 4.01 \mu M$

RAPA $IC_{50} = inactive$

 Table 4.3 Representative results of cell painting assay

Compound	Conc.	Activity	Induction [%]	Highest sim. to a ref. [%]	Toxic
110f	50	active	52.5	81.7	No
110f	30	acive	21.9	74.6	No
110f	10	inactive			No
110c	30	active	49.7	94.9	No
110c	10	inactive			No



5 Synthesis of Bioactive Compounds via C-H Amination

5.1 Introduction

Inert C–H bond functionalization chemistry enables easy access to highly functionalized heterocyclic compounds. This in fact provides an opportunity to evaluate the biological properties of previously unknown chemical structures. This is an additional advantage of C–H bond functionalization chemistry. A novel methodology can supply solutions to synthetic chemistry challenges and additionally when the obtained novel chemical compound, when studied might possess interesting biological activities. That resembles a research at the interface of organic chemistry and biology, while the approach is purely chemistry driven. One such example was reported by Antonchick and co-workers in 2015. [126] The authors developed a novel regioselective amination method for chromones using azoles as the cross-coupling partner (Scheme 5.1). Afterwards, the compounds were evaluated for their bioactivities and found that these novel compounds are potent hedgehog signaling pathway inhibitors (HH inhibitors). Small molecule HH inhibitors are particularly important because of their role as potential anticancer agents. Mutations in the HH signaling pathway have been identified as causes of cancers such as basal cell carcinoma and medulloblastoma. [127]

working hypothesis:

Scheme 5.1 Hedgehog signaling pathway inhibitors *via* C–H amination

Interestingly, the HH signaling pathway has close association with other biological signals like that of bone morphogenetic proteins (BMPs). Small molecule bone morphogenetic protein (BMP)

activators are also an important class of compounds in chemical biology and medicinal chemistry.

5.2 Identification of a novel osteogenic BMP activator chemotype

Bone Morphogenetic Proteins (BMPs) are a group of signalling molecules belonging to the Transforming Growth Factor-β (TGFβ) superfamily. They have crucial roles in various organ systems of the body such as in embryogenesis, adult tissue homeostasis and regeneration. BMPs help to differentiate mesenchymal stem cells to bone. So BMPs play a central role in bone and cartilage formation. Additionally, many processes in early development such as cell growth, apoptosis and cell differentiation are highly dependent on the BMP signaling pathway. BMPs have diverse functions in almost all organ systems and are known to be regulators throughout the body. Deficiency of these proteins have marked effects in various biological systems such as neurological, cardiovascular, skeletal system, etc. [128]

Because of their prominent role in osteogenesis, BMPs and small molecule activators of BMPs are highly relevant for the treatment of skeletal trauma and osteopenic diseases. The available treatments of difficult fractures are based on recombinant proteins. But they are costly and have various adverse effects mainly because of their 10,000-fold beyond doses when compared to physiological concentrations. Small molecules that can activate BMP signaling are highly desirable. BMP activator modalities have various applications in biotechnology, chemical biology and regenerative medicine. However, the discovery and development of such small molecule growth factor/cytokine activators is intrinsically challenging. In this regard, Schade *et al.* devised a physiological, morphogenic cellular screening system that is focused on the BMP pathway for the discovery of novel BMP activators. Using this stem cell-based screening method, almost 7000 compounds were screened for their bone morphogenetic activation abilities. These efforts furnished 4*H*-chromen-4-ones as a novel osteogenic BMP activator chemotype that potentiated BMP signaling outputs.

Interestingly this compound belongs to the compound library of previously reported HH inhibitor chemotype synthesized *via*. C–H bond amination. This is not surprising considering the close association between hedgehog signaling pathways and BMP signaling pathways. Based on this result, other similar compounds were also tested as potential activators of BMPs.

 $IC_{50} \text{ mESC} = 0.2 \pm 0.08 \ \mu\text{M}$

5.3 Design, synthesis and biological evaluation of chromone derivatives

For the structure activity relationship studies, previously reported compounds as well as novel analogues were also evaluated. The synthetic route for the compounds is outlined in the following scheme. At first, the C₃ position of chromone **112** was brominated using hydrobromic acid, in presence of Oxone and triethylamine as base. This was followed by a palladium catalyzed crosscoupling with aryl halide to obtain the C₃-arylated derivatives **114**. The key C–H functionalization at C₂-position using corresponding azoles provided the required compounds **116** for biological studies. Additionally, the triazole ring at C₂-position was replaced with stronger nucleophiles to obtain more structurally diverse analogues (Scheme 5.2).

HBr, Oxone then Et₃N
$$R^{1}$$
 R^{1} R^{2} R^{1} R^{2} R^{1} R^{2} R^{2}

Scheme 5.2 Synthetic route for bioactive chromones

The BMP hit chromenone (2-(1*H*-1,2,4-triazol-1-yl)-3-(4-(trifluoromethyl)phenyl)-4*H*-chromen-4-one) **116a** was prepared on a higher scale to produce the compound in sufficient quantity for

various biological assays and *in vivo* studies. The key amination step was carried out using potassium carbonate as base in presence of 1.5 equivalents of molecular iodine. Reaction with 1 mmol scale of chromone starting material yielded 82% of required compound **116a** (Scheme 5.3). With respect to the mechanism of this amination method, initially molecular iodine reacts with triazole **115a** to form an electrophilic iodine reagent **C**. Reaction between isoflavone and this reagent results in an oxonium ion intermediate **A**. Following nucleophilic substitution and dehydroiodination mediated by base results in the formation of product **116** (Scheme 5.4).

Scheme 5.3 Scale-up synthesis of bioactive chromone 115a

Scheme 5.4 Mechanistic considerations of key C–H amination step

Following the same protocol, a number of analogues were also prepared previously by Antonchick *et al*. The general strategy for the cross-coupling is based on the cross-coupling reaction of various substituted and unsubstituted chromones with different types of azoles in presence of molecular iodine and a base.^[126] In some cases, however, 1,2,4-triazole is used in addition to azole in excess quantities. Also, azole was replaced with more stronger nucleophiles by heating with a base to obtain more structurally diverse analogues. Some of these analogues were resynthesized whenever it was necessary to do so (Scheme 5.5).

Scheme 5.5 Previously known analogues of chromones for resynthesis

Schade *et al.* conducted various biological experiments and fully validated chromone **116a** as a novel osteogenic BMP activator chemotype (Figure 5.1). Based on these encouraging results,

novel chromones mainly focusing on the 2- and 3-positions of the 4*H*-chromen-4-one scaffold were designed and synthesized as potential chemical biology tools for BMP signaling.

CF₃

O

N

N

116a

117

118

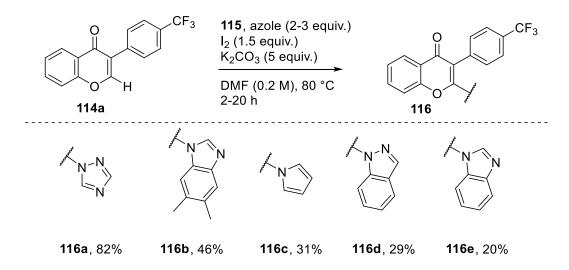
IC₅₀ [
$$\mu$$
M] mESC = 0,2 ± 0,08
EC₅₀ [μ M] C2C12 = 0,24 ± 0,15 (n=4)

inactive

inactive

Figure 5.1 Novel BMP activator analogue and closely related analogues

A total of 29 derivatives including newly synthesized analogues were profiled in the BMP-dependent morphogenic assays (Scheme 5.7 and Scheme 5.8). The data showed that the 3-substituent was quite essential for activity. Compounds containing no substituent at C2-position were inactive. Another analogue which has no *p*-trifluoromethyl moiety on the phenyl ring at C2-position was rendered inactive as well. Other similar compounds screened were also inactive, including compounds such as 4-methoxy phenyl, 3-chlorophenyl or 2-toluyl chromones.



Scheme 5.7 Synthesis of novel analogues *via* direct C–H amination

The 2-position also did not tolerate many alterations as the 2-(1,2,4-triazolyl) substituent turned out essential for robust biological activities and could only be replaced by a pyrrole substituent (116c), albeit with lower potency for this derivative. After that, other derivatives containing cyclic

aliphatic substituents containing nitrogen were also tested in addition to other big heterocyclic compounds (116f-k). Additional combinations that included variations in the six- and seven-position also did not furnish novel BMP activator characteristics. Although a vast number of derivatives were not tested, it was obvious that a small heteroaromatic triazole ring at C₂-position and a *para* electron withdrawing group containing phenyl ring at C₃- position were the most important pharmacophoric features. The other closely related chromones without these substitutions at respective positions would serve as important inactive probes for chemical biology applications. These compounds were evaluated in the human cells to understand their biological properties in the context of osteogenesis. Indeed, Chromone 116a, but not its inactive analogues, efficiently enhanced BMP-4-induced mineralization of human SaoS-2 cells at a dose of 25 nM. Therefore 2,3-disubstituted 4*H*-chromen-4-ones were identified as a new chemotype of BMP potentiators with high potency and efficacy with a unique mode-of-action. This compound (116a) induced a pronounced, kinase-independent, negative TGF-β feedback that enhanced nuclear BMP-Smad signaling outputs. These results provide novel BMP activators and important inactive chromones as novel chemical biology probes and potential therapeutic options.

Scheme 5.8 Synthesis of novel analogues *via* nucleophilic substitution

		Chapter 6
	Summary	

6 Summary

Development of novel transition metal-free C–H functionalization methods have enabled faster, selective and sustainable organic transformations. These methods are highly relevant when it comes to the direct functionalization of otherwise inert bonds and thereby providing useful solutions to the conventional organic chemistry challenges. Additionally, various oxidative and organocatalytic annulation methods give access to various heterocyclic molecules.

A novel regioselective organocatalytic cross-coupling between heteroaromatic *N*-oxides and organosilanes was developed. The methodology enabled synthesis of benzylated *N*-heterocyclic molecules in a transition metal-free manner. Tetrabutylammonium fluoride was found to be the best fluoride activator of organosilanes and triethylsilane as an additive was proved to be advantageous for the transformation. Scope and mechanistic studies of C–H benzylation reactions were also carried out. Based on these data, a plausible mechanism of the reaction has been proposed. Additionally, the versatility of the developed methodology was demonstrated by extending the method for site-selective deoxygenative C–H alkynylation of isoquinoline and quinoline *N*-oxides.

Scheme 6.1 Regioselective cross-coupling of *N*-oxides and organosilanes

A sustainable electrochemical dehydrogenative C(sp²)—H amination involving an electrochemical oxidation-induced nitrenium ion intermediate has been established. The application of electrochemical methods was justified by the metal-free direct electrolytic synthesis of carbazole scaffold as well as direct cross-amination of aromatic and heteroaromatic compounds. The reaction used a simple and easy to use undivided cell. Direct electrolysis using cheap and easily available graphite electrodes provided a straight forward C–H amination method without the use of any expensive transition metals as catalysts or electrodes. Additionally, use of external supporting

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electrolyte was not required in presence of added base and an acidic solvent which is HFIP. This is beneficial because the use of stoichiometric amount of external supporting electrolytes results in larger amount of waste from the reaction. Some of the compounds from this compound library were suggested to have some biological effects as well.

Scheme 6.2 Electrochemical dehydrogenative $C(sp^2)$ –H amination method

Straight forward synthesis of novel bioactive chromones involving an oxidative C–H bond amination with azoles are also carried out. Based on the biological data of active analogues, novel derivatives are designed, synthesized and evaluated biologically for their potential bone morphogenetic protein activating abilities. Additional chromones lacking key substituents at C₂ and C₃ positions were also prepared as closely related probes for biological applications. A novel 2,3-disubstituted 4*H*-chromen-4-one induced a pronounced, kinase-independent, negative TGF-β feedback that enhanced nuclear BMP-Smad signaling outputs.

$$R^{1} \xrightarrow{\text{ii}} R^{2} \xrightarrow{\text{azole,}} R^{1} \xrightarrow{\text{ii}} R^{2}$$

$$29 \text{ compounds}$$

$$\text{novel BMP activator chemotype}$$

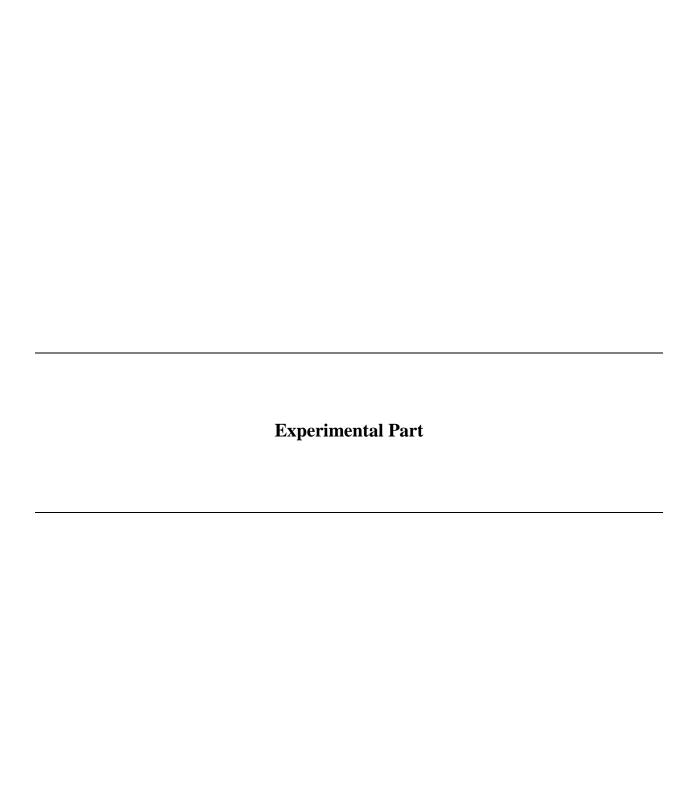
$$CF_{3}$$

$$|C_{50} \approx 0.2 \text{ } \mu\text{M}$$

Scheme 6.3 Design, Synthesis and biological studies of chromone derivatives

In summary, novel metal-free C–H functionalization methods are developed for accessing biologically relevant heteroatoms containing organic molecules. Also, existing C–H functionalization methodologies are employed for the design and synthesis of known bioactive compounds as well as novel potential bioactive compounds.

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7 Experimental Part

7.1 General

Reagents and solvents

Commercially available chemicals were purchased from *Sigma-Aldrich*, *Acros Organics*, *Alfa Aesar*, *Fluorochem*, *VWR Germany*, *ABCR* or *TCI Germany*. Unless otherwise noted, all commercially available compounds and solvents were used as provided without any further purification. Dry dichloromethane was purified by the solvent purification system *M-BRAUN SPS-800*. Solvents for chromatography were laboratory grade.

Chromatography

Analytical thin-layer chromatography (TLC) was performed on Merck silica gel 60 aluminium plates with F₂₅₄ indicator, visualized by irradiation with UV light (254 nm or 365 nm) or staining with KMnO₄-solution (1.5 g KMnO₄, 10 g K₂CO₃, 1.25 mL 10% NaOH in 150 mL H₂O) or staining with *para*-anisaldehyde staining solution (5 mL glacial sulfuric acid, 1.5 mL glacial acetic acid and 3.7 mL *para*-anisaldehyde in 135 mL absolute EtOH).

Column chromatography was performed using silica gel Merck 60 (particle size 0.040-0.063 mm). Solvent mixtures are understood as volume/volume.

Mass spectrometry

Low resolution mass spectra (MS-EI, 70 eV) were collected using an *Agilent Technologies 7890A GC-System* (column: HP-5MS, 30 m × 0.250 mm × 0.25 μm) equipped with an *Agilent Technologies 5975C inert XL MSD with Triple-Axis Detector*. Otherwise Low-resolution mass spectra (MS-ESI) were collected using a *Agilent 1290 LC-MS system, Rapid resolution HD 2.1x50 mm 1.8 μm column*. High resolution mass spectra were recorded on an *LTQ Orbitrap* mass spectrometer coupled to an *Accela HPLC System* (HPLC column: Hypersyl GOLD, 50 mm × 1 mm, 1.9 μm). 122 Experimental |

Nuclear magnetic resonance spectroscopy (NMR)

¹H-NMR, ¹⁹F NMR, ¹³C-NMR and 2D-NMR spectra were recorded on *Bruker DRX400* (400 MHz), *DRX500* (500MHz), *DRX600* (600MHz) and *DRX700* (700 MHZ) spectrometers in

CD₂Cl₂, CDCl₃, or (CD₃)₂SO. Data are reported in the following order: chemical shift (δ) in ppm; multiplicities are indicated s (singlet), d (doublet), dd (doublet of doublets), t (triplet), q (quartet), hept (heptet), m (multiplet); coupling constants (J) are given in Hertz (Hz).

Fourier transform infrared spectroscopy

Fourier transform infrared spectroscopy (FT-IR) spectra were obtained with a *Bruker Tensor 27* spectrometer (ATR, neat) and are reported in terms of frequency of absorption (cm⁻¹).

Electrochemistry

Unless otherwise noted, all reactions and cyclic voltametry (CV) studies were carried out in a 5 mL IKA Electrasyn 2.0 single reaction vial or carousel connected to Electrasyn 2.0. All the electrodes (52 x 5 mm) used for reactions and CV were purchased from IKA.

Biological Studies

All biological experiments were carried out at the Compound Management and Screening Center (COMAS) in Dortmund or at their collaborator's labs.

