Guest editorial

PERSPECTIVES OF TISSUES IN SILICO

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Over the past decade much effort has been invested into the development of in vitro systems as alternatives to animal experiments (Hammad et al., 2013, 2014a, b; Hammad, 2013; Godoy et al., 2010, 2013; Hewitt et al., 2007; Stewart and Marchan, 2012; Gebel et al., 2014; Grinberg et al., 2014). However, in vitro systems still have the limitation that they often do not sufficiently represent the in vivo situation. Moreover, quantitative in vitro to in vivo extrapolation is difficult (Ghallab, 2013; Reif, 2014; Stewart, 2010).

In recent years a concept is emerging that may overcome many of the current limitations of in vitro testing, namely in silico tissues (Hoehme et al., 2010; Schliess et al., 2014). Typically, virtual tissues are based on reconstructions of real tissues, where the exact positions of each individual cell and further relevant structures, e.g. blood vessels, are known in a three-dimensional space (Hoehme et al., 2010; Höhme et al., 2007). In the first step spatio-temporal models are generated from reconstructions (Hammad et al., 2014c). For this purpose the individual cell serves as the smallest unit. Model parameters, such as the probability to divide or to die, and even more complex properties, such as migration rules can be programmed into each cell. This results in a model that can simulate, for example, the spatiotemporal process of tissue damage and regeneration. Key principles how cells in the liver coordinately respond to large destructions to restore functional tissue have been identified by such models (Drasdo et al., 2014a; Hoehme et al., 2010). In next steps, further processes can be integrated into spatio-temporal models, e.g. blood flow or metabolic processes. As an example, Schliess et al. (2014) have integrated metabolic pathway models of ammonia detoxification into spatio-temporal models. This allows simulating ammonia concentrations in the blood circulation and how they are influenced by specific damage patterns of the liver.

In toxicology, modelling especially structure activity and physiologically-basedpharmacokinetic (PBPK) models have a long standing tradition (Schug et al., 2013; Karamanakos et al., 2009; Carlsson et al., 2004; Thiel et al., 2015; Hammad and Ahmed, 2014; Dobrev et al., 2001; El-Masri et al., 1996). However, the advent of spatiotemporal models with the possibility to integrate other model types opens new possibilities. Integrated mathematical models formalize the relationship between individual components to test their interactions in a virtual setting (Drasdo et al., 2014a, b; Widera, 2014). It can be expected that virtual tissue approaches will have a strong impact to understand complex pathophysiologies, especially when processes and interactions have to be elucidated that cannot be directly measured by established methods.

REFERENCES

Carlsson C, Harju M, Bahrami F, Cantillana T, Tysklind M, Brandt I. Olfactory mucosal toxicity screening and multivariate QSAR modeling for chlorinated benzene derivatives. Arch Toxicol. 2004; 78:706-15.

Dobrev ID, Andersen ME, Yang RS. Assessing interaction thresholds for trichloroethylene in combination with tetrachloroethylene and 1,1,1-trichloroethane using gas uptake studies and PBPK modeling. Arch Toxicol. 2001;75:134-44.

Drasdo D, Bode J, Dahmen U, Dirsch O, Dooley S, Gebhardt R, et al. The virtual liver: state of the art and future perspectives. Arch Toxicol. 2014a;88:2071-5.

Drasdo D, Hoehme S, Hengstler JG. How predictive quantitative modelling of tissue organisation can inform liver disease pathogenesis. J Hepatol. 2014b; 61:951-6.

El-Masri HA, Thomas RS, Sabados GR, Phillips JK, Constan AA, Benjamin SA, et al. Physiologically based pharmacokinetic/pharmacodynamic modeling of the toxicologic interaction between carbon tetrachloride and Kepone. Arch Toxicol. 1996;70:704-13.

Gebel T, Foth H, Damm G, Freyberger A, Kramer PJ, Lilienblum W, et al. Manufactured nanomaterials: categorization and approaches to hazard assessment. Arch Toxicol. 2014;88:2191-211.

Ghallab A. In vitro test systems and their limitations. EXCLI J. 2013;12:1024-6.

Godoy P, Lakkapamu S, Schug M, Bauer A, Stewart JD, Bedawi E, et al. Dexamethasone-dependent versus-independent markers of epithelial to mesenchymal transition in primary hepatocytes. Biol Chem. 2010; 391:73–83.

