

Guest editorial:

HUMAN NON-PARENCHYMAL LIVER CELLS FOR CO-CULTIVATION SYSTEMS

Ahmed Ghallab

Department of Forensic and Toxicology, Faculty of Veterinary Medicine,
South Valley University, Qena, Egypt; ghallab@vet.svu.edu.eg

Recently, Pfeiffer and colleagues have published protocols that allow the isolation of human hepatocytes and non-parenchymal liver cells from the same donor (Pfeiffer et al., 2014). Cell isolation is performed with resected liver tissue, usually obtained from hepatectomy because of metastasis from colon cancer. Human liver cells are initially isolated by the conventional two-step EDTA/collagenase perfusion technique. Next, hepatocytes and non-parenchymal cells are separated by low-speed centrifugation. After purification by Percoll density gradient centrifugation, Kupffer cells, sinusoidal endothelial cells and stellate cells are further separated by specific adhesion and by magnetic bead sorting. Typical yields of a single isolation are 1.9×10^6 Kupffer cells, 2.7×10^5 sinusoidal endothelial cells and 4.7×10^5 stellate cells. All cell types can be cultivated either as mono- or co-cultures.

Co-cultivation of non-parenchymal cells and hepatocytes from the same donor may become an important approach of hepatotoxicity testing in future. It is well known that non-parenchymal cells play a critical role in hepatotoxicity (Laskin, 1996; Kantari-Mimoun et al., 2014; Krell et al., 1987). Sinusoidal endothelial cells have been shown to support liver regeneration, where they serve as 'guide-rails' for regenerating hepatocytes and guarantee the rapid re-establishment of functional liver tissue (Hoehme et al., 2010; Schliess et al., 2014). Moreover, sinusoidal endothelial cells have been shown to secrete HGF and Wnt factors during liver regeneration, which stimulate hepatocytes to prolifer-

ate (Ding et al., 2010). They condition the vascular niche by angiocrine signals which can be disturbed after repeated administration of hepatotoxic compounds, leading to activation of stellate cells and fibrosis (Ding et al., 2014). Currently, *in vitro* systems for hepatotoxicity testing are in the focus of toxicological research (Ghallab, 2013; Krell et al., 1987; Schyschka et al., 2013; Grinberg et al., 2014). Although the important role of non-parenchymal cells is out of question (Liu et al., 2013; Hammad et al., 2014; McCuskey et al., 2005; Yee et al., 2003), currently mostly hepatocyte monocultures are tested (Miszczuk et al., 2014; Rodrigues et al., 2013; Watzek et al., 2013; Vinken et al., 2013; Huang et al., 2013). An explanation probably is that cultivation of non-parenchymal cells is still challenging. For example stellate cells tend to be activated spontaneously in culture and sinusoidal endothelial cells may lose capacity to secrete cytokines. Although there is still a long way to go, the technique of Pfeiffer et al. (2014) will improve the availability of human non-parenchymal cells and may facilitate the development of *in vitro* systems that recapitulate the communication between hepatocytes and non-parenchymal cells.

REFERENCES

Ding BS, Nolan DJ, Butler JM, James D, Babazadeh AO, Rosenwaks Z, et al. Inductive angiocrine signals from sinusoidal endothelium are required for liver regeneration. *Nature* 2010;468(7321):310-5.