7.2 Experimental part for the cross-coupling

7.2.1 General procedures

Preparation of heteroaromatic N-oxides

To a solution of quinoline, isoquinoline or pyridine (1 mmol) in dichloromethane (10 mL), 3-chloroperbenzoic acid (1.3 mmol) was added and the reaction was stirred for 24 h at room temperature. Afterwards, residual 3-chloroperbenzoic acid was removed by adding 1 M KOH solution. The aqueous phase was extracted three times with dichloromethane and the combined organic layers were dried over MgSO₄. Column chromatography provided the pure product (eluent: DCM - MeOH). [28]

$$R \xrightarrow{||} N \qquad \frac{m\text{-CPBA (1.3 equiv.)}}{DCM, \text{ rt, 24 h}} \qquad R \xrightarrow{||} R \xrightarrow{||} N$$

Preparation benzyltrimethylsilanes

An oven-dried two neck round-bottom flask containing magnesium powder (0.18 g, 7.5 mmol, 1.5 equiv.) a grain of iodine and dry THF (20 mL) was cooled to 0 °C under nitrogen. Then trimethylsilyl chloride (0.6 mL, 1.2 equiv.) was added, followed by the dropwise addition of a solution of the bromide (5.0 mol, 1.0 equiv.) in THF (5 mL) over a period of 15 min. After the addition was completed, the reaction mixture was stirred overnight and poured 10 mL statured NH₄Cl. The resulting mixture was extracted with EtOAc (3 x 20 mL), the combined organic layers were washed with brine (20 mL), dried MgSO₄ and concentrated to obtain the residue. The residue was purified by flash column chromatography to give the benzyltrimethylsilanes. [91]

General procedures A for the benzylation of heteroaromatic N-oxides

To a solution of *N*-oxide (0.4 mmol) in DMF (1 mL), **2a** (1.20 mmol, 233.15 μ L) was added and the reaction vial was flushed with argon. TBAF (10 - 20 mol%, 40 - 80 μ L) and Et₃SiH (only in

case of isoquinoline and quinoline *N*-oxides) (2 mmol, 329.32 µL) were added respectively, stirred at room temperature and monitored by TLC. After the complete consumption of *N*-oxide, column chromatography of the reaction mixture provided the pure product (Eluent: Petroleum ether-EtOAc).

$$R^{1} \stackrel{\text{II}}{\underset{\stackrel{}{\downarrow}}{\downarrow}} + R^{2} \stackrel{\text{Me}_{3}Si}{\underset{\stackrel{}{\downarrow}}{\downarrow}} + R^{2} \stackrel{\text{TBAF (10 mol\%)}}{\underset{\text{argon}}{\downarrow}} + R^{2} \stackrel{\text{TBAF (10 mol\%)}}{\underset{\text{argon}}{\downarrow}$$

General procedures B for the alkynylation of heteroaromatic N-oxides

To a solution of *N*-oxide (0.2 mmol) in THF (1 mL), ethynyltrimethylsilane (0.8 - 1.2 mmol) was added and the reaction tube was flushed with argon, to which TBAF (5 - 10 mol%) was added using a micropipette (acetylene gas evolution might cause strong effervescence) and stirred at room temperature for specified time. Column chromatography provided the pure product (Eluent: DCM-MeOH).

Procedure for gram-scale C–H bond benzylation of isoquinoline N-oxide

To a solution of isoquinoline *N*-oxide (1g, 6.75 mmol) in DMF (17 mL), **2a** (3.9 mL, 20.25 mmol) was added and the reaction vial was flushed with argon. TBAF (10 mol%, 675 μ L) and Et₃SiH (5.56 mL, 33.76 mmol) were added respectively, stirred at room temperature and monitored by TLC. After the complete consumption of isoquinoline *N*-oxide, column chromatography of the reaction mixture provided the pure product (Eluent: Petroleum ether-EtOAc = 10:1).

7.2.2 Physical data of products

1-Benzylisoquinoline (77a)

Prepared according to general procedure A using isoquinoline *N*-oxide (0.4 mmol) and benzyltrimethylsilane (1.2 mmol). The product was isolated as yellow oil (69 mg, 0.31 mmol, 78%) after 24 h; $R_f = 0.32$ (cyclohexane / ethyl acetate = 5:1); ¹H NMR (400 MHz, CDCl₃) δ 8.49 (d, J = 5.8 Hz, 1H), 8.13 (dd, J = 8.5, 1.1 Hz, 1H), 7.79 (d, J = 8.2 Hz, 1H), 7.63 – 7.59 (m, 1H), 7.57 – 7.47 (m, 2H), 7.29 – 7.20 (m, 4H), 7.18 – 7.11 (m, 1H), 4.67 ppm (s, 2H); ¹³C NMR (126 MHz, CDCl₃) δ 160.1, 141.8, 139.3, 136.6, 129.9, 128.6, 128.5, 127.3, 127.2, 127.2, 126.2, 125.8, 119.9, 41.9 ppm; **FT-IR**: $\tilde{v} = 3027$, 2959, 1673, 1560, 1385, 1153, 1002 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₁₆H₁₄N = 220.1121 found: 220.1120.

2-Benzylquinoline (77b)

Prepared according to general procedure A using quinoline *N*-oxide (0.4 mmol) and benzyltrimethylsilane (1.2 mmol). The product was isolated as yellow oil (69.5 mg, 0.31 mmol, 79%) after 1 h; $R_f = 0.28$ (cyclohexane / ethyl acetate = 5:1); ¹H NMR (500 MHz, CD_2Cl_2) δ 8.09 – 8.04 (m, 2H), 7.80 – 7.77 (m, 1H), 7.73 – 7.71 (m, 1H), 7.50 – 7.53 (m, 1H), 7.36 – 7.21 ppm (m, 6H), 4.33 ppm (s, 2H); ¹³C NMR (126 MHz, CD_2Cl_2) δ 161.6, 148.3, 139.9, 136.7, 129.7, 129.5, 129.3, 128.9, 127.9, 127.1, 126.7, 126.2, 121.9, 45.7 ppm; **FT-IR**: \tilde{v} = 3058, 2848, 2362, 1815, 1597, 1495, 1310, 1029 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{16}H_{14}N$ = 220.1121 found: 220.1119.

2-Benzylpyridine (77c)

Prepared according to general procedure A using pyridine *N*-oxide (0.4 mmol) and benzyltrimethylsilane (1.2 mmol). The product was isolated as yellow oil (51 mg, 0.3 mmol, 76%) after 12 h; $R_f = 0.38$ (cyclohexane / ethyl acetate = 5:1); ${}^{1}H$ NMR (600 MHz, CDCl₃) δ 8.53 – 8.49 (m, 1H), 7.55 – 7.52 (m, 1H), 7.29 – 7.20 (m, 4H), 7.20 – 7.15 (m, 1H), 7.09 – 7.04 (m, 2H), 4.13 ppm (s, 2H); ${}^{13}C$ NMR (151 MHz, CDCl₃) δ 160.8, 149.0, 139.3, 136.5, 129.0, 128.5, 126.3, 123.0, 121.1, 44.5 ppm; **FT-IR**: \tilde{v} = 3061, 2922, 2314, 1879, 1588, 1568, 1309, 1049 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{12}H_{12}N$ = 170.0964 found: 170.0959.

2-Benzyl-6-methylquinoline (77d)

Prepared according to general procedure A using 6-methylquinoline *N*-oxide (0.4 mmol) and benzyltrimethylsilane (1.2 mmol). The product was isolated as yellow solid (68 mg, 0.29 mmol, 73%) after 3 h; $R_f = 0.47$ (cyclohexane / ethyl acetate = 5:1); 1 **H NMR** (500 MHz, CD_2Cl_2) δ 7.99 – 7.92 (m, 2H), 7.55 (d, J = 7.9 Hz, 2H), 7.35 – 7.29 (m, 4H), 7.24 (dd, J = 7.4, 3.2 Hz, 2H), 4.31 (s, 2H), 2.53 ppm (s, 3H); 13 **C NMR** (126 MHz, CD_2Cl_2) δ 160.6, 146.8, 140.1, 136.2, 136.0, 131.9, 129.5, 128.9, 128.8, 127.1, 126.7, 126.7, 121.8, 45.6, 21.6 ppm; **FT-IR**: $\tilde{v} = 3025$, 2512, 1976, 1597, 1433, 1220, 1026 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{17}H_{16}N = 234.1277$ found: 234.1273.

2-Benzyl-6-methoxyquinoline (77e)

Prepared according to general procedure A using 6-methoxyquinoline *N*-oxide (0.4 mmol) and benzyltrimethylsilane (1.2 mmol). The product was isolated as pale orange solid (52 mg, 0.21 mmol, 52%) after 3 h; $R_f = 0.32$ (cyclohexane / ethyl acetate = 5:1); ¹H NMR (500 MHz, CD₂Cl₂) δ 7.97 (d, J = 8.4 Hz, 1H), 7.93 (d, J = 9.2 Hz, 1H), 7.34 (dd, J = 9.2, 2.8 Hz, 1H), 7.33 – 7.27 (m, 4H), 7.25 – 7.19 (m, 2H), 7.08 (d, J = 2.9 Hz, 1H), 4.28 (s, 2H), 3.91 ppm (s, 3H); ¹³C NMR (126 MHz, CD₂Cl₂) δ 159.0, 157.7, 144.2, 140.2, 135.5, 130.6, 129.4, 128.8, 128.0, 126.6, 122.2, 122.1,

105.5, 55.8, 45.4 ppm; **FT-IR**: $\tilde{v} = 3082$, 2962, 2362, 1664, 1493, 1374, 1289, 1071 cm⁻¹; **HR-MS**: calc. for $[M+H]^+ C_{17} H_{16} ON = 250.1226$ found: 250.1223.

2-Benzyl-8-methylquinoline (77f)

Prepared according to general procedure A using 8-methylquinoline *N*-oxide (0.4 mmol) and benzyltrimethylsilane (1.2 mmol). The product was isolated as yellow oil (31 mg, 0.13 mmol, 33%) after 5 h; $R_f = 0.72$ (cyclohexane / ethyl acetate = 5:1); ¹**H NMR** (500 MHz, CD_2Cl_2) δ 8.03 (d, J = 8.4 Hz, 1H), 7.62 (dd, J = 8.1, 1.5 Hz, 1H), 7.56 – 7.54 (m, 1H), 7.40 – 7.33 (m, 3H), 7.32 – 7.24 (m, 3H), 7.23 – 7.19 (m, 1H), 4.32 (s, 2H), 2.80 ppm (t, J = 0.8 Hz, 3H); ¹³**C NMR** (126 MHz, CD_2Cl_2) δ 160.3, 147.2, 140.1, 137.2, 136.9, 129.7, 129.5, 128.8, 128.8, 128.7, 126.9, 126.6, 125.9, 125.8, 125.7, 121.5, 45.9, 18.0 ppm ; **FT-IR**: $\tilde{v} = 3026$, 2850, 1935, 1614, 1466, 1210, 972 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{17}H_{16}N = 234.1277$ found: 234.1275.

2-Benzyl-4-methylquinoline (77g)

Prepared according to general procedure A using 4-methylquinoline *N*-oxide (0.4 mmol) and benzyltrimethylsilane (1.2 mmol). The product was isolated as pale yellow solid (67 mg, 0.28 mmol, 71%) after 1 h; $R_f = 0.4$ (cyclohexane / ethyl acetate = 5:1); ${}^{1}\mathbf{H}$ NMR (500 MHz, CD₂Cl₂) δ 8.07 – 8.03 (m, 1H), 7.97 (dd, J = 8.3, 1.4 Hz, 1H), 7.72 – 7.68 (m, 1H), 7.55 – 7.52 (m, 1H), 7.37 – 7.28 (m, 4H), 7.26 – 7.20 (m, 1H), 7.13 (d, J = 1.1 Hz, 1H), 4.28 (s, 2H), 2.63 ppm (s, 3H); ${}^{13}\mathbf{C}$ NMR (126 MHz, CD₂Cl₂) δ 161.2, 148.1, 145.0, 140.0, 129.8, 129.5, 128.8, 128.8, 127.2, 126.7, 125.9, 124.0, 122.5, 45.7, 18.8 ppm; **FT-IR**: $\tilde{\mathbf{v}} = 3023$, 2521, 1976, 1506, 1341, 1188, 1018 cm⁻¹; **HR-MS**: calc. for [M+H] $^+$ C₁₇H₁₆N = 234.1277 found: 234.1274.

2-Benzyl-4-methylpyridine (77h)

Prepared according to general procedure A using 4-methylpyridine *N*-oxide (0.4 mmol) and benzyltrimethylsilane (1.2 mmol). The product was isolated as yellow oil (54 mg, 0.29 mmol, 73%) after 12 h; $R_f = 0.25$ (cyclohexane / ethyl acetate = 5:1); 1 H NMR (600 MHz, CD_2Cl_2) δ 8.36 (d, J = 5.1 Hz, 1H), 7.33 – 7.24 (m, 4H), 7.24 – 7.15 (m, 1H), 6.98 (d, J = 1.7 Hz, 1H), 6.95 (dd, J = 5.0, 1.6 Hz, 1H), 4.08 ppm (s, 2H), 2.28 ppm (s, 3H); 13 C NMR (151 MHz, CD_2Cl_2) δ 161.1, 149.4, 148.0, 140.5, 129.4, 128.8, 126.5, 124.2, 122.6, 44.8, 21.1 ppm; FT-IR: $\tilde{v} = 3084$, 2922, 2030, 1601, 1400, 1216, 1030 cm⁻¹; HR-MS: calc. for [M+H]⁺ $C_{13}H_{14}N = 184.1121$ found: 184.1117.

1-(4-Isopropylbenzyl)isoquinoline (77i)

Prepared according to general procedure A using isoquinoline *N*-oxide (0.4 mmol) and 4-isopropylbenzyltrimethylsilane (1.2 mmol). The product was isolated as a yellow oil (48 mg, 0.18 mmol, 46%) after 24 h; $R_f = 0.28$ (cyclohexane / ethyl acetate = 5:1); ${}^{1}\mathbf{H}$ NMR (600 MHz, CD₂Cl₂) δ 8.47 (d, J = 5.7 Hz, 1H), 8.22 – 8.20 (m, 1H), 7.85 – 7.83 (m, 1H), 7.67 – 7.65 (m, 1H), 7.58 – 7.55 (m, 2H), 7.21 (d, J = 8.2 Hz, 2H), 7.13 (d, J = 8.1 Hz, 2H), 4.63 (s, 2H), 2.89 – 2.82 (m, 1H), 1.21 ppm (d, J = 7.0 Hz, 6H); ${}^{13}\mathbf{C}$ NMR (151 MHz, CD₂Cl₂) δ 160.8, 147.2, 142.4, 137.3, 136.9, 130.1, 128.9, 127.7, 127.5, 127.4, 126.8, 126.0, 119.9, 41.7, 34.0, 24.1 ppm; **FT-IR**: $\tilde{\mathbf{v}} = 3009$, 2957, 1907, 1585, 1419, 1290, 1153, 1018 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₁₉H₂₀N = 262.1590 found: 262.1587.

1-([1,1'-Biphenyl]-4-ylmethyl)isoquinoline (77j)

Prepared according to general procedure A using isoquinoline *N*-oxide (0.4 mmol) and ([1,1'-biphenyl]-4-ylmethyl)trimethylsilane (1.2 mmol). The product was isolated as a white solid (54 mg, 0.18 mmol, 46%) after 24 h; $R_f = 0.26$ (cyclohexane / ethyl acetate = 5:1); ¹H NMR (700 MHz, CD_2Cl_2) δ 8.50 (d, J = 5.7 Hz, 1H), 8.23 (d, J = 8.5 Hz, 1H), 7.86 (d, J = 8.2 Hz, 1H), 7.69 – 7.66 (m, 1H), 7.62 – 7.55 (m, 4H), 7.54 – 7.50 (m, 2H), 7.42 (t, J = 7.7 Hz, 2H), 7.37 (d, J = 8.0 Hz, 2H), 7.33 (t, J = 7.4 Hz, 1H), 4.72 ppm (s, 2H); ¹³C NMR (176 MHz, CD_2Cl_2) δ 160.5, 142.4, 141.1, 139.4, 139.2, 136.9, 130.2, 129.5, 129.1, 127.7, 127.6, 127.5, 127.4, 127.4, 127.2, 126.0, 120.0, 41.8 ppm; **FT-IR**: $\tilde{v} = 3027$, 2925, 2516, 1975, 1620, 1599, 1238, 1006 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{22}H_{18}N = 296.1434$ found: 296.1429.

1-(Naphthalen-1-ylmethyl) isoquinoline (77k)

Prepared according to general procedure A using isoquinoline *N*-oxide (0.4 mmol) and trimethyl (naphthalen-1-ylmethyl) silane (1.2 mmol). The product was isolated as a pale yellow solid (41 mg, 0.15 mmol, 38%) after 24 h; $R_f = 0.26$ (cyclohexane / ethyl acetate = 5:1); ¹**H NMR** (600 MHz, CD_2Cl_2) δ 8.51 (d, J = 5.7 Hz, 1H), 8.24 – 8.22 (m, 1H), 7.86 – 7.84 (m, 1H), 7.81 – 7.77 (m, 1H), 7.75 (dd, J = 10.8, 8.0 Hz, 2H), 7.70 (d, J = 1.6 Hz, 1H), 7.66 – 7.64 (m, 1H), 7.61 (dd, J = 5.7, 0.9 Hz, 1H), 7.55 – 7.53 (m, 1H), 7.47 – 7.39 (m, 3H), 4.83 ppm (s, 2H); ¹³**C NMR** (151 MHz, CD_2Cl_2) δ 160.4, 142.4, 137.7, 136.9, 133.9, 132.5, 130.2, 128.3, 127.9, 127.8, 127.7, 127.7, 127.5, 127.5, 127.3, 126.3, 126.0, 125.8, 120.1, 42.4 ppm; **FT-IR**: $\tilde{v} = 3016$, 2959, 1976, 1599, 1389, 1238, 1067 cm⁻¹; **HR-MS**: calc. for $[M+H]^+C_{20}H_{16}N = 270.1277$ found: 270.1276.