Godoy P, Hewitt NJ, Albrecht U, Andersen ME, Ansari N, Bhattacharya S, et al. Recent advances in 2D and 3D in vitro systems using primary hepatocytes, alternative hepatocyte sources and non-parenchymal liver cells and their use in investigating mechanisms of hepatotoxicity, cell signaling and ADME. Arch Toxicol. 2013;87:1315–530.

Grinberg M, Stöber RM, Edlund K, Rempel E, Godoy P, Reif R, et al. Toxicogenomics directory of chemically exposed human hepatocytes. Arch Toxicol. 2014;88:2261-87.

Hammad S. Advances in 2D and 3D in vitro systems for hepatotoxicity testing. EXCLI J. 2013;12:993-6.

Hammad S, Ahmed H. Biomarker: the universe of chemically induced gene expression alterations in human hepatocyte. EXCLI J. 2014;13:1275-7.

Hammad S, Marchan R, Hengstler JG. Cutting-edge topics in research on animal sciences. J Exp Appl Animal Sci. 2013;1:1-3.

Hammad S, Abdallah MF, Abdou AM. Current difficulties to replace animal experiments in toxicology. J Exp Appl Animal Sci. 2014a;1:269-70.

Hammad S, Omar MA, Abdel-Wareth AAA, Ahmed H. Pitfalls of in vitro systems: why we still need animal experiments? J Exp Appl Animal Sci. 2014b;1: 271-2.

Hammad S, Hoehme S, Friebel A, von Recklinghausen I, Othman A, Begher-Tibbe B, et al. Protocols for staining of bile canalicular and sinusoidal networks of human, mouse and pig livers, three-dimensional reconstruction and quantification of tissue microarchitecture by image processing and analysis. Arch Toxicol. 2014c;88:1161-83.

Hewitt NJ, Lechón MJ, Houston JB, Hallifax D, Brown HS, Maurel P, et al. Primary hepatocytes: current understanding of the regulation of metabolic enzymes and transporter proteins, and pharmaceutical practice for the use of hepatocytes in metabolism, enzyme induction, transporter, clearance, and hepatotoxicity studies. Drug Metab Rev. 2007;39:159-234.

Höhme S, Hengstler JG, Brulport M, Schäfer M, Bauer A, Gebhardt R, et al. Mathematical modelling of liver regeneration after intoxication with CCl(4). Chem Biol Interac. 2007;168:74–93.

Hoehme S, Brulport M, Bauer A, Bedawy E, Schormann W, Hermes M, et al. Prediction and validation of cell alignment along microvessels as order principle to restore tissue architecture in liver regeneration. Proc Natl Acad Sci USA 2010;107:10371–6.

Karamanakos PN, Trafalis DT, Geromichalos GD, Pappas P, Harkitis P, Konstandi M, et al. Inhibition of rat hepatic CYP2E1 by quinacrine: molecular modeling investigation and effects on 4-(methyl nitrosamino)-1-(3-pyridyl)-1-butanone (NNK)-induced mutagenicity. Arch Toxicol. 2009;83:571-80.

Reif R. Concepts of predictive toxicology. EXCLI J. 2014;13:1292-4.

Schliess F, Hoehme S, Henkel SG, Ghallab A, Driesch D, Böttger J, et al. Integrated metabolic spatial-temporal model for the prediction of ammonia detoxification during liver damage and regeneration. Hepatology. 2014;60:2040-51. Schug M, Stöber R, Heise T, Mielke H, Gundert-Remy U, Godoy P, et al. Pharmacokinetics explain in vivo/in vitro discrepancies of carcinogen-induced gene expression alterations in rat liver and cultivated hepatocytes. Arch Toxicol. 2013;87:337-45.

Stewart JD. In vitro test systems in toxicology. EXCLI J. 2010;9:156-8.

Stewart JD, Marchan R. Current developments in toxicology. EXCLI J. 2012;11:692-702.

Thiel C, Schneckener S, Krauss M, Ghallab A, Hofmann U, Kanacher T, et al. A systematic evaluation of the use of physiologically based pharmacokinetic modeling for cross-species extrapolation. J Pharm Sci. 2015;104:191-206.

Widera A. Integrated spatiotemporal-metabolic modelling bridges the gap between metabolism on the cellular level and organ function. EXCLI J. 2014; 13:1289-91.