- Ding BS, Cao Z, Lis R, Nolan DJ, Guo P, Simons M, et al. Divergent angiocrine signals from vascular niche balance liver regeneration and fibrosis. *Nature* 2014; 505(7481):97-102.
- Ghallab A. In vitro test systems and their limitations. *EXCLI J* 2013;12:1024-6.
- Grinberg M, Stöber RM, Edlund K, Rempel E, Godoy P, Reif R, et al. Toxicogenomics directory of chemically exposed human hepatocytes. *Arch Toxicol*. 2014;88:2261-87.
- Hammad S, Hoehme S, Friebel A, von Recklinghausen I, Othman A, Begher-Tibbe B, et al. Protocols for staining of bile canalicular and sinusoidal networks of human, mouse and pig livers, three-dimensional reconstruction and quantification of tissue microarchitecture by image processing and analysis. *Arch Toxicol*. 2014;88:1161-83.
- Hoehme S, Brulport M, Bauer A, Bedawy E, Schormann W, Hermes M, et al. Prediction and validation of cell alignment along microvessels as order principle to restore tissue architecture in liver regeneration. *Proc Natl Acad Sci USA*. 2010;107:10371-6.
- Huang CS, Lii CK, Lin AH, Yeh YW, Yao HT, Li CC, et al. Protection by chrysin, apigenin, and luteolin against oxidative stress is mediated by the Nrf2-dependent up-regulation of heme oxygenase 1 and glutamate cysteine ligase in rat primary hepatocytes. *Arch Toxicol*. 2013;87:167-78.
- Kantari-Mimoun C, Castells M, Klose R, Meinecke AK, Lemberger UJ, Rautou PE, et al. Resolution of liver fibrosis requires myeloid cell-driven sinusoidal angiogenesis. *Hepatology*. 2014 Dec 5. doi: 10.1002/hep.27635. [Epub ahead of print].
- Krell H, Metz J, Jaeschke H, Höke H, Pfaff E. Drug-induced intrahepatic cholestasis: characterization of different pathomechanisms. *Arch Toxicol*. 1987;60:124-30.
- Laskin DL. Sinusoidal lining cells and hepatotoxicity. *Toxicol Pathol*. 1996;24:112-8.
- Liu Y, Gardner CR, Laskin JD, Laskin DL. Classical and alternative activation of rat hepatic sinusoidal endothelial cells by inflammatory stimuli. *Exp Mol Pathol*. 2013;94:160-7.
- McCuskey RS, Bethea NW, Wong J, McCuskey MK, Abril ER, Wang X, et al. Ethanol binging exacerbates sinusoidal endothelial and parenchymal injury elicited by acetaminophen. *J Hepatol*. 2005;42:371-7.
- Miszcuk GS, Barosso IR, Zucchetti AE, Boaglio AC, Pellegrino JM, Sánchez Pozzi EJ, et al. Sandwich-cultured rat hepatocytes as an in vitro model to study canalicular transport alterations in cholestasis. *Arch Toxicol*. 2014 Jun 10. [Epub ahead of print].
- Pfeiffer E, Kegel V, Zeilinger K, Hengstler JG, Nüssler AK, Seehofer D, et al. Isolation, characterization, and cultivation of human hepatocytes and non-parenchymal liver cells. *Exp Biol Med (Maywood)*. 2014 Nov 12. [Epub ahead of print].
- Rodrigues AV, Rollison HE, Martin S, Sarda S, Schulz-Utermoehl T, Stahl S, et al. In vitro exploration of potential mechanisms of toxicity of the human hepatotoxic drug fenclozic acid. *Arch Toxicol*. 2013;87:1569-79.
- Schliess F, Hoehme S, Henkel SG, Ghallab A, Driesch D, Böttger J, et al. Integrated metabolic spatial-temporal model for the prediction of ammonia detoxification during liver damage and regeneration. *Hepatology*. 2014;60:2040-51.
- Schyschka L, Sánchez JJ, Wang Z, Burkhardt B, Müller-Vieira U, Zeilinger K, et al. Hepatic 3D cultures but not 2D cultures preserve specific transporter activity for acetaminophen-induced hepatotoxicity. *Arch Toxicol*. 2013;87:1581-93.
- Vinken M, Maes M, Cavill R, Valkenburg D, Ellis JK, Decroock E, et al. Proteomic and metabolomic responses to connexin43 silencing in primary hepatocyte cultures. *Arch Toxicol*. 2013;87:883-94.
- Watzek N, Scherbl D, Schug M, Hengstler JG, Baum M, Habermeyer M, et al. Toxicokinetics of acrylamide in primary rat hepatocytes: coupling to glutathione is faster than conversion to glycidamide. *Arch Toxicol*. 2013;87:1545-56.
- Yee SB, Hanumegowda UM, Copple BL, Shibuya M, Ganey PE, Roth RA. Endothelial cell injury and coagulation system activation during synergistic hepatotoxicity from monocrotaline and bacterial lipopolysaccharide coexposure. *Toxicol Sci*. 2003;74:203-14.