1-(Naphthalen-2-ylmethyl)isoquinoline (77l)

Prepared according to general procedure A using isoquinoline *N*-oxide (0.4 mmol) and Trimethyl(naphthalen-2-ylmethyl)silane (1.2 mmol). The product was isolated as a yellow oil (50 mg, 0.19 mmol, 47%) after 24 h; $R_f = 0.26$ (cyclohexane / ethyl acetate = 5:1); ¹**H NMR** (700 MHz, CD_2Cl_2) δ 8.45 (d, J = 5.7 Hz, 1H), 8.24 – 8.19 (m, 1H), 8.17 (d, J = 8.5 Hz, 1H), 7.93 – 7.86 (m, 2H), 7.76 (d, J = 8.2 Hz, 1H), 7.68 (dd, J = 8.3, 6.8 Hz, 1H), 7.61 (d, J = 5.7 Hz, 1H), 7.55 – 7.51 (m, 3H), 7.33 (t, J = 7.6 Hz, 1H), 7.07 (d, J = 7.1 Hz, 1H), 5.13 ppm (s, 2H); ¹³**C NMR** (176 MHz, CD_2Cl_2) δ 160.3, 142.4, 136.7, 136.2, 134.2, 132.6, 130.3, 129.0, 127.8, 127.8, 127.6, 127.3, 127.2, 126.4, 126.0, 125.8, 125.8, 124.4, 120.0, 39.2 ppm; **FT-IR**: $\tilde{v} = 3026$, 2925, 1976, 1620, 1498, 1408, 1261, 1015 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{20}H_{16}N = 270.1277$ found: 270.1269.

2-(4-Isopropylbenzyl) quinoline (77m)

Prepared according to general procedure A using quinoline *N*-oxide (0.4 mmol) and 4-isopropylbenzyltrimethylsilane (1.2 mmol). The product was isolated as yellow oil (70 mg, 0.27 mmol, 67%) after 6 h; $R_f = 0.40$ (cyclohexane / ethyl acetate = 5:1); ¹H NMR (500 MHz, CD₂Cl₂) δ 8.10 – 8.03 (m, 2H), 7.80 (dd, J = 8.1, 1.5 Hz, 1H), 7.73 – 7.69 (m, 1H), 7.53 – 7.50 (m, 1H), 7.31 – 7.24 (m, 3H), 7.22 – 7.17 (m, 2H), 4.30 (s, 2H), 2.89 (p, J = 6.9 Hz, 1H), 1.24 ppm (d, J = 6.9 Hz, 6H); ¹³C NMR (126 MHz, CD₂Cl₂) δ 161.8, 148.3, 147.4, 137.2, 136.6, 129.7, 129.4, 129.3, 127.9, 127.1, 126.9, 126.2, 121.9, 45.4, 34.1, 24.2 ppm; FT-IR: \tilde{v} = 3052, 2869, 1794, 1598, 1382, 1197, 1018 cm⁻¹; HR-MS: calc. for [M+H]⁺ C₁₉H₂₀N = 262.1590 found: 262.1587.

2-(4-Methoxybenzyl)quinoline (77n)

Prepared according to general procedure A using quinoline *N*-oxide (0.4 mmol) and 4-methoxybenzyltrimethylsilane (1.2 mmol). The product was isolated as yellow oil (70 mg, 0.28 mmol, 70%) after 12 h; $R_f = 0.32$ (cyclohexane / ethyl acetate = 5:1); ¹H NMR (500 MHz, CD₂Cl₂)

 δ 8.08 – 8.00 (m, 2H), 7.79 (dd, J = 8.2, 1.4 Hz, 1H), 7.71 – 7.68 (m, 1H), 7.51 – 7.49 (m, 1H), 7.28 – 7.20 (m, 3H), 6.87 – 6.82 (m, 2H), 4.25 (s, 2H), 3.76 ppm (s, 3H); ¹³C NMR (126 MHz, CD₂Cl₂) δ 162.0, 158.7, 148.2, 136.6, 131.9, 130.4, 129.6, 129.2, 127.9, 127.1, 126.2, 121.8, 114.2, 55.5, 44.8 ppm; **FT-IR**: \tilde{v} = 3058, 2954, 2489, 1886, 1652, 1464, 1280, 1106 cm⁻¹; **HR-MS**: calc. for [M+H]⁺C₁₇H₁₆ON = 250.1226 found: 250.1224.

2-(4-Chlorobenzyl)quinoline (770)

Prepared according to general procedure A using quinoline *N*-oxide (0.4 mmol) and 4-chlorobenzyltrimethylsilane (1.2 mmol). The product was isolated as yellow oil (86 mg, 0.34 mmol, 84%) after 6 h; $R_f = 0.36$ (cyclohexane / ethyl acetate = 5:1); ¹**H NMR** (500 MHz, CD₂Cl₂) δ 8.08 (d, J = 8.5 Hz, 1H), 8.02 (dd, J = 8.3, 1.1 Hz, 1H), 7.80 (dd, J = 8.1, 1.5 Hz, 1H), 7.72 – 7.69 (m, 1H), 7.53 – 7.50 (m, 1H), 7.30 – 7.22 (m, 5H), 4.29 ppm (s, 2H); ¹³**C NMR** (126 MHz, CD₂Cl₂) δ 161.0, 148.3, 138.5, 136.9, 132.4, 130.9, 129.8, 129.2, 128.9, 127.9, 127.1, 126.4, 121.8, 44.9 ppm; **FT-IR**: $\tilde{v} = 3037$, 2962, 2490, 1818, 1599, 1374, 1220, 1154, 1014 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₁₆H₁₃N³⁵Cl = 254.0731 found: 254.0729; C₁₆H₁₃N³⁷Cl = 256.0701 found: 256.0695.

2-(2,4,6-Trimethylbenzyl)quinoline (77p)

Prepared according to general procedure A using quinoline *N*-oxide (0.4 mmol) and 2, 4, 6-trimethylbenzyltrimethylsilane (1.2 mmol). The product was isolated as white solid (73 mg, 0.28 mmol, 70%) after 6 h; $R_f = 0.44$ (cyclohexane / ethyl acetate = 5:1); ${}^{1}H$ NMR (500 MHz, CD₂Cl₂) δ 7.99 (dd, J = 8.5, 2.5 Hz, 2H), 7.77 (dd, J = 8.2, 1.5 Hz, 1H), 7.67 – 7.70 (m, 1H), 7.47 – 7.50 (m, 1H), 6.98 (d, J = 8.5 Hz, 1H), 6.92 (s, 2H), 4.35 (s, 2H), 2.28 ppm (d, J = 14.0 Hz, 9H); ${}^{13}C$ NMR (126 MHz, CD₂Cl₂) δ 161.4, 148.3, 137.6, 136.6, 136.3, 133.2, 129.6, 129.2, 129.1, 127.8, 127.0, 126.0, 120.5, 39.0, 20.5 ppm; **FT-IR**: $\tilde{v} = 2960$, 2350, 1619, 1503, 1426, 1204, 1016 cm⁻¹; **HR-MS**: calc. for [M+H]⁺C₁₉H₂₀N = 262.1590 found: 262.1587.

2-(4-Isopropylbenzyl) pyridine (77q)

Prepared according to general procedure A using pyridine *N*-oxide (0.4 mmol) and isopropylbenzyltrimethylsilane (1.2 mmol). The product was isolated as yellow oil (25 mg, 0.12 mmol, 30%) after 12 h; $R_f = 0.24$ (cyclohexane / ethylacetate = 5:1); 1 **H NMR** (500 MHz, CD_2Cl_2) δ 8.50 – 8.49 (m, 1H), 7.60 – 7.57 (m, 1H), 7.20 – 7.13 (m, 5H), 7.12 – 7.09 (m, 1H), 4.08 (s, 2H), 2.90 - 2.84 (m, 1H), 1.22 ppm (d, J = 7.0 Hz, 6H); 13 C NMR (126 MHz, CD_2Cl_2) δ 161.6, 149.6, 147.3, 137.5, 136.7, 129.2, 126.8, 123.3, 121.5, 44.5, 34.0, 24.1 ppm; **FT-IR**: $\tilde{v} = 3007$, 2160, 1588, 1420, 1221, 1019 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{15}H_{18}N = 212.1434$ found: 212.1428.

4-Methyl-2-(naphthalen-2-ylmethyl) pyridine (77r)

Prepared according to general procedure A using 4-methylpyridine *N*-oxide (0.4 mmol) and trimethyl (naphthalen-2-ylmethyl) silane (1.2 mmol). The product was isolated as yellow oil (68 mg, 0.29 mmol, 72%) after 24 h; $R_f = 0.16$ (cyclohexane / ethyl acetate = 5:1); ¹H NMR (500 MHz, CD_2Cl_2) δ 8.37 (d, J = 5.0 Hz, 1H), 7.83 – 7.75 (m, 4H), 7.73 – 7.69 (m, 1H), 7.49 – 7.38 (m, 4H), 4.25 (s, 2H), 2.27 ppm (s, 3H); ¹³C NMR (126 MHz, CD_2Cl_2) δ 160.9, 149.3, 137.9, 134.0, 132.5, 128.3, 128.0, 127.9, 127.8, 127.8, 127.6, 126.3, 125.7, 124.3, 122.6, 44.9, 21.0 ppm; FT-IR: $\tilde{v} = 3179$, 2920, 2158, 1668, 1437, 1270, 1017 cm⁻¹; HR-MS: calc. for [M+H]⁺C₁₇H₁₆N = 234.1277 found: 234.1275.

1-(Phenylethynyl)isoquinoline (83)

To a mixture of isoquinoline *N*-oxide (0.2 mmol) in DMF (1 mL), trimethyl(phenylethynyl)silane (0.6 mmol) was added. The reaction vial was flushed with argon and then 10 mol% of TBAF was added and stirred for 2 h. Purification by column chromatography (EtOAc/Petroleum ether) yielded the desired product as a dark brown solid (41 mg, 0.18 mmol, 91%); $R_f = 0.36$ (cyclohexane / ethyl acetate = 5:1); 1 H NMR (500 MHz, CDCl₃) δ 8.56 (d, J = 5.7 Hz, 1H), 8.53 – 8.51 (m, 1H), 7.88 – 7.83 (m, 1H), 7.77 – 7.63 (m, 5H), 7.44 – 7.41 ppm (m, 3H); 13 C NMR (126 MHz, CDCl₃) δ 144.3, 142.8, 136.0, 132.3, 130.9, 129.4, 129.4, 128.6, 128.3, 128.2, 127.1, 127.0, 122.2, 120.8, 94.4, 86.7 ppm; **FT-IR**: \tilde{v} = 3055, 2514, 2159, 2030, 1596, 1353, 1315, 1014 cm⁻¹; **HR-MS**: calc. for [M+H] $^{+}$ C₁₇H₁₂N = 230.0964 found: 230.0961.

1,2-Di(isoquinolin-1-yl)ethyne (85a)

Prepared according to general procedure B using isoquinoline *N*-oxide (0.2 mmol) and ethynyltrimethylsilane (0.8 mmol); The product was obtained as a pale brown solid (25 mg, 0.09 mmol, 89%); $R_f = 0.33$ (dichloromethane / methanol = 50:1); ¹H NMR (400 MHz, CD_2Cl_2) δ 8.65 (dd, J = 9.2, 6.4 Hz, 4H), 7.93 (d, J = 7.7 Hz, 2H), 7.77 ppm (t, J = 6.6 Hz, 6H); ¹³C NMR (101 MHz, CD_2Cl_2) δ 143.8, 143.6, 136.2, 131.1, 130.1, 128.8, 127.4, 127.1, 121.7, 90.6 ppm; **FT-IR**: $\tilde{v} = 2924$, 2853, 2510, 2159, 2030, 1976, 1204 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{20}H_{13}N_2 = 281.1073$ found: 281.1068.

1,2-Di(quinolin-2-yl)ethyne (85b)

Prepared according to general procedure B using quinoline *N*-oxide (0.2 mmol) and ethynyltrimethylsilane (1.2 mmol). The product was obtained as a pale yellow solid (24 mg, 0.09 mmol, 86%); $R_f = 0.4$ (dichloromethane / methanol = 50:1); ¹H NMR (500 MHz, CD_2Cl_2) δ 8.23 (dd, J = 8.5, 0.8 Hz, 2H), 8.11 (dd, J = 8.5, 1.1 Hz, 2H), 7.88 (dd, J = 8.2, 1.4 Hz, 2H), 7.80 – 7.77 (m, 2H), 7.73 (d, J = 8.4 Hz, 2H), 7.63 – 7.60 ppm (m, 2H); ¹³C NMR (126 MHz, CD_2Cl_2) δ 148.3, 142.5, 136.3, 130.1, 129.3, 127.6, 127.5, 127.4, 124.4, 88.5 ppm; **FT-IR**: $\tilde{v} = 3052$, 2521, 2159, 2030, 1976, 1500, 1207 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{20}H_{13}N_2 = 281.1073$ found: 281.1068.

1,2-Bis(6-methylquinolin-2-yl)ethyne (85c)

Prepared according to general procedure B using 6, 7-dimethoxyisoquinoline *N*-oxide (0.2 mmol) and ethynyltrimethylsilane (1.2 mmol). The product was obtained as a pale brown solid (20 mg, 0.07 mmol, 65%); $R_f = 0.62$ (dichloromethane / methanol = 50:1); ¹H NMR (400 MHz, CDCl₃) δ 8.07 (dd, J = 15.0, 8.8 Hz, 4H), 7.74 (d, J = 8.4 Hz, 2H), 7.62 – 7.55 (m, 4H), 2.55 ppm (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 146.8, 141.8, 137.8, 135.8, 132.7, 129.1, 127.6, 126.5, 124.8, 89.0, 21.8 ppm; **FT-IR**: $\tilde{v} = 3059$, 2513, 2159, 2030, 1976, 1491, 1285, 1162 cm⁻¹; **HR-MS**: calc. for $[M+H]^+C_{22}H_{17}N_2 = 309.1386$ found: 309.1386.

1,2-Bis(6-methoxyquinolin-2-yl)ethyne (85d)

Prepared according to general procedure B using 6-methoxyquinoline *N*-oxide (0.2 mmol) and ethynyltrimethylsilane (1.2 mmol). The product was obtained as a brown solid (16 mg, 0.05 mmol, 47%); $R_f = 0.33$ (dichloromethane / methanol = 50:1); ¹**H NMR** (500 MHz, CDCl₃) δ 8.08 (d, J =

8.8 Hz, 4H), 7.75 (d, J = 8.4 Hz, 2H), 7.41 (dd, J = 9.2, 2.8 Hz, 2H), 7.08 (d, J = 2.7 Hz, 2H), 3.95 ppm (s, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 158.8, 144.1, 139.9, 135.4, 130.7, 128.8, 125.2, 123.5, 105.0, 88.9, 55.8 ppm; **FT-IR**: $\tilde{v} = 3002$, 2964, 2518, 2159, 2026, 1976, 1305, 1148 cm⁻¹; **HR-MS**: calc. for [M+H]⁺C₂₂H₁₇O₂N₂ = 341.1284 found: 341.1287.

1,2-Bis(8-methylquinolin-2-yl)ethyne (85e)

Prepared according to general procedure B using 8-methylquinoline *N*-oxide (0.2 mmol) and ethynyltrimethylsilane (1.2 mmol). The product was obtained as a white solid (10 mg, 0.04 mmol, 35%); $R_f = 0.7$ (dichloromethane / methanol = 50:1); ¹H NMR (400 MHz, CD₂Cl₂) δ 8.20 (d, J = 8.4 Hz, 2H), 7.72 (t, J = 7.8 Hz, 4H), 7.64 (d, J = 7.0 Hz, 2H), 7.50 (t, J = 7.6 Hz, 2H), 2.86 ppm (s, 6H); ¹³C NMR (126 MHz, CD₂Cl₂) δ 147.8, 141.8, 137.8, 136.9, 130.6, 127.8, 127.7, 126.0, 124.6, 88.8, 18.2 ppm; **FT-IR**: $\tilde{v} = 3059$, 2952, 2732, 2503, 2159, 1976, 1499, 1303, 1080 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₂₂H₁₇N₂ = 309.1386 found: 309.1381.

1,2-Bis(6,7-dimethoxyisoquinolin-1-yl)ethyne (85f)

Prepared according to general procedure B using 6, 7-dimethoxyisoquinoline *N*-oxide (0.2 mmol) and ethynyltrimethylsilane (1.2 mmol). The product was obtained as pale yellow solid (28 mg, 0.07 mmol, 69%); $R_f = 0.36$ (dichloromethane / methanol = 50:1); ¹H NMR (400 MHz, CDCl₃) δ 8.46 (d, J = 5.5 Hz, 2H), 7.92 (s, 2H), 7.58 (d, J = 5.7 Hz, 2H), 7.11 (s, 2H), 4.11 (s, 6H), 4.05 ppm (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 153.4, 151.2, 141.2, 140.2, 132.8, 126.2, 119.9,

104.9, 104.5, 56.3, 56.0 ppm; **FT-IR**: $\tilde{v} = 2931$, 2518, 2159, 2027, 1976, 1426, 1027 cm⁻¹; **HR-MS**: calc. for $[M+H]^+ C_{24}H_{21}O_4N_2 = 401.1496$ found: 401.1490.

1,2-Bis(7-bromoisoquinolin-1-yl)ethyne (85g)

Prepared according to general procedure B using 7-bromoisoquinoline *N*-oxide (0.2 mmol) and ethynyltrimethylsilane (1.2 mmol). The product was obtained as a pale brown solid (26 mg, 0.06 mmol, 59%); $R_f = 0.33$ (dichloromethane / methanol = 50:1); ¹H NMR (500 MHz, CDCl₃) δ 8.83 (d, J = 1.9 Hz, 2H), 8.68 (d, J = 5.6 Hz, 2H), 7.86 (dd, J = 8.7, 1.9 Hz, 2H), 7.79 (d, J = 8.7 Hz, 2H), 7.74 ppm (d, J = 5.7 Hz, 2H); ¹³C NMR (126 MHz, CDCl₃) δ 142.9, 141.7, 134.7, 134.3, 129.1, 128.6, 122.7, 121.3, 90.3 ppm; FT-IR: $\tilde{v} = 2918$, 2495, 2159, 2031, 1976, 1541, 1373, 1012 cm⁻¹; HR-MS: calc. for [M+H]⁺ C₂₀H₁₁N₂⁷⁹Br₂ = 436.9283 found: 436.9282 C₂₀H₁₁N₂⁷⁹Br⁸¹Br = 438.9263 found: 438.9258 C₂₀H₁₁N₂⁸¹Br₂ = 440.9243 found: 440.9236.

1, 2-Bis (7-phenylisoquinolin-1-yl) ethyne (85h)

Prepared according to general procedure B using 7-phenylisoquinoline 2-oxide (0.2 mmol) and ethynyltrimethylsilane (1.2 mmol). The product was obtained as a pale brown solid (17 mg, 0.04 mmol, 39%); $R_f = 0.64$ (dichloromethane / methanol = 50:1); ${}^{1}H$ NMR (500 MHz, CDCl₃) δ 8.87 – 8.83 (m, 2H), 8.65 (d, J = 5.6 Hz, 2H), 8.03 (dd, J = 8.5, 1.8 Hz, 2H), 7.97 (d, J = 8.5 Hz, 2H), 7.77 (d, J = 5.6 Hz, 2H), 7.76 – 7.71 (m, 4H), 7.45 – 7.39 (m, 4H), 7.39 – 7.34 ppm (m, 2H); ${}^{13}C$ NMR (126 MHz, CDCl₃) δ 143.5, 142.9, 141.9, 140.2, 135.5, 131.3, 130.4, 129.4, 128.4, 128.0,

124.9, 121.7, 91.3 ppm; **FT-IR**: $\tilde{v} = 2956$, 2514, 2361, 2159, 2096, 1670, 1487, 1144 cm⁻¹; **HR-MS**: calc. for [M+H] $^+$ C₃₂ H₂₁ N₂ = 433.1699 found: 433.1699.

7.3 Experimental part for the electrochemical amination

7.3.1 General procedures

General procedures for the preparation of biaryl acetanilides

To a solution of bromoacetanilide (0.25 mmol) in MeCN (10 mL) and H₂O (0.5 mL), borate (0.28 mmol), K₂CO₃ (0.38 mmol) and Pd(PPh₃)₄ (0.025 mmol) were added and the reaction tube was flushed with argon. Degassing was done 3 times and stirred at 100 °C overnight. After the completion, reaction mixture was diluted with water, extracted with EtOAC, dried over anhydrous Na₂SO₄. The organic extracts were filtered and concentrated under reduced pressure. Purification by column-chromatography on silica gel afforded the pure product (Eluent: Petroleum ether-EtOAc). [116]

Anilides (1 mmol) were taken in TFA (2 mL). Then Pd(OAc)₂ (0.1 mmol) followed by AgOAc (2 mmol) were added to it. The reaction was heated at 110 °C for the maximum consumption of starting material. Then the reaction was quenched with aqueous sodium bicarbonate solution and extracted with EtOAc, washed with water, brine dried and purified by column chromatography (Eluent: Petroleum ether-EtOAc).^[129]

General procedure A for intramolecular C-H amination

A 5 mL glass vial was charged with 2-amidobiaryl substrate (0.2 mmol, 1 equiv.) to which 1,1,1,3,3,3-hexfluoroisopropanol (0.06 M, 3 mL) followed by sodium ethoxide (21 wt. % solution in ethanol, 0.15 mmol, 0.75 equiv., 56 μ L) were added. Lid with graphite electrodes (52 x 5 mm)

was attached and electrolysis was carried out with a constant current of 3 mA until 2.8 F/mol charge was passed (~ 5 h) at room temperature. After the reaction, the reaction mixture was transferred to a 50 mL round bottom flask to which electrodes were washed with methanol. Solvents were evaporated under reduced pressure and the crude product was purified by silica gel column chromatography to obtain pure product (eluent: petroleum ether- ethyl acetate).

graphite graphite
$$I = 3 \text{ mA}, Q = 2.8 \text{ F/mol}$$
NAOEt (0.75 equiv.)

HFIP (0.06 M), rt

General procedure B for intermolecular C-H amination

A 5 mL glass vial was charged with corresponding *N*-arylamide (0.2 mmol, 1 equiv.) and (hetero)arene partner (2-5 equiv.) to which 1,1,1,3,3,3-hexfluoroisopropanol (0.06 M, 3 mL) followed by potassium *tert*-butoxide (0.4 mmol, 2 equiv., 45.3 mg) were added. Lid with graphite electrodes (52 x 5 mm) was attached and electrolysis was carried out with a constant current of 3 mA until 2.8 F/mol charge was passed (~ 5 h) at room temperature. After the reaction, the reaction mixture was transferred to a 50 mL round bottom flask to which electrodes were washed with methanol. Solvents were evaporated under reduced pressure and the crude product was purified by silica gel column chromatography to obtain pure product (eluent: *n*-pentane- acetone or petroleum ether- ethyl acetate).

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 $I = 3 \text{ mA}, Q = 2.8 \text{ F/mol}$
 $I = 3 \text{ mA$

General procedure for cyclovoltammetric (CV) studies

A 5 mL glass vial was charged with substrate (1 mM) and tetrabutylammonium tetrafluoroborate (0.1 M) in solvent (3 mL). Lid equipped with glassy carbon anode, reference electrode (Ag/AgCl

in 3 M aqueous KCl) and platinum plated cathode was attached to the vial and voltammograms were recorded with scan rates as mentioned.

7.3.2 Electrolytic parameters of reaction

Constant current, I = 3 mA

Charge passed, Q = 2.8 F/mol

Time, t for 0.2 mmol scale reaction = 5 h

Electrode materials for both WE and CE = Graphite electrodes

Maximum concentration of *in situ* generated electrolyte = 0.05 M for intramolecular aminations

= 0.13 M for intermolecular aminations

Voltage range of reactions for 3 mA CCE = 8 - 19 V for intramolecular aminations using NaOEt

= 3 - 9 V for intermolecular aminations using KOtBu

Charge (coulomb, C) = current (ampere, A) \times time (second, s) = $0.003 \times 18000 = 54$ C

Faradaic efficiency range calculated = 26 - 55 %

$$Faradaic\ efficiency = \frac{Q\ theoretical}{Q\ experimental} \times reaction\ yield\ \%$$

Exposed surface area of electrodes calculated using A = 2 (wl+hl+hw), where l is exposed length (5 mm), h is exposed height (2 mm) and w is exposed width (16 mm). = 244 mm²

Current density = area of junction/current value = $244/3 = 0.012 \text{ mA/mm}^2$

7.3.3 Evidences for nitrenium ion intermediate

The HFIP mono adduct was isolated and characterized from a reaction in the absence of any coupling partner. The formation of 103' indicates a delocalization of positive charge of nitrenium ion in the aromatic ring. The HFIP di adduct was observed in GC-MS spectra for a reaction in the absence of any coupling partner. This indicates a delocalization of the positive charge of the nitrenium ion in the aromatic ring.

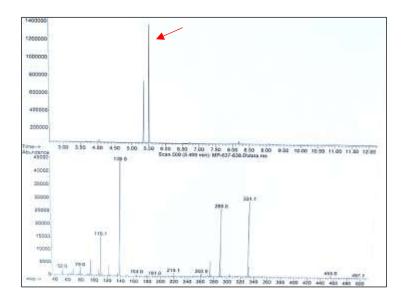


Figure S4. GC-MS-FID spectra for HFIP mono-adduct of 103.

Competitive formation of the HFIP mono adduct was observed in GC-MS spectra for many intramolecular reactions as the major side product. Attempts to minimize this side product were mainly not successful. The formation of **88a'** indicates a delocalization of positive charge of the nitrenium ion in the aromatic ring.

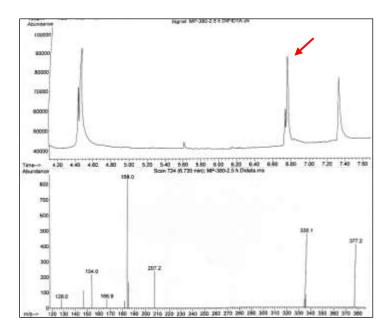


Figure S5. GC-MS-FID spectra for HFIP mono-adduct of 88a.

7.3.4 Physical data of products

1-(9*H***-carbazol-9-yl)ethan-1-one (89a)**

Prepared according to general procedure A; The product was isolated as white solid (26 mg, 0.13 mmol, 63%) after electrolysis; $R_f = 0.53$ (cyclohexane / ethyl acetate = 4:1); 1 **H NMR** (500 MHz, CD₂Cl₂) δ 8.21 (d, J = 8.4 Hz, 2H), 8.02 (d, J = 7.7 Hz, 2H), 7.49 (t, J = 7.8 Hz, 2H), 7.40 (t, J = 7.5 Hz, 2H), 2.86 ppm (s, 3H); 13 **C NMR** (126 MHz, CD₂Cl₂) δ 170.6, 139.2, 127.8, 126.8, 124.1, 120.3, 116.7, 28.1 ppm; **FT-IR**: $\tilde{v} = 2924$, 2531, 1977, 1678, 1430, 1295, 1236, 1151, 1122, 942 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₁₄H₁₂ON = 210.0913 found: 210.0913.

9*H*-carbazole-9-carbaldehyde (89b)

Prepared according to general procedure A; The product was isolated as white solid (23 mg, 0.12 mmol, 58%) after electrolysis; $R_f = 0.57$ (cyclohexane / ethyl acetate = 4:1); 1 **H NMR** (700 MHz, CDCl₃) δ 9.69 (s, 1H), 8.58 (s, 1H), 7.98 (dd, J = 7.2, 3.6 Hz, 2H), 7.71 (s, 1H), 7.53 – 7.45 (m, 2H), 7.43 ppm (t, J = 7.4 Hz, 2H); 13 C NMR (176 MHz, CDCl₃) δ 157.6, 137.9, 137.3, 127.9, 127.2, 126.3, 124.8, 124.5, 120.9, 120.0, 117.0, 110.1 ppm; **FT-IR**: $\tilde{v} = 2927$, 2514, 1977, 1673, 1589, 1361, 1302, 1216, 1081, 946 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₁₃H₁₀ON = 196.0757 found: 196.0759.

1-(2-fluoro-9*H*-carbazol-9-yl)ethan-1-one (89c)

Prepared according to general procedure A; The product was isolated as pale yellow solid (24 mg, 0.11 mmol, 53%) after electrolysis; $R_f = 0.47$ (cyclohexane / ethyl acetate = 4:1); ¹H NMR (500 MHz, CDCl₃) δ 8.11 – 8.03 (m, 2H), 7.93 (dd, J = 7.7, 0.8 Hz, 1H), 7.89 (dd, J = 8.5, 5.6 Hz, 1H), 7.49 – 7.42 (m, 1H), 7.42 – 7.35 (m, 1H), 7.16 – 7.08 (m, 1H), 2.86 ppm (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 170.1, 162.5 (d, J = 243.0 Hz), 139.4 (d, J = 12.4 Hz), 138.8, 127.0, 126.1, 124.0, 122.5, 120.5 (d, J = 10.1 Hz), 119.8, 115.9, 111.7 (d, J = 24.0 Hz), 104.6 (d, J = 29.4 Hz), 27.7 ppm; FT-IR: $\tilde{v} = 3049$, 2159, 1977, 1692, 1382, 1365, 1131, 1062, 1022, 942 cm⁻¹; HR-MS: calc. for [M+H]⁺ $C_{14}H_{11}$ ONF = 228.0819 found: 228.0820.

1-(2-chloro-9*H*-carbazol-9-yl)ethan-1-one (89d)

Prepared according to general procedure A; The product was isolated as pale yellow solid (35 mg, 0.14 mmol, 72%) after electrolysis; $R_f = 0.50$ (cyclohexane / ethyl acetate = 4:1); 1 H NMR (500 MHz, CDCl₃) δ 8.32 (s, 1H), 8.06 (d, J = 8.4 Hz, 1H), 7.93 (d, J = 7.7 Hz, 1H), 7.84 (d, J = 8.2 Hz, 1H), 7.51 – 7.44 (m, 1H), 7.40 – 7.33 (m, 2H), 2.85 ppm (s, 3H); 13 C NMR (126 MHz, CDCl₃) δ 170.0, 139.3, 138.7, 133.2, 127.6, 125.9, 124.9, 124.3, 124.0, 120.5, 120.1, 117.1, 116.0, 27.8 ppm; **FT-IR**: $\tilde{v} = 3142, 2159, 1689, 1595, 1468, 1451, 1364, 1271, 1187, 913 cm⁻¹;$ **HR-MS** $: calc. for [M+H]⁺ <math>C_{14}H_{11}O$ N³⁵Cl = 244.0524 found: 244.0525 $C_{14}H_{11}O$ N³⁷Cl = 246.0494 found: 246.0495.

1-(2-bromo-9*H*-carbazol-9-yl)ethan-1-one (89e)

Prepared according to general procedure A; The product was isolated as pale yellow solid (40 mg, 0.14 mmol, 69%) after electrolysis; $R_f = 0.63$ (cyclohexane / ethyl acetate = 4:1); 1 **H NMR** (500 MHz, CDCl₃) δ 8.49 (s, 1H), 8.06 (d, J = 8.5 Hz, 1H), 7.94 (d, J = 7.7 Hz, 1H), 7.83 – 7.77 (m, 1H), 7.53 – 7.46 (m, 2H), 7.39 (t, J = 7.5 Hz, 1H), 2.85 (s, 3H) ppm; 13 **C NMR** (126 MHz, CDCl₃) δ 170.0, 139.5, 138.5, 127.8, 127.1, 125.8, 125.2, 124.0, 121.1, 120.8, 120.1, 119.9, 116.0, 27.8

ppm; **FT-IR**: $\tilde{v} = 3132$, 2924, 2526, 1976, 1594, 1409, 1325, 1274, 1034, 906 cm⁻¹; **HR-MS**: calc. for $[M+H]^+$ $C_{14}H_{11}ON^{79}Br = 288.0018$ found: 288.0023 $C_{14}H_{11}ON^{81}Br = 289.9998$ found: 290.0002.

1-(2-(tert-butyl)-9H-carbazol-9-yl)ethan-1-one (89f)

Prepared according to general procedure A; The product was isolated as white solid (39 mg, 0.15 mmol, 74%) after electrolysis; $R_f = 0.59$ (cyclohexane / ethyl acetate = 4:1); ¹**H NMR** (500 MHz, CD₂Cl₂) δ 8.30 (s, 1H), 8.18 (d, J = 8.4 Hz, 1H), 7.98 (dd, J = 7.7, 1.4 Hz, 1H), 7.93 (d, J = 8.1 Hz, 1H), 7.51 – 7.42 (m, 2H), 7.41 – 7.34 (m, 1H), 2.87 (s, 3H), 1.44 ppm (s, 9H); ¹³**C NMR** (126 MHz, CD₂Cl₂) δ 170.7, 151.6, 139.6, 139.4, 127.3, 126.8, 124.3, 124.0, 121.9, 120.1, 119.7, 116.7, 113.6, 35.9, 32.0, 28.2 ppm; **FT-IR**: $\tilde{v} = 2961$, 2159, 1691, 1601, 1497, 1460, 1325, 1206, 1018, 860 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₁₄H₁₂ON = 266.1539 found: 266.1541.

1-(2-phenyl-9*H*-carbazol-9-yl)ethan-1-one (89g)

Prepared according to general procedure A; The product was isolated as pale yellow solid (25 mg, 0.09 mmol, 44%) after electrolysis; $R_f = 0.47$ (cyclohexane / ethyl acetate = 4:1); ¹H NMR (500 MHz, CDCl₃) δ 8.51 (d, J = 1.5 Hz, 1H), 8.19 (d, J = 8.4 Hz, 1H), 8.06 – 8.01 (m, 2H), 7.72 – 7.69 (m, 2H), 7.64 (dd, J = 8.0, 1.5 Hz, 1H), 7.52 – 7.47 (m, 3H), 7.43 – 7.37 (m, 2H), 2.93 ppm (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 170.3, 141.6, 141.0, 139.4, 139.1, 129.0, 127.7, 127.6, 127.4, 126.4, 125.6, 123.9, 123.3, 120.1, 120.1, 116.3, 115.3, 28.0 ppm; FT-IR: $\tilde{v} = 2924$, 2159, 1977, 1695, 1459, 1448, 1306, 1265, 1013, 913 cm⁻¹; HR-MS: calc. for [M+H]⁺ C₂₀H₁₆ON = 286.1226 found: 286.1229.

1-(2-ethyl-9*H*-carbazol-9-yl)ethan-1-one (89h)

Prepared according to general procedure A using a (0.2 mmol); The product was isolated as pale yellow solid (21 mg, 0.14 mmol, 44%) after electrolysis; $R_f = 0.53$ (cyclohexane / ethyl acetate = 4:1); ¹H NMR (500 MHz, CDCl₃) δ 8.15 (d, J = 8.4 Hz, 1H), 8.09 (s, 1H), 7.95 (d, J = 7.2 Hz, 1H), 7.89 (d, J = 7.9 Hz, 1H), 7.48 – 7.41 (m, 1H), 7.40 – 7.33 (m, 1H), 7.24 (d, J = 7.9 Hz, 1H), 2.88 (s, 3H), 2.84 (q, J = 7.6 Hz, 2H), 1.34 ppm (t, J = 7.6 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 170.3, 144.3, 139.2, 138.7, 126.9, 126.7, 124.4, 123.9, 123.7, 119.7, 119.7, 116.2, 115.8, 29.9, 28.0, 16.4 ppm; **FT-IR**: $\tilde{v} = 2965$, 2866, 2116, 1621, 1421, 1325, 1292, 1192, 1057, 922 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₁₆H₁₆ON = 238.1226 found: 238.1228.

1-(3-chloro-9*H*-carbazol-9-yl)ethan-1-one (89i)

Prepared according to general procedure A; The product was isolated as yellow solid (18 mg, 0.07 mmol, 37%) after electrolysis; $R_f = 0.50$ (cyclohexane / ethyl acetate = 4:1); 1 **H NMR** (400 MHz, CDCl₃) δ 8.22 (d, J = 8.9 Hz, 1H), 8.08 (d, J = 8.5 Hz, 1H), 7.94 – 7.90 (m, 2H), 7.52 – 7.48 (m, 1H), 7.42 – 7.37 (m, 2H), 2.85 ppm (s, 3H); 13 C NMR (101 MHz, CDCl₃) δ 169.9, 139.0, 137.2, 129.5, 128.1, 127.8, 127.4, 125.5, 124.0, 120.3, 119.7, 117.8, 116.1, 27.8 ppm; **FT-IR**: $\tilde{v} = 3060$, 2933, 2510, 1616, 1582, 1484, 1329, 1128, 1022, 982 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{14}H_{11}ON^{35}Cl = 244.0524$ found: 244.0525 $C_{14}H_{11}ON^{37}Cl = 246.0494$ found: 246.0495.

1-(4-methyl-9*H*-carbazol-9-yl)ethan-1-one (89j)

Prepared according to general procedure A; The product was isolated as pale white solid (16 mg, 0.07 mmol, 36%) after electrolysis; $R_f = 0.57$ (cyclohexane / ethyl acetate = 4:1); ¹H NMR (400 MHz, CD₂Cl₂) δ 8.29 (d, J = 8.5 Hz, 1H), 8.16 (d, J = 7.7 Hz, 1H), 8.07 (d, J = 8.5 Hz, 1H), 7.54 - 7.45 (m, 1H), 7.46 - 7.34 (m, 2H), 7.19 (d, J = 7.4 Hz, 1H), 2.87 (s, 3H), 2.84 ppm (s, 3H); ¹³C NMR (101 MHz, CD₂Cl₂) δ 170.8, 139.4, 139.3, 133.8, 127.6, 127.3, 127.1, 126.0, 125.0, 124.0, 123.1, 116.5, 114.1, 28.4, 21.5 ppm; FT-IR: $\tilde{v} = 2923$, 2515, 1479, 1456, 1448, 1434, 1416, 1313, 1247, 983 cm⁻¹; HR-MS: calc. for [M+H]⁺ C₁₅H₁₄ON = 224.1070 found: 224.1070.

1-(isopropyl-9*H*-carbazol-9-yl)ethan-1-one (89k) and 1-(1-isopropyl-9*H*-carbazol-9-yl)ethan-1-one (89k')

Prepared according to general procedure A; Combined yield of regioisomers = 46% (23 mg, 0.09 mmol, ratio of isolated isomers is 2.3:1); Pale yellow oils; $R_{f(2i)} = 0.53$ (cyclohexane / ethyl acetate = 4:1); ${}^{1}\mathbf{H}$ NMR (500 MHz, CDCl₃) δ 8.24 (d, J = 8.4 Hz, 1H), 8.09 (d, J = 8.6 Hz, 1H), 8.00 (dd, J = 7.6, 0.7 Hz, 1H), 7.85 (d, J = 1.8 Hz, 1H), 7.49 – 7.44 (m, 1H), 7.41 – 7.34 (m, 2H), 3.09 (hept, J = 7.2 Hz, 1H), 2.87 (s, 3H), 1.36 ppm (d, J = 6.9 Hz, 6H); ${}^{13}\mathbf{C}$ NMR (126 MHz, CDCl₃) δ 170.1, 144.7, 139.1, 137.1, 127.3, 126.7, 126.2, 123.7, 119.8, 117.4, 116.5, 116.1, 34.1, 27.8, 24.5 ppm. FT-IR: $\tilde{\mathbf{v}} = 3071$, 2961, 2851, 2666, 1452, 1368, 1240, 1026, 982 cm⁻¹; HR-MS: calc. for [M+H]⁺ $C_{17}H_{18}$ ON = 252.1383 found: 252.1385; $R_{f(2i^*)} = 0.69$ (cyclohexane / ethyl acetate = 4:1); ${}^{1}\mathbf{H}$ NMR (500 MHz, CDCl₃) δ 8.00 – 7.92 (m, 2H), 7.82 (dd, J = 7.0, 1.8 Hz, 1H), 7.49 – 7.34 (m, 4H), 3.30 (hept, J = 6.8 Hz, 1H), 2.67 (s, 3H), 1.32 ppm (d, J = 6.8 Hz, 6H); ${}^{13}\mathbf{C}$ NMR (126 MHz, CDCl₃) δ 172.3, 140.3, 137.8, 137.7, 128.2, 127.2, 126.9, 125.2, 125.0, 123.6, 120.0, 117.5, 114.8, 30.5, 26.8, 23.8 ppm.

1-(2-(tert-butyl)-9*H*-carbazol-9-yl)ethan-1-one (89l)

Prepared according to general procedure A; The product was isolated as white solid (41 mg, 0.16 mmol, 77%) after electrolysis; $R_f = 0.59$ (cyclohexane / ethyl acetate = 4:1); ${}^{1}\mathbf{H}$ NMR (500 MHz, CD₂Cl₂) δ 8.30 (s, 1H), 8.18 (d, J = 8.4 Hz, 1H), 7.98 (dd, J = 7.7, 1.4 Hz, 1H), 7.93 (d, J = 8.1 Hz, 1H), 7.51 – 7.42 (m, 2H), 7.41 – 7.34 (m, 1H), 2.87 (s, 3H), 1.44 ppm (s, 9H); ${}^{13}\mathbf{C}$ NMR (126 MHz, CD₂Cl₂) δ 170.7, 151.6, 139.6, 139.4, 127.3, 126.8, 124.3, 124.0, 121.9, 120.1, 119.7, 116.7, 113.6, 35.9, 32.0, 28.2 ppm; **FT-IR**: $\tilde{\mathbf{v}} = 2961$, 2159, 1691, 1601, 1497, 1460, 1325, 1206, 1018, 860 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₁₄H₁₂ON = 266.1539 found: 266.1541. All physical data are also given above (2g).

1-(2-phenoxy-9*H*-carbazol-9-yl)ethan-1-one (89m)

Prepared according to general procedure A; The product was isolated as white solid (31 mg, 0.10 mmol, 51%) after electrolysis; $R_f = 0.57$ (cyclohexane / ethyl acetate = 4:1); ${}^{1}\mathbf{H}$ NMR (500 MHz, CD_2Cl_2) δ 8.12 (d, J = 8.3 Hz, 1H), 8.02 - 7.93 (m, 3H), 7.51 - 7.42 (m, 1H), 7.42 - 7.35 (m, 3H), 7.19 - 7.11 (m, 1H), 7.11 - 7.04 (m, 3H), 2.80 ppm (s, 3H); ${}^{13}\mathbf{C}$ NMR (126 MHz, CD_2Cl_2) δ 170.6, 158.1, 157.4, 140.3, 139.4, 130.4, 127.1, 126.6, 124.2, 123.9, 122.5, 121.0, 120.0, 119.2, 116.4, 115.8, 108.2, 28.1 ppm; **FT-IR**: $\tilde{\mathbf{v}} = 2925$, 2364, 2031, 1704, 1584, 1420, 1369, 1297, 1036, 827 cm⁻¹; **HR-MS**: calc. for $[\mathbf{M}+\mathbf{H}]^+$ $C_{20}H_{16}O_2\mathbf{N} = 302.1176$ found: 302.1178.

1-(2-methyl-9*H*-carbazol-9-yl)ethan-1-one (89n)

Prepared according to general procedure A; The product was isolated as white solid (20 mg, 0.09 mmol, 45%) after electrolysis; $R_f = 0.47$ (cyclohexane / ethyl acetate = 4:1); ${}^{1}H$ NMR (500 MHz, CD₂Cl₂) δ 8.18 (d, J = 8.4 Hz, 1H), 8.05 (s, 1H), 7.99 – 7.95 (m, 1H), 7.88 (d, J = 7.8 Hz, 1H), 7.48 – 7.41 (m, 1H), 7.41 – 7.34 (m, 1H), 7.24 – 7.21 (m, 1H), 2.85 (s, 3H), 2.54 ppm (s, 3H); ${}^{13}C$

NMR (126 MHz, CD₂Cl₂) δ 170.7, 139.6, 139.2, 138.2, 127.2, 126.9, 125.3, 124.4, 124.0, 120.0, 119.9, 117.1, 116.7, 28.2, 22.6 ppm; **FT-IR**: $\tilde{\mathbf{v}} = 2918, 2519, 2029, 1977, 1600, 1459, 1369, 1196, 982, 808 cm⁻¹;$ **HR-MS**: calc. for [M+H]⁺ C₁₅H₁₄ON = 224.1070 found: 224.1070.

1-(2-methoxy-9*H*-carbazol-9-yl)ethan-1-one (89o)

Prepared according to general procedure A; The product was isolated as pale yellow solid (22 mg, 0.09 mmol, 46%) after electrolysis; $R_f = 0.43$ (cyclohexane / ethyl acetate = 4:1); ¹H NMR (400 MHz, CDCl₃) δ 8.03 (d, J = 7.7 Hz, 1H), 7.93 - 7.86 (m, 2H), 7.83 (d, J = 8.5 Hz, 1H), 7.43 - 7.32 (m, 2H), 6.97 (dd, J = 8.5, 2.3 Hz, 1H), 3.92 (s, 3H), 2.85 ppm (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 170.3, 159.9, 140.2, 138.5, 126.8, 126.0, 123.8, 120.3, 119.8, 119.3, 115.9, 111.5, 102.1, 55.9, 27.8 ppm; **FT-IR**: $\tilde{\mathbf{v}} = 3150$, 2182, 1597, 1498, 1372, 1278, 1201, 1163, 1025, 945 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₁₅H₁₄O₂N = 240.1019 found: 240.1020.

1-(2-isopropyl-9*H*-carbazol-9-yl)ethan-1-one (89p)

Prepared according to general procedure A; The product was isolated as pale yellow solid (33 mg, 0.13 mmol, 66%) after electrolysis; $R_f = 0.59$ (cyclohexane / ethyl acetate = 4:1); ¹H NMR (500 MHz, CDCl₃) δ 8.15 (d, J = 7.9 Hz, 2H), 7.96 (d, J = 7.5 Hz, 1H), 7.90 (d, J = 7.9 Hz, 1H), 7.47 – 7.42 (m, 1H), 7.39 – 7.34 (m, 1H), 7.29 (dd, J = 7.9, 1.4 Hz, 1H), 3.10 (hept, J = 6.9 Hz, 1H), 2.89 (s, 3H), 1.36 ppm (d, J = 6.9 Hz, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 170.3, 149.1, 139.2, 138.8, 126.9, 126.7, 124.5, 123.7, 122.5, 119.7, 119.7, 116.2, 114.5, 35.1, 28.0, 24.5 ppm; **FT-IR**: $\tilde{v} = 2958$, 2867, 2160, 1695, 1496, 1422, 1284, 1241, 1104, 936 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{17}H_{18}ON = 252.1383$ found: 252.1385.

1-(7*H*-benzo[*c*]carbazol-7-yl)ethan-1-one (89q)

Prepared according to general procedure A; The product was isolated as white solid (33 mg, 0.13 mmol, 64%) after electrolysis; $R_f = 0.35$ (cyclohexane / ethyl acetate = 4:1); 1 **H NMR** (400 MHz, CD₂Cl₂) δ 8.82 (d, J = 8.5 Hz, 1H), 8.63 – 8.55 (m, 1H), 8.48 (d, J = 9.2 Hz, 1H), 8.33 – 8.25 (m, 1H), 8.03 (d, J = 8.2 Hz, 1H), 7.95 (d, J = 9.1 Hz, 1H), 7.79 – 7.70 (m, 1H), 7.62 – 7.48 (m, 3H), 2.95 ppm (s, 3H); 13 C NMR (101 MHz, CD₂Cl₂) δ 171.0, 138.7, 137.5, 131.3, 129.5, 129.2, 128.8, 127.9, 127.3, 126.6, 125.3, 124.3, 124.2, 122.7, 119.7, 116.5, 116.4, 28.6 ppm; **FT-IR**: $\tilde{v} = 3049$, 2159, 1977, 1692, 1382, 1365, 1131, 1062, 1022, 942 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₁₈H₁₄ON = 260.1070 found: 260.1072.

1-(2-(*tert*-butyl)-7-fluoro-9*H*-carbazol-9-yl)ethan-1-one (89r)

Prepared according to general procedure A; The product was isolated as pale yellow solid (39 mg, 0.14 mmol, 69%) after electrolysis; $R_f = 0.67$ (cyclohexane / ethyl acetate = 4:1); ¹H NMR (500 MHz, CD_2Cl_2) δ 8.12 (d, J = 1.6 Hz, 1H), 8.04 (dd, J = 11.1, 2.4 Hz, 1H), 7.94 – 7.83 (m, 2H), 7.52 – 7.44 (m, 1H), 7.18 – 7.06 (m, 1H), 2.85 (s, 3H), 1.43 ppm (s, 9H); ¹³C NMR (126 MHz, CD_2Cl_2) δ 170.6, 162.6 (d, J = 241.5 Hz), 151.2, 140.1 (d, J = 12.4 Hz), 139.7, 123.9, 123.0, 122., 120.7 (d, J = 10.1 Hz), 119.6, 113.2, 111.7 (d, J = 23.9 Hz), 104.8 (d, J = 29.4 Hz), 35.8, 32.0, 28.1 ppm; **FT-IR**: $\tilde{v} = 2952$, 2865, 1996, 1498, 1367, 1264, 1020, 1009, 952 cm⁻¹; **HR-MS**: calc. for $[M+H]^+$ $C_{18}H_{19}$ ONF = 284.1445 found: 284.1449.

Note: Compounds 110a-110p exist as rotamers because of the restricted rotation around C-N bond. Major rotamer peaks are reported except for 110m.

N-(4-methoxyphenyl)-N-(4-methylnaphthalen-1-yl)acetamide (110a)

Prepared according to general procedure B using *N*-(4-methoxyphenyl)acetamide (**108a**, 0.2 mmol, 1 equiv.) and 1-methylnaphthalene (**109a**, 0.4 mmol, 2 equiv.); The product was isolated as pale yellow oil (56.1 mg, 0.18 mmol, 92%) after electrolysis; $R_f = 0.22$ (pentane / acetone = 4:1); ¹H NMR (500 MHz, CD₂Cl₂) δ 8.12 – 8.00 (m, 2H), 7.63 – 7.54 (m, 2H), 7.45 – 7.27 (m, 4H), 6.84 (dd, J = 34.0, 8.6 Hz, 2H), 3.73 (s, 3H), 2.73 (s, 3H), 1.88 ppm (s, 3H); major rotamer ¹³C NMR (126 MHz, CD₂Cl₂) δ 171.4, 157.8, 138.5, 136.2, 134.4, 131.2, 129.0, 127.6, 127.4, 127.2, 127.0, 126.9, 125.5, 123.9, 114.2, 56.0, 23.7, 19.8 ppm; **FT-IR**: $\tilde{v} = 3001$, 2836, 2121, 1710, 1667, 1507, 1440, 1421, 1320, 1243, 986 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₂₀H₂₀O₂N = 306.1489 found: 306.1490.

N-(4-ethoxyphenyl)-N-(4-methylnaphthalen-1-yl)acetamide (110b)

Prepared according to general procedure B using *N*-(4-ethoxyphenyl)acetamide (**108b**, 0.2 mmol, 1 equiv.) and 1-methylnaphthalene (**109a**, 0.4 mmol, 2 equiv.); The product was isolated as white solid (55 mg, 0.17 mmol, 56%) after electrolysis; $R_f = 0.28$ (pentane / acetone = 4:1); ¹H NMR (500 MHz, CD₂Cl₂) δ 8.10 – 8.03 (m, 2H), 7.63 – 7.56 (m, 2H), 7.44 – 7.27 (m, 4H), 6.89 – 6.75 (m, 2H), 4.01 – 3.93 (m, 2H), 2.73 – 2.61 (m, 3H), 2.20 – 1.86 (m, 3H), 1.41 – 1.32 (m, 3H) ppm; major rotamer ¹³C NMR (126 MHz, CD₂Cl₂) δ 171.5, 157.3, 138.5, 136.2, 134.4, 131.2, 129.0, 127.7, 127.3, 127.2, 127.0, 127.0, 125.5, 123.9, 114.8, 64.2, 23.7, 19.8, 15.1 ppm; **FT-IR**: \tilde{v} =

2978, 2384, 1990, 1596, 1421, 1390, 1241, 1107, 1044, 922 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{21}H_{22}O_2N = 320.1645$ found: 320.1647.

N-(4-methoxy-2-methylphenyl)-*N*-(4-methylnaphthalen-1-yl)acetamide (110c)

Prepared according to general procedure B using *N*-(4-methoxy-2-methylphenyl)acetamide (**108c**, 0.2 mmol, 1 equiv.) and 1-methylnaphthalene (**109a**, 0.4 mmol, 2 equiv.); The product was isolated as white solid (40.2 mg, 0.13 mmol, 63%) after electrolysis; $R_f = 0.23$ (pentane / acetone = 4:1); ¹**H NMR** (500 MHz, CD₂Cl₂) δ 8.17 – 7.97 (m, 2H), 7.69 – 7.54 (m, 2H), 7.37 – 7.20 (m, 2H), 7.15 – 6.95 (m, 1H), 6.83 (d, J = 2.9 Hz, 1H), 6.54 (dd, J = 8.8, 2.9 Hz, 1H), 3.78 and 3.73 (two s, total 3H), 2.71 and 2.66 (two s, total 3H), 2.44 – 1.90 ppm (m, 6H); ¹³**C NMR** (151 MHz, CD₂Cl₂) δ 171.3, 158.7, 137.0, 135.8, 134.3, 127.7, 127.4, 127.1, 126.9, 126.8, 126.4, 125.6, 125.4, 124.3, 124.1, 116.6, 112.2, 55.8, 31.2, 23.2, 19.8 ppm; **FT-IR**: $\tilde{v} = 3053$, 2836, 1977, 1666, 1596, 1389, 1294, 1180, 917 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₂₁H₂₂O₂N = 320.1645 found: 320.1647.

4-(4-methylnaphthalen-1-yl)-2H-benzo[b][1,4] oxazin-3(4H)-one (110d)

Prepared according to general procedure B using 2H-benzo[b][1,4]oxazin-3(4H)-one (**108d**, 0.2 mmol, 1 equiv.) and 1-methylnaphthalene (**109a**, 0.4 mmol, 2 equiv.); The product was isolated as yellow solid (43.9 mg, 0.15 mmol, 76%) after electrolysis; $R_f = 0.44$ (pentane / acetone = 4:1); ¹**H NMR** (500 MHz, CD₂Cl₂) δ 8.14 (d, J = 8.5 Hz, 1H), 7.69 – 7.56 (m, 2H), 7.55 – 7.45 (m, 2H), 7.40 (d, J = 7.4 Hz, 1H), 7.11 (dd, J = 8.1, 1.2 Hz, 1H), 7.02 – 6.95 (m, 1H), 6.85 – 6.63 (m, 1H), 6.25 (dd, J = 8.1, 1.3 Hz, 1H), 4.92 (d, J = 15.0 Hz, 1H), 4.86 (d, J = 15.0 Hz, 1H), 2.80 ppm (s,

3H); ¹³C **NMR** (126 MHz, CD₂Cl₂) δ 165.0, 145.3, 137.2, 134.3, 131.4, 131.2, 130.5, 127.7, 127.5, 127.1, 127.1, 125.7, 124.5, 123.2, 123.1, 117.4, 117.2, 68.9, 19.9 ppm; **FT-IR**: $\tilde{\mathbf{v}} = 2873$, 2349, 2114, 1996, 1686, 1592, 1459, 1394, 1377, 1265, 1221, 1193, 930 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₁₉H₁₆O₂N = 290.1176 found: 290.1178.

5-bromo-1-(4-methylnaphthalen-1-yl)indolin-2-one (110e)

Prepared according to general procedure B using 5-bromoindolin-2-one (**108e**, 0.2 mmol, 1 equiv.) and 1-methylnaphthalene (**109a**, 0.4 mmol, 2 equiv.); The product was isolated as pale yellow solid (38.7 mg, 0.11 mmol, 55%) after electrolysis; $R_f = 0.45$ (pentane / acetone = 4:1); ¹H NMR (500 MHz, CD₂Cl₂) δ 8.13 (d, J = 8.4 Hz, 1H), 7.64 – 7.57 (m, 2H), 7.52 – 7.46 (m, 3H), 7.39 (d, J = 7.4 Hz, 1H), 7.27 – 7.23 (m, 1H), 6.22 (d, J = 8.4 Hz, 1H), 3.87 (d, J = 22.5 Hz, 1H), 3.79 (d, J = 22.5 Hz, 1H), 2.78 ppm (s, 3H); ¹³C NMR (126 MHz, CD₂Cl₂) δ 174.7, 146.1, 137.2, 134.2, 131.2, 130.1, 129.6, 128.2, 127.2, 127.1, 127.1, 127.0, 126.6, 125.62, 123.5, 115.4, 111.5, 36.4, 19.9 ppm; FT-IR: $\tilde{v} = 2915$, 2848, 2385, 2115, 1994, 1721, 1462, 1350, 1239, 969 cm⁻¹; HR-MS: calc. for [M+H]⁺ C₁₉H₁₅ON⁷⁹Br = 352.0331 found: 352.0333 C₁₉H₁₅ON⁸¹Br = 354.0311 found: 354.0312.

5-chloro-1-(4-methylnaphthalen-1-yl)indolin-2-one (110f)

Prepared according to general procedure B using 5-chloroindolin-2-one (**108f**, 0.2 mmol, 1 equiv.) and 1-methylnaphthalene (**109a**, 0.4 mmol, 2 equiv.); The product was isolated as white solid (36.8

mg, 0.12 mmol, 60%) after electrolysis; $R_f = 0.31$ (pentane / acetone = 4:1); ¹H NMR (500 MHz, CDCl₃) δ 8.12 – 8.08 (m, 1H), 7.61 – 7.56 (m, 2H), 7.50 – 7.43 (m, 2H), 7.41 – 7.34 (m, 2H), 7.09 (dd, J = 8.3, 2.1 Hz, 1H), 6.27 (d, J = 8.4 Hz, 1H), 3.89 (d, J = 22.5 Hz, 1H), 3.82 (d, J = 22.5 Hz, 1H), 2.77 ppm (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 174.7, 145.0, 136.8, 133.9, 129.7, 129.0, 128.1, 128.1, 127.0, 126.7, 126.2, 125.9, 125.2, 125.1, 123.1, 110.9, 36.1, 19.7 ppm; FT-IR: $\tilde{v} = 2664$, 2116, 1719, 1512, 1481, 1356, 1270, 1195, 1110, 1023, 943 cm⁻¹; HR-MS: calc. for [M+H]⁺ $C_{19}H_{15}ON^{35}Cl = 308.0837$ found: 308.0836 $C_{19}H_{15}ON^{37}Cl = 310.0807$ found: 310.0807.

5-fluoro-1-(4-methylnaphthalen-1-yl)indolin-2-one (110g)

Prepared according to general procedure B using 5-fluoroindolin-2-one (**108g**, 0.2 mmol, 1 equiv.) and 1-methylnaphthalene (**109a**, 0.4 mmol, 2 equiv.); The product was isolated as yellow solid (15 mg, 0.05 mmol, 26%) after electrolysis; $R_f = 0.34$ (pentane / acetone = 4:1); ¹**H NMR** (500 MHz, CD_2Cl_2) δ 8.16 – 8.10 (m, 1H), 7.64 – 7.57 (m, 2H), 7.52 – 7.45 (m, 2H), 7.39 (d, J = 7.3 Hz, 1H), 7.17 – 7.11 (m, 1H), 6.87 – 6.79 (m, 1H), 6.26 (dd, J = 8.6, 4.4 Hz, 1H), 3.87 (d, J = 22.4 Hz, 1H), 3.79 (d, J = 22.6 Hz, 1H), 2.78 ppm (s, 3H); ¹³C NMR (126 MHz, CD_2Cl_2) δ 175.1, 159.7 (d, J = 23.4 Hz), 143.0, 137.1, 134.3, 130.1 (d, J = 34.9 Hz), 127.2, 127.0, 127.0, 126.6, 126.6, 126.5, 125.6, 123.6, 114.5 (d, J = 23.5 Hz), 112.9 (d, J = 25.0 Hz), 110.5 (d, J = 8.3 Hz). 36.9, 19.9 ppm; **FT-IR**: $\tilde{v} = 2915$, 2349, 1724, 1480, 1357, 1216, 1110, 1061, 983 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{19}H_{15}ONF = 292.1132$ found: 292.1134.

1-(3,4-dimethylnaphthalen-1-yl)indolin-2-one (110h)

Prepared according to general procedure B using indolin-2-one (**108h**, 0.2 mmol, 1 equiv.) and 1,2-dimethylnaphthalene (**109b**, 0.4 mmol, 2 equiv.); The product was isolated as yellow solid (24.1 mg, 0.08 mmol, 42%) after electrolysis; $R_f = 0.22$ (cyclohexane / ethyl acetate = 4:1); ¹**H NMR** (500 MHz, CDCl₃) δ 8.13 (d, J = 8.6 Hz, 1H), 7.59 (d, J = 8.5 Hz, 1H), 7.57 – 7.51 (m, 1H), 7.41 – 7.34 (m, 3H), 7.15 – 7.04 (m, 2H), 6.35 (d, J = 7.6 Hz, 1H), 3.90 (d, J = 22.4 Hz, 1H), 3.83 (d, J = 22.5 Hz, 1H), 2.67 (s, 3H), 2.53 ppm (s, 3H); ¹³**C NMR** (126 MHz, CDCl₃) δ 175.5, 146.6, 134.2, 133.7, 133.6, 129.4, 128.6, 128.5, 128.1, 126.6, 125.8, 124.7, 124.6, 124.4, 123.1, 122.8, 110.0, 36.3, 20.9, 14.9 ppm; **FT-IR**: \tilde{v} = 3667, 2976, 2398, 2115, 1994, 1718, 1512, 1486, 1385, 1299, 1192, 980 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₂₀H₁₈ON = 288.1383 found: 288.1384.

5-chloro-1-(3,4-dimethylnaphthalen-1-yl)indolin-2-one (110i)

Prepared according to general procedure B using 5-chloroindolin-2-one (**108f**, 0.2 mmol, 1 equiv.) and and 1,2-dimethylnaphthalene (**109b**, 0.4 mmol, 2 equiv.); The product was isolated as white solid (49 mg, 0.15 mmol, 76%) after electrolysis; $R_f = 0.28$ (cyclohexane / ethyl acetate = 4:1); ¹**H NMR** (500 MHz, CDCl₃) δ 8.13 (d, J = 8.6 Hz, 1H), 7.58 – 7.50 (m, 2H), 7.43 – 7.36 (m, 1H), 7.36 – 7.34 (m, 1H), 7.33 (s, 1H), 7.11 – 7.06 (m, 1H), 6.26 (d, J = 8.4 Hz, 1H), 3.89 (d, J = 22.5 Hz, 1H), 3.81 (d, J = 22.5 Hz, 1H), 2.67 (s, 3H), 2.53 ppm (s, 3H); ¹³**C NMR** (126 MHz, CDCl₃) δ 174.8, 145.1, 134.2, 133.9, 133.7, 129.4, 128.3, 128.2, 128.1, 128.1, 126.7, 125.9, 125.9, 125.0, 124.8, 122.9, 110.9, 36.2, 21.2, 15.0 ppm; **FT-IR**: $\tilde{v} = 2922$, 2546, 1977, 1609, 1512, 1386, 1286, 1194, 1069, 999 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₂₀H₁₇ON³⁵Cl = 322.0993 found = 322.0995, C₂₀H₁₇ON³⁷Cl = 324.0964 found = 324.0964.

N-(3,4-dimethylnaphthalen-1-yl)-*N*-(4-methoxyphenyl)acetamide (110j)

Prepared according to general procedure B using *N*-(4-methoxyphenyl)acetamide (**108a**, 0.2 mmol, 1 equiv.) and 1, 2-dimethylnaphthalene (**109b**, 0.4 mmol, 2 equiv.); The product was isolated as white solid (55 mg, 0.17 mmol, 86%) after electrolysis; $R_f = 0.16$ (cyclohexane / ethyl acetate = 4:1); major rotamer ¹**H NMR** (500 MHz, CDCl₃) δ 8.09 (d, J = 8.2 Hz, 1H), 8.02 (d, J = 8.1 Hz, 1H), 7.59 – 7.48 (m, 2H), 7.33 (d, J = 9.3 Hz, 3H), 6.81 (d, J = 9.0 Hz, 2H), 3.74 (s, 3H), 2.63 (s, 3H), 2.49 (s, 3H), 1.92 ppm (s, 3H); ¹³**C NMR** (126 MHz, CDCl₃) δ 171.6, 157.4, 137.5, 135.9, 134.2, 133.7, 132.7, 129.9, 129.6, 128.4, 126.6, 126.3, 124.6, 123.3, 114.1, 55.5, 23.6, 20.9, 14.9 ppm; **FT-IR**: \tilde{v} = 2931, 2835, 2089, 1667, 1598, 1506, 1461, 1367, 1293, 1121, 909 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₂₁H₂₂O₂N = 320.1645 found = 320.1647.

N-mesityl-*N*-(4-methoxyphenyl)acetamide (110k)

Prepared according to general procedure B using *N*-(4-methoxyphenyl)acetamide (**108a**, 0.2 mmol, 1 equiv.) and mesitylene (**109c**, 1 mmol, 5 equiv.); The product was isolated as yellow oil (27.1 mg, 0.09 mmol, 46%) after electrolysis; $R_f = 0.32$ (pentane / acetone = 4:1); major rotamer ¹H NMR (500 MHz, CD_2Cl_2) δ 7.20 – 7.15 (m, 2H), 6.98 (s, 2H), 6.81 – 6.76 (m, 2H), 3.75 (s, 3H), 2.31 (s, 3H), 2.15 (s, 6H), 1.83 (s, 3H); ¹³C NMR (126 MHz, CD_2Cl_2) δ 170.7, 157.0, 138.8, 136.7, 134.6, 130.4, 127.9, 125.5, 113.9, 55.9, 23.7, 21.3, 18.3 ppm; **FT-IR**: $\tilde{v} = 2915$, 2349, 1995, 1667, 1585, 1482, 1367, 1294, 1197, 1111, 1033 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{18}H_{22}O_2N = 284.1645$ found: 284.1647.

N-(4-isopropylphenyl)-*N*-mesitylacetamide (110l)

Prepared according to general procedure B using *N*-(4-isopropylphenyl)acetamide (**108i**, 0.2 mmol, 1 equiv.) and mesitylene (**109c**, 1 mmol, 5 equiv.); The product was isolated as yellow oil (27.1 mg, 0.09 mmol, 46%) after electrolysis; $R_f = 0.43$ (pentane / acetone = 4:1); ¹**H NMR** (500 MHz, CDCl₃) δ 7.21 – 7.14 (m, 2H), 7.13 – 7.04 (m, 2H), 6.97 (s, 2H), 2.84 (hept, J = 6.9 Hz, 1H), 2.32 (s, 3H), 2.17 (s, 6H), 1.89 (s, 3H), 1.20 ppm (d, J = 6.9 Hz, 6H); ¹³**C NMR** (126 MHz, CDCl₃) δ 170.7, 145.3, 138.5, 138.3, 138.3, 136.3, 130.0, 126.5, 123.4, 33.6, 24.1, 23.8, 21.2, 18.2 ppm; **FT-IR**: $\tilde{\mathbf{v}} = 2959$, 1672, 1509, 1481, 1366, 1296, 1192, 1056, 982 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $\mathbf{C}_{20}\mathbf{H}_{26}\mathbf{ON} = 296.2009$ found: 296.2011.

N-(2,4-dimethoxyphenyl)-*N*-mesitylacetamide (110m)

Prepared according to general procedure B using N-(2,4-dimethoxyphenyl)acetamide (**108j**, 0.2 mmol, 1 equiv.) and mesitylene (**109c**, 1 mmol, 5 equiv.) in 1:1 DCM:HFIP mixture (0.06 M, 3 mL); The product was isolated as yellow oil (28 mg, 0.09 mmol, 45%) after electrolysis; $R_f = 0.20$ (pentane / acetone = 4:1); 1:1 rotamers ¹H NMR (500 MHz, CDCl₃) δ 6.96 – 6.71 (m, 3H), 6.53 (dd, J = 4.0, 2.7 Hz, 1H), 6.39 – 6.31 (m, 1H), 3.89 and 3.88 (two s, total 3H), 3.78 and 3.75 (two s, 3H), 2.30 and 2.28 (two s, total 3H), 2.24, 2.21 and 2.12 (three s, total 6H), 1.97 and 1.91 ppm (two s, total 3H); ¹³C NMR (126 MHz, CDCl₃) δ 172.2, 170.4, 159.2, 159.1, 155.1, 154.8, 139.3, 139.2, 137.6, 136.8, 136.3, 136.0, 135.2, 130.0, 129.8, 129.3, 127.1, 126.4, 124.7, 123.6, 104.8, 104.6, 100.0, 99.4, 56.0, 55.6, 55.6, 55.5, 22.7, 21.6, 21.1, 19.0, 18.2, 17.9 ppm; **FT-IR**: \tilde{v} = 2921,

2841, 2160, 1668, 1586, 1452, 1294, 1160, 938, 877 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ $C_{19}H_{24}O_3N$ = 314.1751 found: 314.1752.

tert-butyl 2-(N-(4-methoxyphenyl)acetamido)-3-methyl-1H-indole-1-carboxylate (110n)

Prepared according to general procedure B using *N*-(4-methoxyphenyl)acetamide (**108a**, 0.2 mmol, 1 equiv.) and *tert*-butyl 3-methyl-1*H*-indole-1-carboxylate (**109d**, 0.4 mmol, 2 equiv.); Yellow solid; The product was isolated as yellow oil (36 mg, 0.09 mmol, 46%) after electrolysis; $R_f = 0.22$ (cyclohexane / ethyl acetatae = 4:1); ¹H NMR (500 MHz, CD₂Cl₂) δ 8.15 (d, J = 8.4 Hz, 1H), 7.54 (d, J = 7.7 Hz, 1H), 7.41 – 7.35 (m, 1H), 7.32 – 7.23 (m, 3H), 6.85 – 6.79 (m, 2H), 3.76 (s, 3H), 2.20 (s, 3H), 2.04 (s, 3H), 1.57 ppm (s, 9H); ¹³C NMR (126 MHz, CD₂Cl₂) δ 171.6, 157.8, 149.9, 135.0, 134.9, 133.6, 128.7, 126.4, 125.9, 123.4, 119.8, 116.1, 115.6, 114.1, 85.1, 55.9, 28.4, 23.5, 8.5 ppm; **FT-IR**: $\tilde{v} = 2930$, 2836, 2116, 1991, 1685, 1476, 1395, 1286, 936 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₂₃H₂₇O₄N₂ = 395.1965 found = 395.1964.

$$\begin{array}{c|c}
O & & \\
O & & \\
O & & \\
CF_3 & \\
CF_3 & \\
\end{array}$$

N-(10-((1,1,1,3,3,3-hexafluoropropan-2-yl)oxy)anthracen-9-yl)-N-(4-methoxyphenyl)acetamide (110o)

Prepared according to general procedure B using N-(4-methoxyphenyl)acetamide (**108a**, 0.2 mmol, 1 equiv.) and anthracene (**109e**, 0.4 mmol, 2 equiv.); The product was isolated as yellow oil (60.1 mg, 0.12 mmol, 60%) after electrolysis; $R_f = 0.38$ (pentane / acetone = 4:1); ¹H NMR

(400 MHz, CD₂Cl₂) δ 8.36 (dd, J = 7.5, 2.5 Hz, 2H), 8.22 – 8.13 (m, 2H), 7.69 – 7.59 (m, 4H), 7.29 (dd, 2H), 6.76 (dd, 2H), 5.40 (h, J = 5.5 Hz, 1H), 3.71 (s, 3H), 1.70 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 171.7, 157.1, 147.8, 135.4, 132.4, 130.0, 128.3, 127.0, 125.5, 124.1, 123.8, 123.4, 121.7, 121.4 (q, J = 284.7 Hz), 114.0, 55.5, 23.7 ppm; ¹⁹F NMR (565 MHz, CDCl₃) δ -71.42 ppm; **FT-IR**: $\tilde{\mathbf{v}}$ = 2937, 2844, 2349, 1991, 1671, 1509, 1439,1376, 1223, 1054, 905 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₂₆H₂₀O₃NF₆= 508.1342 found: 508.1335.

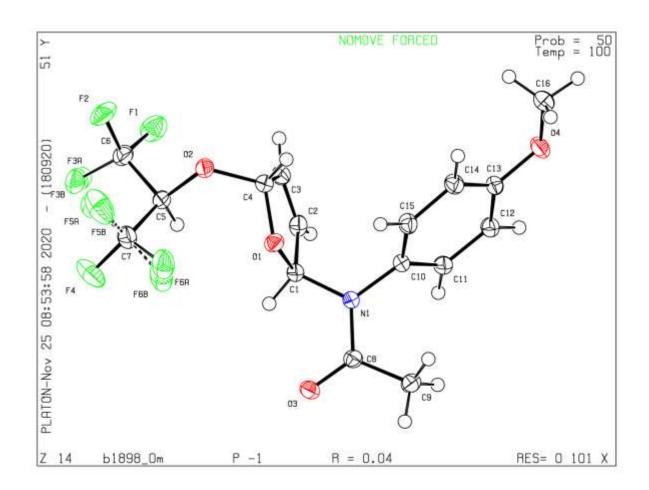
N-5-((1,1,1,3,3,3-hexafluoropropan-2-yl)oxy)-2,5-dihydrofuran-2-yl)-N-(4-methoxyphenyl)acetamide (110p)

Prepared according to general procedure B using *N*-(4-methoxyphenyl)acetamide (**108a**, 0.2 mmol, 1 equiv.) and furan (**109f**, 1 mmol, 5 equiv.); The product was isolated as pale yellow solid (72 mg, 0.18 mmol, 90%) after electrolysis; $R_f = 0.23$ (cyclohexane / ethyl acetatae = 4:1); ¹H NMR (500 MHz, CDCl₃) δ 7.47 – 7.42 (m, 1H), 7.14 (s, 1H), 6.91 (s, 1H), 6.86 (d, *J* = 7.8 Hz, 2H), 6.03 (d, *J* = 5.8 Hz, 1H), 5.82 – 5.75 (m, 2H), 4.50 (hept, *J* = 5.8 Hz, 1H), 3.81 (s, 3H), 1.83 ppm (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 172.1, 159.7, 133.8, 131.2, 130.5, 129.2, 121.7 (d, *J* = 284.2 Hz), 114.5, 107.7, 90.3, 70.9 (hept, *J* = 32.7 Hz), 55.5, 23.3 ppm (*see crystal data*); **FT-IR**: \tilde{v} = 3149, 2850, 2361, 2159, 1977, 1667, 1513, 1373, 1279, 1096, 963 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₁₆H₁₆O₄NF₆ = 400.0978 found = 400.0976.

N-(2-((1,1,1,3,3,3-hexafluoropropan-2-yl)oxy)-4-methoxyphenyl)acetamide (103')

Physical data of **103**': ¹**H NMR** (500 MHz, CDCl₃) δ 8.17 (d, J = 9.0 Hz, 1H), 7.34 (s, 1H), 6.69 (dd, J = 9.0, 2.7 Hz, 1H), 6.58 (d, J = 2.7 Hz, 1H), 4.84 (hept, J = 5.7 Hz, 1H), 3.79 (s, 3H), 2.18 ppm (s, 3H); ¹³**C NMR** (126 MHz, CDCl₃) δ 168.3, 156.7, 147.4, 123.4, 122.5, 121.0 (q, J = 284.8 Hz), 109.1, 102.7, 76.7 (hept, J = 33.6 Hz), 55.8, 24.5 ppm.

7.3.5 Crystal data



CCDC 2018010

Compound	110p
CCDC	2018010
Data block name	data_b1898_0m
Identification code	B1898_0m
Empirical formula	$C_{16}H_{15}F_6NO_4$
Formula weight	399.29
Temperature/K	100.0
Crystal system	triclinic
Space group	P-1
a/Å	7.3160(5)
b/Å	11.3203(9)
c/Å	11.6708(10)
α/°	112.918(3)
β/°	92.672(4)
γ/°	107.503(3)
Volume/Å ³	834.17(12)
Z	2
$ ho_{ m calc} g/cm^3$	1.590
μ/mm^{-1}	0.156
F(000)	408.0
Crystal size/mm ³	$0.749 \times 0.505 \times 0.144$
Radiation	$MoK\alpha (\lambda = 0.71073)$
2Θ range for data collection/°	4.314 to 61.056
Index ranges	$-10 \le h \le 10$, $-16 \le k \le 16$, $-16 \le l \le 16$
Reflections collected	56063
Independent reflections	5096 [$R_{int} = 0.0275$, $R_{sigma} = 0.0160$]
Data/restraints/parameters	5096/0/333
Goodness-of-fit on F ²	1.040
Final R indexes [I>= 2σ (I)]	$R_1 = 0.0359, wR_2 = 0.0938$
Final R indexes [all data]	$R_1 = 0.0402, wR_2 = 0.0977$
Largest diff. peak/hole / e Å ⁻³	0.48/-0.30

7.4 Experimental part for the synthesis of bioactive chromones

7.4.1 General procedures

Preparation of key building blocks as starting materials

HBr (2.2 equiv.)
OXONE® (1.2 equiv.)
$$Et_3N \text{ (6 equiv.)}$$

$$CH_2Cl_2 \text{ (0.25 M)}$$

3-bromo-4*H*-chromen-4-one was prepared according to literature known procedure by addition of hydrobromic acid to the mixture of chromone and *OXONE*[®] in CH₂Cl₂ followed by treatment of Et₃N in moderate to good yields. The analytical data was identical with the reported.^[130]

3-(4-(trifluoromethyl)phenyl)-4H-chromen-4-one was prepared according to literature known procedure following a palladium catalyzed cross-coupling reaction. The analytical data was identical with the reported. [126]

Procedure A for the coupling between chromone and azoles

$$CF_3$$
 CF_3 CF_3

Isoflavone (0.1 mmol) and azole (0.2 mmol) were dissolved in a 4 mL screw-capped vial with 0.5 mL of anhydrous DMF. Then molecular iodine (0.15 mmol), followed by anhydrous K₂CO₃ were added at room temperature. The reaction mixture was stirred at 80 °C for 12 h. The reaction mixture was quenched with a saturated solution of sodium thiosulphate and extracted with

dichloromethane, washed with water, brine and dried over anhydrous Na₂SO₄. The organic extracts were filtered and concentrated under reduced pressure. Purification by column-chromatography on silica gel afforded the pure product.

Procedure B for the coupling between chromone and azoles

Chromone (0.1 mmol) and triazole (0.11 mmol) were dissolved in a 4 mL screw-capped vial with 0:5 mL of anhydrous DMF. Then molecular iodine (0.15 mmol) was added at room temperature and the reaction mixture was stirred at 80 °C for 12 h. Azole (0.2-0.4 mmol) followed by anhydrous K₂CO₃ were added to the reaction mixture and the reaction was continued for 3-14 h. Then the reaction mixture was quenched with saturated solution of sodium thiosulphate and extracted with dichloromethane, washed with water, brine, dried over anhydrous Na₂SO₄. The organic extracts were filtered and concentrated under reduced pressure. Purification by column chromatography on silica gel afforded the pure product.

General procedure C for the nucleophilic substitution

1 equiv. of 2-(1H-1,2,4-triazol-1-yl)-4H-chromen-4-one was transferred to a vial followed by the addition of dry DMF (0.1 M). After that 2 equiv. of nucleophile was added. The mixture was heated at the mentioned temperature. The progress of the reaction was monitored by TLC. Upon completion, the reaction mixture was diluted with EtOAc (10 mL) and washed 3 times with H₂O (3*10 mL). The organic phase was dried by sodium sulfate and concentrated in vacuo. The pure product was obtained by column chromatography.

7.4.2 Physical data of products

2-(1*H*-1,2,4-triazol-1-yl)-3-(4-(trifluoromethyl)phenyl)-4*H*-chromen-4-one (116a)

Prepared according to general procedure A. The product was isolated as white solid (31.8 mg, 0.09 mmol, 89%); $R_f = 0.28$ (cyclohexane / ethyl acetate = 3:2); ${}^{1}H$ NMR (500 MHz, CD_2Cl_2) δ 8.29 (d, J = 8.0 Hz, 1H), 8.09 (d, J = 14.5 Hz, 2H), 7.81 (t, J = 7.7 Hz, 1H), 7.61 (dd, J = 13.0, 8.2 Hz, 3H), 7.54 (t, J = 7.6 Hz, 1H), 7.33 (d, J = 8.0 Hz, 2H) ppm; ${}^{13}C$ NMR (126 MHz, $CDCl_3$) δ 176.8, 154.4, 153.2, 149.7, 144.9, 135.2, 133.3, 130.9 (d, J = 32.7 Hz), 130.5, 126.7 (d, J = 8.8 Hz), 125.8 (q, J = 3.6 Hz), 125.0, 123.0, 122.8, 118.1, 116.6 ppm; **FT-IR**: $\tilde{v} = 3120$, 1643, 1613, 1579, 1503, 1468, 1422, 1381, 1330, 1291, 1273, 1241, 1124, 1106, 1071 cm $^{-1}$; **HR-MS**: calc. for [M+H] $^+$ $C_{18}H_{11}O_2N_3F_3$: 358.0798 found: 358.0799.

2-(5,6-dimethyl-1H-benzo[d]imidazol-1-yl)-3-(4-(trifluoromethyl)phenyl)-4H-chromen-4-one (116b)

Prepared according to general procedure A. The product was isolated as pale yellow solid (20 mg, 0.05 mmol, 46%); $R_f = 0.30$ (cyclohexane / ethyl acetate = 3:2); ${}^{1}H$ NMR (500 MHz, CD₂Cl₂) δ 8.30 (dd, J = 8.1, 1.7 Hz, 1H), 7.87 – 7.80 (m, 1H), 7.63 (dd, J = 8.6, 1.0 Hz, 1H), 7.60 – 7.54 (m, 3H), 7.51 (s, 2H), 7.40 – 7.33 (m, 3H), 2.37 ppm (s, 6H).; ${}^{13}C$ NMR (126 MHz, CD₂Cl₂) δ 177.1, 154.9, 151.9, 142.3, 141.2, 135.2, 135.0, 133.9, 131.2, 131.2, 130.6 (d, J = 32.3 Hz), 126.9, 126.8, 126.0 (q, J = 3.8 Hz), 125.6, 123.7, 123.5, 121.2, 118.4, 115.8, 112.7, 20.8, 20.5 ppm; **FT-IR** $\tilde{v} =$

3631, 2925, 2514, 2028, 1977, 1641, 1571, 1508, 1397, 1266, 1160, 960 cm⁻¹; **HR-MS**: calc. for $[M+H]^+$ $C_{25}H_{18}O_2N_2F_3 = 435.1315$ found: 435.1311.

2-(1*H*-pyrrol-1-yl)-3-(4-(trifluoromethyl)phenyl)-4*H*-chromen-4-one (116c)

Prepared according to general procedure B. The product was isolated as yellow solid (11 mg, 0.03 mmol, 31%); $R_f = 0.42$ (cyclohexane / ethyl acetate = 3:2); ¹H NMR (700 MHz,CD₂Cl₂) δ 8.29 – 8.20 (m, 1H), 7.80 (d, J = 7.6 Hz, 1H), 7.70 – 7.49 (m, 7H), 7.35 (d, J = 7.6 Hz, 2H), 6.37 ppm (d, J = 4.3 Hz, 1H).; ¹³C NMR (176 MHz, CD₂Cl₂) δ 177.5, 154.8, 153.0, 143.5, 135.8, 135.1, 131.8, 131.3, 130.2 (q, J = 32.5 Hz), 126.8, 126.6, 125.6 (q, J = 4.0 Hz), 123.6, 118.5, 114.7, 109.1 ppm; FT-IR: $\tilde{v} = 3676$, 2988, 2159, 1726, 1632, 1573, 1464, 1321, 1161, 1067, 908 cm⁻¹; HR-MS: calc. for [M+H]⁺ C₂₀H₁₃O₂NF₃ = 356.0893 found: 357.0847.

2-(1H-indazol-1-yl)-3-(4-(trifluoromethyl)phenyl)-4H-chromen-4-one (116d)

Prepared according to general procedure A. The product was isolated as yellow solid (12 mg, 0.03 mmol, 30%); $R_f = 0.53$ (cyclohexane / ethyl acetate = 3:2); ${}^{1}H$ NMR (500 MHz, CD_2Cl_2) δ 8.19 (dd, J = 7.9, 1.5 Hz, 1H), 7.78 (d, J = 8.1 Hz, 2H), 7.67 (m, 6H), 7.48 – 7.43 (m, 1H), 7.40 (d, J = 8.4 Hz, 1H), 7.32 – 7.26 (m, 1H), 6.96 ppm (s, 1H); ${}^{13}C$ NMR (126 MHz, CD_2Cl_2) δ 175.1, 157.1, 153.6, 140.0, 136.0, 134.5, 133.7, 133.6, 132.1, 131.7, 130.5 (d, J = 32.5 Hz), 126.7 (q, J = 3.8 Hz), 126.1, 125.7, 123.8, 123.6, 123.6, 117.4, 116.7, 106.5, 103.9 ppm; FT-IR: $\tilde{v} = 2923$, 2522, 2362, 2159, 1977, 1617, 1419, 1399, 1237, 1122, 1020 cm $^{-1}$; HR-MS: calc. for [M+H] $^+$ $C_{23}H_{14}O_2N_2F_3 = 407.1002$ found: 407.1000.

2-(1*H*-benzo[d]imidazol-1-yl)-3-(4-(trifluoromethyl)phenyl)-4*H*-chromen-4-one (116e)

Prepared according to general procedure A. The product was isolated as pale-yellow solid (8 mg, 0.02 mmol, 20%); $R_f = 0.30$ (cyclohexane / ethyl acetate = 3:2); ${}^{1}H$ NMR (500 MHz, CD_2Cl_2) δ 8.30 (dd, J = 7.9, 1.7 Hz, 1H), 7.86 - 7.82 (m, 1H), 7.79 - 7.76 (m, 1H), 7.65 - 7.54 (m, 6H), 7.41 – 7.37 ppm (m, 4H); ${}^{13}C$ NMR (126 MHz, CD_2Cl_2) δ 177.1, 154.9, 151.5, 143.8, 142.1, 135.3, 134.8, 132.8, 131.2, 130.7 (q, J = 32.5 Hz), 127.2 (d, J = 18.9 Hz), 126.9, 126.9, 126.1 (q, J = 3.8 Hz), 125.5, 124.7, 123.6, 121.3, 118.4, 116.2, 112.6 ppm; **FT-IR**: $\tilde{v} = 2672$, 2384, 2115, 1991, 1624, 1503, 1353, 1292, 1162, 960 cm $^{-1}$; **HR-MS**: calc. for [M+H] $^+$ $C_{23}H_{14}O_2N_2F_3 = 407.1002$ found: 407.0999.

2-(4-methylpiperazin-1-yl)-3-(4-(trifluoromethyl)phenyl)-4*H*-chromen-4-one (116f)

Prepared according to general procedure C. The product was isolated as pale yellow solid (7 mg, 0.02 mmol, 18%); $R_f = 0.50$ (dichloromethane / methanol = 9:1); ${}^{1}\mathbf{H}$ NMR (700 MHz, CDCl3) δ 8.19 (dd, J = 7.8, 1.7 Hz, 1H), 7.66 (d, J = 8.1 Hz, 2H), 7.62 – 7.59 (m, 1H), 7.56 (d, J = 8.0 Hz, 2H), 7.39 – 7.33 (m, 2H), 3.28 (t, J = 5.0 Hz, 4H), 2.36 (t, J = 5.0 Hz, 4H), 2.27 ppm (s, 3H); ${}^{13}\mathbf{C}$ NMR (176 MHz, CDCl₃) δ 176.1, 161.6, 153.4, 138.1, 132.8, 131.1, 129.0 (q, J = 32.3 Hz), 126.4, 125.3 (q, J = 3.7 Hz), 125.3, 123.6, 123.1, 116.6, 103.5, 54.5, 47.7, 46.1 ppm ; **FT-IR**: $\tilde{\mathbf{v}} = 2925$, 2029, 1615, 1548, 1464, 1293, 1162, 1019, 879 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₂₁H₂₀O₂N₂F₃ = 389.1471 found: 389.1473.

2-morpholino-3-(4-(trifluoromethyl)phenyl)-4*H*-chromen-4-one (116g)

Prepared according to general procedure C. The product was isolated as pale yellow solid (5 mg, 0.01 mmol, 24%); $R_f = 0.35$ (cyclohexane / ethyl acetate = 3:2); ¹H NMR (500 MHz, CDCl₃) δ 8.20 (dd, J = 7.9, 1.6 Hz, 1H), 7.67 (d, J = 8.1 Hz, 2H), 7.65 – 7.58 (m, 1H), 7.58 (d, J = 8.0 Hz, 2H), 7.40 – 7.34 (m, 2H), 3.66 – 3.60 (m, 4H), 3.28 – 3.22 ppm (m, 4H); ¹³C NMR (126 MHz, CDCl₃) δ 176.1, 161.4, 153.4, 137.8, 132.9, 131.1, 129.2 (d, J = 32.6 Hz), 126.8, 126.5, 125.5 (q, J = 3.9 Hz), 125.2, 123.0, 116.6, 103.7, 66.4, 48.1 ppm; **FT-IR**: $\tilde{v} = 2921$, 2854, 2090, 1614, 1515, 1406, 1320, 1161, 999, 898 cm⁻¹; **HR-MS**: calc. for [M+H]⁺ C₂₀H₁₇O₃NF₃ = 376.1155 found: 376.1156.

2-(benzylamino)-3-(4-(trifluoromethyl)phenyl)-4H-chromen-4-one (116h)

Prepared according to general procedure C. The product was isolated as brown solid (6 mg, 0.02 mmol, 27%); $R_f = 0.45$ (cyclohexane / ethyl acetate = 3:2); ${}^{1}H$ NMR (700 MHz,CDCl₃) δ 8.21 (dd, J = 7.8, 1.7 Hz, 1H), 7.69 (d, J = 8.0 Hz, 2H), 7.62 – 7.57 (m, 1H), 7.54 (d, J = 8.0 Hz, 2H), 7.39 – 7.35 (m, 4H), 7.34 – 7.29 (m, 3H), 5.31 (t, J = 6.1 Hz, 1H), 4.64 ppm (d, J = 6.1 Hz, 2H); ${}^{13}C$ NMR (176 MHz, CDCl₃) δ 173.8, 160.6, 153.0, 137.5, 136.6, 132.3, 131.6, 129.8 (q, J = 32.5 Hz), 129.2, 128.7, 128.2, 127.4, 126.4, 126.3 (q, J = 3.7 Hz), 125.2, 123.4, 116.5, 99.9, 45.8 ppm; FT-IR: $\tilde{v} = 3741$, 3032, 2921, 2850, 2115, 1736, 1698, 1581, 1484, 1319, 1166, 909 cm⁻¹; HR-MS: calc. for [M+H] $^+$ C₁₆H₁₄N = C₂₃H₁₇O₂NF₃ = 396.1206 found: 396.1200.

2-(piperidin-1-yl)-3-(4-(trifluoromethyl)phenyl)-4*H*-chromen-4-one (116i)

Prepared according to general procedure C. The product was isolated as pale yellow solid (7 mg, 0.02 mmol, 19%); $R_f = 0.54$ (cyclohexane / ethyl acetate = 3:2); ¹H NMR (700 MHz, CDCl₃) δ 8.19 (dd, J = 7.6, 1.9 Hz, 1H), 7.65 (d, J = 7.8 Hz, 2H), 7.60 – 7.55 (m, 3H), 7.37 – 7.33 (m, 2H), 3.22 – 3.19 (m, 4H), 1.62 – 1.56 (m, 2H), 1.52 (p, J = 5.7 Hz, 4H). ¹³C NMR (176 MHz, CDCl₃) δ 176.0, 162.1, 153.4, 138.7, 132.6, 130.9, 128.6 (q, J = 32.4 Hz), 126.4, 125.2 (q, J = 3.8 Hz), 124.9, 123.7, 123.2, 116.5, 103.0, 49.3, 25.7, 24.2 ppm; FT-IR: $\tilde{v} = 2928$, 2841, 1732, 1614, 1542, 1425, 1250, 1106, 1068, 955, 893 cm⁻¹; HR-MS: calc. for [M+H]⁺ C₂₁H₁₉O₂NF₃= 374.1362 found: 374.1363.

2-(pyrrolidin-1-yl)-3-(4-(trifluoromethyl)phenyl)-4H-chromen-4-one (116j)

Prepared according to general procedure C. The product was isolated as pale yellow solid (10 mg, 0.03 mmol, 28%); $R_f = 0.43$ (cyclohexane / ethyl acetate = 3:2); ${}^{1}H$ NMR (700 MHz, CDCl₃) δ 8.19 (dd, J = 7.8, 1.7 Hz, 1H), 7.64 – 7.60 (m, 2H), 7.58 – 7.53 (m, 1H), 7.51 – 7.45 (m, 2H), 7.35 – 7.31 (m, 2H), 3.26 – 3.21 (m, 4H), 1.95 – 1.60 ppm (m, 4H); ${}^{13}C$ NMR (176 MHz, CDCl₃) δ 175.4, 159.4, 153.4, 138.6, 132.4, 132.2, 128.7 (q, J = 32.3 Hz), 126.3, 125.3, 124.6 (q, J = 3.7 Hz), 123.7, 122.9, 116.2, 100.7, 50.1, 25.5 ppm; **FT-IR**: $\tilde{v} = 2665$, 2384, 2118, 1996, 1727, 1614, 1512, 1468, 1357, 1222, 1098, 954, 883 cm⁻¹; **HR-MS**: calc. for [M+H] $^{+}$ C₂₀H₁₇O₂NF₃ = 360.1206 found: 360.1207.

2-(piperazin-1-yl)-3-(4-(trifluoromethyl)phenyl)-4*H*-chromen-4-one (116k)

Prepared according to general procedure C. The product was isolated as pale yellow solid (12 mg, 0.03 mmol, 32%); $R_f = 0.44$ (dichloromethane / methanol = 9:1); ${}^{1}H$ NMR (700 MHz, CDCl₃) δ 8.20 (dd, J = 7.8, 1.7 Hz, 1H), 7.66 (d, J = 8.1 Hz, 2H), 7.63 – 7.58 (m, 1H), 7.57 (d, J = 8.0 Hz, 2H), 7.39 – 7.33 (m, 2H), 3.64 (s, 1H), 3.27 – 3.23 (m, 4H), 2.85 – 2.81 ppm (m, 4H); ${}^{13}C$ NMR (176 MHz, CDCl₃) δ 176.1, 161.7, 153.4, 138.1, 132.8, 131.1, 128.9 (q, J = 32.4 Hz), 126.4, 125.3 (q, J = 3.7 Hz)., 125.1, 123.6, 123.1, 116.6, 103.5, 48.9, 45.7 ppm; **FT-IR**: $\tilde{v} = 2954$, 2854, 2359, 2159, 1976, 1727, 1677, 1597, 1466, 1425, 1342, 1276, 1068, 1020, 942 cm⁻¹; **HR-MS**: calc. for $[M+H]^+$ $C_{20}H_{18}O_2N_2F_3 = 375.1315$ found: 375.1315.

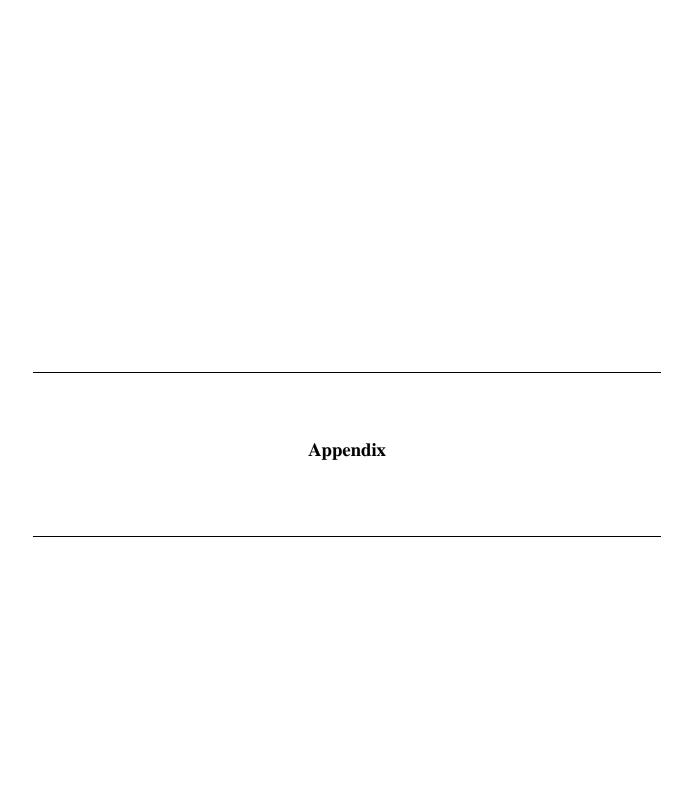
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9 Appendix

9.1 List of Abbreviations

Å Ångström

Ac acetyl

anhyd. anhydrous

aq. aqueous
Ar argon

BDD boron doped diamond

BDE bond dissociation energy

BMP bone morphogenetic protein

Bn benzyl

Boc *tert*-butyloxycarbonyl

Bu butyl

Bz benzoyl

calc. calculated

cat. catalyst

CDC cross dehydrogenative coupling

CE counter electrode

COMAS Compound Management and Screening Center

conc. concentration

const. constant

conv. conversion

CV cyclic voltammetry

DCE 1,2-dichloroethane

DCM dichloromethane

DDQ 2,3-dichloro-5,6-dicyano-1,4-benzoquinone

DIPEA diisopropylethylamine

DMF dimethylformamide

DMSO dimethylsulfoxide

EC₅₀ half maximal effective concentration

List of Abbreviations i

EDG electron-donating group
ESI electronspray ionization

Et ethyl

et al. et alii (and others)

 Et_3N triethylamine Et_2O diethylether EtOAc ethyl acetate equiv. equivalent

EWG electron withdrawing group

F Faraday

FT-IR Fourier-Transform Infrared spectroscopy

G graphite

GC glassy carbon

h hour

HFIP 1,1,1,3,3,3-hexafluoro-2-propanol

Hh hedgehog

HPLC high-performance liquid chromatography

HR-MS high resolution mass spectrometry

Hz Hertz
I current

IC₅₀ half-maximal inhibitory concentration

*i*Pr iso-propyl

J coupling constants

K₂CO₃ potassium carbonate

m- metaMe methyl

MeCN acetonitrile

mESCs Mouse embryonic stem cells

NaOEt sodium ethoxide

n.d. not detected

NMR nuclear magnetic resonance

List of Abbreviations ii

Nu nucleophile

o- ortho OAc acetate p- para Ph phenyl

PIDA iodosobenzene diacetate

PIFA iodosobenzene bis(trifluoroacetate)

PPh₃ triphenylphosphine ppm parts per million rt room temperature

p-TsOH *para* -toluenesulfonic acid

Q charge passed R_f retention factor

RVC reticulated vitreous catbon

SAR structure-activity relationship
SCE saturated calomel electrode

SET single electron transfer

sat. saturated

TBAF tetrabutylammonium fluoride

tBu tert-butyl

temp. temperature

THF tetrahydrofuran
TM transition metal

t_R retention time

Ts para-tosyl

TsCl para-toluenesulfonyl chloride

TS transitionstate

V voltage

WE working electrode

List of Abbreviations iii

9.2 Acknowledgement

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1. <u>Puthanveedu, M.</u>; Khamraev, V.; Brieger, L.; Strohmann, C.; Antonchick, A. P., Electrochemical Dehydrogenative C(sp²)-H Amination. *Chem. Eur. J.* 2021, 27, 8008.

2. Wesseler, F.; Riege, D.; <u>Puthanveedu, M.</u>; Halver, J.; Müller, E.; Bertrand, J.; Mercola, M.; Antonchick, A. P.; Ziegler, S.; Sievers, S.; Waldmann, H.; Schade, D., Probing embryonic mesodermal differentiation enables identification of small molecule bone morphogenetic protein activators (*manuscript submitted*).

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- 3. <u>Puthanveedu, M.</u>; Antonchick A. P. Aromatic C–H Functionalization with Iodine(I/III) Catalysts or Iodine (III) Reagents in *Iodine Catalysis in Organic Synthesis*, co-edited by Kazuaki Ishihara and Kilian Muñiz. Wiley-VCH. 2020 (*book chapter in press*).
- 4. <u>Puthanveedu, M.</u>; Polychronidou, V.; Antonchick, A. P., Catalytic Selective Metal-Free Cross-Coupling of Heteroaromatic *N*-Oxides with Organosilanes. *Org. Lett.* 2019, *21*, 3407-3411.
- 5. Mishra, A.; Vats, T. K.; <u>Puthanveedu, M.</u>; Das, A.; Deb, I., Rhodium-Catalyzed sp² C–H Acetoxylation of *N*-Aryl Azaindoles/*N*-Heteroaryl Indolines. *J. Org. Chem.* 2017, 82, 12406–12415.

Conferences

- 04/2021 Mahesh Puthanveedu, Vladislav Khamraev, Lukas Brieger, Carsten Strohmann, Andrey P. Antonchick "Electrochemical Dehydrogenative C(sp²)-H Amination" 8th IMPRS-LM student symposium, Dortmund, Germany, April 2021 (poster presentation)
- 07/2019 Mahesh Puthanveedu, Vasiliki Polychronidou, Andrey P. Antonchick "Catalytic Metal-Free Cross-Coupling of Heteroaromatic *N*-Oxides with Organosilanes" European Symposium on Organic Chemistry, Vienna, Austria, July 2019 (poster presentation)
- Mahesh Puthanveedu, Aniket Mishra, Tripta Kumari Vats, Indu Bhusan Deb "Rhodium-Catalyzed sp² C–H Acetoxylation of *N*-Aryl Azaindoles" International Symposium on Chemical Biology and Drug Discovery, Kolkata, India, March 2016 (poster presentation)

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9.4 Eidesstattliche Versicherung (Affidavit)

Eidesstattliche Versicherung (Affidavit)

Puthanveedu, Mahesh	215726			
Name, Vorname (Sumame, first name)	Matrikel-Nr. (Enrolment number)			
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Affidavit ix